Distribution Agreement

In presenting this thesis or dissertation as a partial fulfillment of the requirements for an advanced degree from Emory University, I hereby grant to Emory University and its agents the non-exclusive license to archive, make accessible, and display my thesis or dissertation in whole or in part in all forms of media, now or hereafter known, including display on the world wide web. I understand that I may select some access restrictions as part of the online submission of this thesis or dissertation. I retain all ownership rights to the copyright of the thesis or dissertation. I also retain the right to use in future works (such as articles or books) all or part of this thesis or dissertation.

-DocuSigned by: Daniel Gromer Signature: -E9C9FC461A4F4CE...

Daniel Gromer Name <u>3/27/2024 | 9:03</u> PM EDT Date TitleThe Relationship Between the Immunogenicity and Reactogenicity of a SeasonalInfluenza Vaccine Delivered by Microneedle Patch or Hypodermic Needle

Author Daniel Gromer

Degree Master of Science

Program Clinical Research

Approved by the Committee

-DocuSigned by:

— DocuSigned by:

DocuSigned by:

DocuSigned by: Daniel Graciaa 4F20987BEE764A2...

Jordan Kempker *Advisor*

Amita Manatunga Advisor

Nadine Rouphael *Committee Member*

Daniel Graciaa Committee Member

Committee Member

Committee Member

Accepted by the Laney Graduate School:

Kimberly Jacob Arriola, Ph.D, MPH Dean, James T. Laney Graduate School

Date

The Relationship Between the Immunogenicity and Reactogenicity of a Seasonal Influenza Vaccine Delivered by Microneedle Patch or Hypodermic Needle

By

Daniel Gromer

B.A., University of Pennsylvania, 2012

M.D., University of Pennsylvania, 2018

Advisor: Nadine Rouphael, M.D., M.Sc.

An abstract of

A thesis submitted to the Faculty of the

James T. Laney School of Graduate Studies of Emory University

in partial fulfillment of the requirements for the degree of

Master of Science

in Clinical Research

2024

Abstract

The Relationship Between the Immunogenicity and Reactogenicity of a Seasonal Influenza Vaccine Delivered by Microneedle Patch or Hypodermic Needle

By Daniel Gromer

<u>Background</u>: Vaccine immunogenicity and reactogenicity each depend on recipient and vaccine characteristics. An association between these two vaccine outcomes would have major implications for clinical care, public health, and vaccine development. We hypothesized that healthy adults who reported higher reactogenicity from seasonal inactivated influenza vaccine (IIV) developed higher antibody titers compared with those who reported lower reactogenicity.

<u>Methods</u>: We performed a secondary analysis of the TIV-MNP 2015 study, a randomized phase 1 clinical trial comparing the immunogenicity and reactogenicity of a trivalent IIV delivered by microneedle patch (MNP) or intramuscular injection (IM). We created composite scores of solicited adverse events (Global, Systemic, and Local) as the exposure and hemagglutination inhibition (HAI) antibody titers against the H1N1, H3N2, and B antigens in the vaccine as the outcome. To account for longitudinal outcome data, we used mixed model analysis of variance to estimate geometric mean titers (GMTs), GMT ratios (GMRs), and titer fold change ratios (FCRs) and modified Poisson generalized estimating equations to estimate risk ratios (RRs) of HAI seroprotection and seroconversion. We adjusted for several confounders and generated estimates separately by vaccine delivery method.

<u>Results</u>: The IM (n=25) and MNP (n=50) groups were balanced in baseline characteristics. Longitudinal estimates of H3N2 HAI GMTs were associated with the Systemic and Local scores among the IM group. Within the IM group, those with high reaction scores had lower baseline H3N2 HAI GMTs (Global GMR 0.5, p=0.06; Systemic GMR 0.4, p=0.01; Local GMR 0.3, p=0.01) and twice the titer fold change by day 28 (Global FCR 2.0, p=0.04; Systemic FCR 1.9, p=0.07; Local FCR 2.0, p=0.15) compared with those with low reaction scores. Those with high Local scores had a higher risk of HAI seroconversion (RR 1.4, p=0.03).

<u>Conclusion</u>: These results suggest that heightened reactogenicity to intramuscular inactivated influenza vaccine is related to low baseline humoral immunity to an included antigen. Participants with greater reactogenicity developed greater antibody titer fold change after 4 weeks, though the overall magnitude of response was similar or lower compared with low-reactogenicity participants.

The Relationship Between the Immunogenicity and Reactogenicity of a Seasonal Influenza Vaccine Delivered by Microneedle Patch or Hypodermic Needle

By

Daniel Gromer

B.A., University of Pennsylvania, 2012M.D., University of Pennsylvania, 2018

Advisor: Nadine Rouphael, M.D., M.Sc.

A thesis submitted to the Faculty of the

James T. Laney School of Graduate Studies of Emory University

in partial fulfillment of the requirements for the degree of

Master of Science

in Clinical Research

2024

Table of Contents

Introduction	1
Methods	8
Results	14
Discussion	19
Tables	27
Table 1. Baseline Participant Characteristics	27
Table 2. Longitudinal Associations Between Reaction Score and HAI GMT	28
Table 3A-C. Longitudinal Associations Between Alternative Cut Point Reaction Scores and HAI GM	
Table 4A-C. Reaction Score and HAI GMT by Study Day	32
Table 5. IM Study Group Reaction Score and H3N2 HAI GMT by Study Day	35
Table 6A-C. Reaction Score and HAI Titer Fold Change by Study Day	36
Table 7. Extreme Reaction Scores and HAI GMT by Study Day	39
Table 8. Extreme Reaction Scores and HAI Titer Fold Change by Study Day	40
Table 9. Reaction Severity and Longitudinal HAI GMT	41
Table 10. Reaction Severity and HAI Titer Fold Change by Study Day	42
Table 11. Reaction Duration and Longitudinal HAI GMT	43
Table 12. Reaction Duration and HAI Titer Fold Change by Study Day	44
Table 13. Longitudinal Associations Between Reaction Score and HAI Seroprotection	45
Table 14. HAI Seroprotection by Vaccine Route and Day	46
Table 15A-C. Longitudinal Associations Between Alternative Cut Point Reaction Scores and HAI Seroprotection	47
Table 16. Longitudinal Associations Between Reaction Score and HAI Seroconversion	50
Table 17A-C. Reaction Score and HAI Seroconversion by Study Day	51
Table 18A-C. Longitudinal Associations Between Alternative Cut Point Reaction Scores and HAI Seroconversion	54
Table 19A-C. Alternative Cut Point Reaction Scores and HAI Seroconversion by Study Day	57
Figures	63
Figure 1. Conceptual Model	63
Figure 2. Parent Study CONSORT Diagram	64
Figure 3. Adverse Events by Vaccine Route	65
Figure 4. Select Local Adverse Events by Time and Vaccine Route	66

References		
Figure 8. H3	N2 HAI Titer by Time and Reaction Score	
Figure 7. Re	action Score Bivariable Distributions	69
Figure 6. Re	action Score Univariable Distributions	
Figure 5. Co	rrelation Between Adverse Events	

Introduction

Vaccines are an effective public health measure for preventing and mitigating infectious diseases. A vaccine's immunity-boosting capacity, termed immunogenicity, and near-term adverse event profile, termed reactogenicity, depend on recipient characteristics, such as age,^{1–4} assigned sex,^{1,5,6} and potentially body mass index (BMI),^{1,7–12} as well as prior immunization^{13,14} and the platform, route, and dose of vaccine.^{2,4,15–18} Limited information exists regarding whether vaccine reactogenicity and immunogenicity are related to each other.

There are several reasons to pursue this question. One reason is to determine whether postvaccine symptoms could serve as a clinical indicator of population immunity in the early stages of a pandemic emergency. Knowing that a specific reaction or cadre of reactions implies that a vaccine has been effective in generating adequate immunity could allow public health officials to allocate a scarce vaccine better among a large group of people. More likely, learning about the balance between immunogenicity and reactogenicity will positively impact patient-provider communication by helping clinicians communicate with their patients. Over recent decades, numerous studies have linked low vaccine confidence to patient and parent fears about vaccine adverse events and higher vaccine confidence to the presence of a strong patient-provider relationship and trust in the medical and public health establishments.^{19–26} Defining the association between immunogenicity and reactogenicity may help clinicians to counsel patients about the meaning behind post-vaccine adverse events and build the trust needed to inspire vaccine confidence. Beyond affecting how clinicians communicate about what adverse events mean, investigating the immunogenicity-reactogenicity relationship can also provide them and their patients with guidance on how to manage adverse events. When discussing post-vaccine symptoms, a frequent and logical question is whether taking antipyretic analgesics to suppress vaccine reactions affects vaccine-derived immunity. Because fever after vaccination may herald increased immunity,²⁷ the concern is that taking antipyretics around the time of vaccination may blunt a necessary inflammatory response and dampen immunogenicity. Some evidence in children suggests that antipyretics taken at the time of vaccination may negatively impact immunogenicity,²⁸ but minimal investigation has been done in adults, who can provide more detailed reactogenicity data. Determining whether immunogenicity and reactogenicity are linked in analyses of adult participant data can serve as an appropriate starting point for future prospective studies of vaccines and medications such as acetaminophen and nonsteroidal anti-inflammatory drugs (NSAIDs). Ultimately, this line of study can enable medical providers and public health institutions to accurately coach people about the risks and benefits of taking medications to make tolerating vaccines easier.

From a biomedical research perspective, understanding the immunogenicity-reactogenicity relationship also has important implications for the future of vaccine design. Because vaccine and recipient characteristics impact vaccine responses, increasing the granularity of our knowledge about how variables such as sex, hormonal milieu, vaccine dose, and inclusion of adjuvants alter these responses may bring a new age of personalized vaccinology.

Though questions about the association between immunogenicity and reactogenicity had been asked infrequently prior to 2020, the pace of the implementation of SARS-CoV-2 vaccines to address the COVID-19 pandemic reinvigorated inquiry about the immunogenicity-reactogenicity relationship in the scientific literature and the lay press.^{29–31} Several publications featuring large samples, advanced biostatistical methods, and different SARS-CoV-2 vaccines subsequently found positive associations between reactogenicity and immunogenicity, particularly after multiple doses of mRNA vaccines or after participants with prior infection were exposed to vaccine antigen.^{32–39} Specific post-vaccination reactions (e.g., fever) and composite scores of reactions were both associated with increased immunity.^{34–37,40–48} While the estimation of this relationship for SARS-CoV-2 vaccines represents an exciting step, it is unclear if vaccines against other pathogens, such as influenza, carry the same immunogenicity-reactogenicity association.

Strains of influenza virus mutate perpetually, contributing to their ongoing evasion of human immunity through antigenic drift.^{49–51} As a result, the seasonal inactivated influenza vaccine (IIV) is manufactured yearly, anticipating antigenic drift, and is universally recommended in the United States (US) for individuals aged 6 months and older.^{4,52–54} To address the variety of distinct influenza viruses that circulate in each season, the seasonal IIV delivers multiple antigens designed to stimulate short-lived immunity against predicted strains. During the 2010s, seasonal IIVs approved in the US were either trivalent or quadrivalent, containing 1 influenza A/H3N2 antigen, and 1 or 2 influenza B antigens.

The antigens contained within seasonal IIVs and the strains they are intended to generate immunity against differ regarding their effects on and importance to human health. An A/H1N1 influenza strain caused the first influenza pandemic of the 21st century in 2009.^{55–57} Between 2010 and 2015, the seasonal IIV included a stable H1N1 antigen similar to the 2009 pandemic strain.^{58,59} As of the 2014–2015 influenza season, A/H1N1 influenza made up a minority of circulating virus and contributed to a minority of severe cases of disease.^{58,59}

In contrast, influenza A/H3N2 is usually dominant or co-dominant as the circulating virus and leads to most adult hospitalizations and deaths.^{59–61} The H3N2 antigen included in the seasonal IIV changes on a yearly basis and mismatches can occur between the circulating virus and the predicted antigen contained within the vaccine. This was the case in the 2014–2015 influenza season, when the vaccine's H3N2 antigen did not closely match the dominant circulating H3N2 virus.^{59,60}

Influenza B presents a different prediction problem than influenza A/H3N2. Since the 1980s, there have been 2 distinct co-circulating lineages of influenza B, which undergoes antigenic drift more slowly than influenza A.^{62,63} Efforts to predict which of the 2 lineages will dominate in a specific influenza season have largely failed, with trivalent IIVs in the US containing the dominant antigen less than 50% of the time between 2000 and early 2011.⁶³ Though influenza B causes fewer hospitalizations and deaths in adults, it is particularly notable for causing disease in children.⁶⁴ To mitigate the negative impact of frequent vaccine-strain mismatches, many trivalent influenza vaccines (TIVs) were transitioned to quadrivalent influenza vaccines (QIVs) by including 2 influenza B antigens during the 2010s and early 2020s.^{52,53,63,65,66}

One major challenge in estimating the immunogenicity-reactogenicity relationship of influenza vaccines comes in the form of confounding by human variation. Even in a clinical trial without variation in the platform, route, and dose of the vaccine of interest, heterogeneity in recipient characteristics may impact the estimation of the effect of reactogenicity on immunogenicity. Among multiple SARS-CoV-2 vaccines, younger age, female sex, and prior COVID-19 have all been associated with increases in both immunogenicity and reactogenicity, while the effect of BMI has yet to be shown consistently.^{33–35,37–39,67,68} Age^{1–4} and assigned sex^{1,5,6} affect seasonal IIV responses similarly, though prior antigen exposure via vaccination may be associated with decreased immunogenicity due to imprinting.^{13,14} The available data are inconsistent on the topic of the effect of BMI on influenza vaccine responses, perhaps in part due to differences in how BMI is categorized and expressed in various modeling approaches.^{1,7–12} Though differential vaccine effects are widely recognized and have been for decades, the implications of this heterogeneity in outcomes remains understudied and the approach to vaccination of large populations often follows a one-size-fits-all model. Seasonal influenza is currently the only pathogen for which we have dedicated vaccines for older (≥ 65) and younger (< 65) adult populations, ^{53,69–71} and there is currently no tailored vaccine approach to address differences in sex, BMI, vaccination history, or any other variable. Thus, models investigating the association between reactogenicity and immunogenicity must adjust for multiple confounding variables to achieve accurate estimation.

Heterogeneity in the platform, route, and dose of influenza vaccines represents a second major challenge in estimating the immunogenicity-reactogenicity relationship. The number and dose of

antigen, as well as the presence of an adjuvant, vary among licensed seasonal IIVs, precluding simple comparisons between trial participants who have received vaccines from different manufacturers. One way to circumvent this latter issue may be to analyze trials of the same vaccine delivered across distinct routes.

The seasonal IIV is generally offered, as with most vaccines worldwide, as an intramuscular (IM) injection via hypodermic needle.^{16,72} While manufacturing, regulatory, and distribution systems have matured around the constraints of this convention, global dominance of this method of vaccine delivery presents several challenges. IM vaccines require an intact cold chain, large amounts of storage space, and trained healthcare workers to administer them. Needle use results in frequent adverse safety events and needle phobia.⁷³ Non-single-use vials of vaccine result in wasted doses. Besides these significant logistical and acceptability issues, IM vaccines also suffer from immunologic drawbacks.^{15,74,75} The intramuscular compartment contains low numbers of antigen-presenting cells (APCs) as compared to mucosal sites and the skin. Exposing these alternative sites to vaccine antigens has repeatedly resulted in improved immunity against numerous infections.^{16,74,76}

As a result, there is growing enthusiasm for novel methods of achieving mucosal or intradermal vaccination. One such method is the microneedle patch (MNP), an array of sub-millimeter needles attached to a patch backing that delivers antigens to the intradermal space.^{15,16} Along with the logistical benefits that MNPs promise, such as low-cost manufacturing, thermostability, easy transportation, lack of waste, increased safety, and possibly the convenience of self-administration,^{54,77–80} several preclinical and clinical studies of MNP vaccines have shown equal

or enhanced immunogenicity over traditional IM vaccination.^{74,81} MNPs have also demonstrated a distinct adverse event profile from their IM counterparts, causing a higher frequency of redness, swelling, and itching where the patches were applied.^{54,82–84}

Therefore, to assess the relationship between reactogenicity and immunogenicity with a seasonal influenza vaccine, we performed a secondary analysis of TIV-MNP 2015, the completed first-inhuman phase 1 clinical trial of a seasonal IIV delivered by MNP, which was compared with IM injection of the same IIV.⁵⁴ Our aim was to estimate the effect of reactogenicity on immunogenicity for each delivery method, adjusting for measured confounding variables. We hypothesized that healthy, non-pregnant adults who reported higher reactogenicity from IIV developed markers of increased immunity compared with those who reported lower reactogenicity.

Methods

Parent Study Design

We used primary data obtained in the TIV-MNP 2015 trial,⁵⁴ a partly blinded, randomized, placebo-controlled, phase 1 study, described in brief below.

Setting and Participants

The TIV-MNP 2015 study included 100 healthy, non-pregnant, immunocompetent adults aged 18–49 years, who were recruited by the Hope Clinic of the Emory Vaccine Center in Atlanta, Georgia in the summer of 2015. Key exclusion criteria included influenza infection or vaccination during the 2014–2015 season, $BMI > 35 \text{ kg/m}^2$, recent blood donation, vaccination, or experimental product receipt, various acute and chronic medical or psychiatric conditions, and receipt of specified immunosuppressive or immunomodulatory medications. Please see clinicaltrials.gov, NCT02438423, for further details.

Randomization and Procedures

Participants were randomized in a 1:1:1:1 fashion to receive either IIV via microneedle patch (MNP_{IIV-HCW}), IIV via intramuscular injection (IM_{IIV}), or placebo via microneedle patch (MNP_{placebo}), all administered by a healthcare worker, or IIV via microneedle patch selfadministered by the study participant (MNP_{IIV-self}) under healthcare worker supervision. The IIV was composed of 15µg of each of the following influenza vaccine strains: A/Christchurch/16/2010, NIB-74 (H1N1), A/Texas/50/2012, NYMC X-223 (H3N2), B/Massachusetts/2/2012, NYMC BX-51 (B). Participants were followed for 180 days. Solicited local and systemic adverse events were assessed daily for 8 days after study product administration, by questionnaire initially and clinic visit if necessary. Adverse events were graded by severity (0 representing no event and 4 representing life-threatening event) based on the Food and Drug Administration toxicity grading schema.⁸⁵ Blood samples were drawn for immunogenicity testing on days 0, 28, and 180. We excluded all participants in the MNP_{placebo} group, as the association of interest was specific to those receiving IIV. Two additional participants did not provide blood samples, and therefore did not have any immunogenicity measurements available. We treated these values as missing at random and excluded the outcome data from statistical comparisons. There was no loss to follow up.

Variables

Reactogenicity

We chose measurements of reactogenicity derived from solicited adverse events as the exposure variables. Reactogenicity was divided into local and systemic adverse events. Local adverse events included swelling (induration), pain, redness (erythema), itching (pruritus), and tenderness. Systemic adverse events included fatigue, joint pain (arthralgia), body ache (myalgia), fever, shivering or shaking body movements, malaise, nausea, sweating, and headache.

Primary Measures of Reactogenicity

The number of unique solicited adverse events recorded by each participant at any point during the 8-day reporting period was summed to make continuous variables. We generated separate sums for local events, systemic events, and all events, and termed these Local, Systemic, and Global reaction scores. We examined the distributions of these scores using descriptive statistics and univariable and bivariable plots, visually determined cut points to separate high and low levels of each score, and generated dichotomous categorical variables as our primary reactogenicity measures.

Secondary Measures of Reactogenicity

We grouped participants by the severity of adverse events. Any participant recording an event of grade 2 or greater at any point during the study was included in the high severity group, and all others were included in the low severity group.

We also grouped participants by the duration of adverse events. Any participant with an event beginning on day 0 or day 1 and lasting greater than 2 continuous days (i.e., still present on day 2 or day 3, respectively), was included in the prolonged duration group, and all others were included in the short duration group.

Immunogenicity

We chose measurements of immunogenicity as the outcome variables. Immunogenicity was measured by hemagglutination inhibition (HAI) antibody titer. HAI measurement was performed by blinded Hope Clinic Laboratory staff using previously described methods.⁸⁶

Primary Measure of Immunogenicity

We chose HAI antibody titers as the primary measure of immunogenicity. HAI titers were inverted and represented as continuous numerical data. They were log-transformed for regression analyses and then back-transformed to generate geometric mean titers (GMTs) and the ratios between GMTs (geometric mean titer ratios, or GMRs).

Secondary Measures of Immunogenicity

We modeled HAI titer fold change as a measure of immunogenicity. We calculated HAI titer fold change by subtracting baseline log-transformed HAI from post-vaccine (day 28 or day 180) log-transformed HAI for each participant.

We also chose HAI seroprotection and HAI seroconversion as dichotomous categorical measures of immunogenicity. Seroprotection is defined as an HAI titer \geq 1:40. Seroconversion is defined as a post-vaccination measurement with a minimum 4-fold increase in HAI titer (if the baseline titer is \geq 1:10) or an HAI titer \geq 1:40 (if the baseline titer is < 1:10).

Covariates

Other measured covariates, recorded at enrollment, included BMI, sex, race, ethnicity, and IIV receipt in the prior two influenza seasons. All covariates were recorded by participant questionnaire except for BMI, which was measured with standard equipment in the clinic. All were represented as categorical variables. BMI was categorized as ≤ 25 , 25 to < 30, and ≥ 30 .

Sensitivity Analysis

To assess the robustness of findings generated with our primary measures of reactogenicity, we repeated our analyses using multiple different cut points for dichotomizing reaction scores. Additionally, we subsetted the participants using the 15 highest and 15 lowest values of each reaction score and performed comparisons between these groups of participants with extreme values of each reaction score. For example, we compared those with the 15 highest Global scores to those with the 15 lowest Global scores.

Statistical Analysis

We constructed a directed acyclic graph (DAG, Figure 1) to express our conceptual model of the effect of reactogenicity on immunogenicity and determine which variables to adjust for in multivariable modeling. To further characterize potential confounders and effect modifiers, we performed bivariable comparisons between selected variables and both the exposure and outcome measures. Covariates with statistically significant associations with both the exposure and outcome at the 95% confidence level were marked as potential confounders for adjustment in the multivariable analyses. Additionally, we used multiplicative terms to assess for interactions to include in multivariable models.

For the primary analyses, we used mixed model analysis of variance (ANOVA) to estimate GMTs and GMRs, using the exposure variable as the between-participant factor, time as the within-participant factor, and a compound symmetric covariance structure. We included vaccine delivery method (IM or MNP) in interaction terms with these factors, including a three-way interaction term, to generate separate estimates for participants receiving IM or MNP IIV on each study day. We used analogous methods for models estimating HAI titer fold change, which were expressed as ratios between groups (fold change ratios, or FCRs). For analyses with HAI seroprotection and seroconversion outcomes, we used modified Poisson regression^{87,88} and generalized estimating equations (GEEs)^{89,90} to generate risk ratios (RRs).

All statistical testing was performed in SAS version 9.4 (Cary, NC, USA). We used the MIXED procedure to generate estimates with mixed model ANOVAs and the GENMOD procedure to generate estimates with modified Poisson regression and GEEs to account for repeated outcome measures. Data visualization was performed with R v4.2.1 (R Foundation for Statistical Computing, Vienna, Austria).

Results

The initial enrolled sample included 100 participants. The original trial flow diagram is displayed in Figure 2. After the 25 participants in the MNP_{placebo} group were excluded, we excluded the outcome data of 2 additional participants who did not submit blood samples for analysis. We combined the MNP_{IIV-HCW} and MNP_{IIV-self} groups into a single MNP group to compare with the IM group. The two groups were balanced in baseline characteristics (Table 1). The median H1N1 HAI antibody titer was 80 and the median H3N2 and B HAI titers were 40.

Figure 3 displays the fraction of participants who received either IM or MNP IIV and had each type of reaction. The IM group reported more systemic events of every type except fever, which was only reported by a single participant, who was in the MNP group. The MNP group reported more local pruritus, redness, and swelling events, and the IM group reported more local pain events. Redness and swelling peaked within 24 hours of vaccination, resolving within 48 hours for the IM group and over 1–2 weeks for the MNP group (Figure 4).

Figure 5 displays the correlation matrix between all types of adverse events. Most Pearson correlation coefficients were close to 0 and there was no discernible hierarchical clustering pattern between adverse events.

Based on the distributions of the Global, Systemic, and Local reaction scores generated for all participants, we visually dichotomized the scores into high and low levels (Figures 6 and 7).

Those with a Global reaction score ≥ 4 , a Systemic reaction score ≥ 3 , or a Local reaction score ≥ 3 were assigned the high level for the corresponding score.

To screen for significant associations between reactogenicity and immunogenicity, we generated unadjusted and adjusted longitudinal GMR estimates and hypothesis tests for each vaccine antigen comparing participants in the IM or MNP groups with high and low Global, Systemic, and Local reaction scores (Table 2). We found no significant associations using the Global score but noted a consistent association among the IM group between antibody titers against the H3N2 antigen and both the Systemic and Local scores. We also noted associations between the Local score and the H1N1 and H3N2 HAI titers in the MNP group. These associations were not robust to changes in the dichotomization cut points of the reaction scores (Table 3A-C). In this sensitivity analysis, we also noted associations among the IM group between the B antigen and both the Global and Systemic scores with the lowest cut points, but these were not statistically significant after adjustment (Table 3A-B).

When we used interaction terms to generate time-specific estimates (Table 4A-C), we found that the previously noted associations among the MNP group were not driven by any significant difference on any specific day. In contrast, the IM group associations were driven by baseline differences in H3N2 HAI titer (reproduced from Table 4B in Table 5). Among participants in the IM group, adjusting for BMI, sex, race, ethnicity, and prior IIV, those with a high Systemic reaction score had 0.4 (95% CI 0.2, 0.8) times the day 0 H3N2 HAI GMT compared with those who had a low Systemic score. Similarly, those with a high Local score had 0.3 (95% CI 0.1, 0.8) times the day 0 H3N2 HAI GMT compared with those in the IM group as sociation score. Those in

the IM group with a high Systemic score developed similar H3N2 GMTs on day 28 compared with those who had a low Systemic score. In contrast, those with a high Local score appeared to develop lower H3N2 GMTs on day 28 compared with those who had a low Local score, though this did not reach statistical significance. Finally, those with high Systemic and Local scores appeared to demonstrate lower H3N2 GMTs on day 180 compared with their low-score counterparts, though these findings also did not reach statistical significance (Table 5, Figure 8).

We generated analogous unadjusted and adjusted estimates for the secondary outcome of HAI titer fold change (Table 6A-C). Among the IM group, prior to adjustment for confounding variables, those with any type of high reaction score had more than 2 times the day 28 H3N2 HAI titer fold change compared with their low-score counterparts. This finding remained similar after adjustment and was statistically significant when using the Global reaction score (Table 6B). Though it was not indicated by the longitudinal GMR estimates, we found a similar pattern among the MNP group using the Global reaction score and the B antigen, where a high score was associated with lower day 0 HAI titer (Table 4C) and greater day 28 HAI titer fold change (Table 6C).

As a sensitivity analysis, we selected participants with the 15 highest and 15 lowest values of each reaction score and performed unadjusted comparisons between these smaller groups of participants with extreme values. Among the IM group, those with the highest Local scores had lower H3N2 HAI GMTs compared to those with the lowest Local scores (Table 7), and those with the highest Systemic scores had a higher H3N2 HAI titer fold change on day 28 compared to those with the lowest Systemic scores (Table 8). Additionally, those in the IM group with the

highest Local reaction scores had higher H1N1 GMTs and a lower H1N1 HAI titer fold change on day 28 compared to those with the lowest Local scores, but this association was not found in the primary analyses.

We then grouped participants by either reaction severity or reaction duration, agnostic of reaction type, and estimated the relationship between these secondary reactogenicity measures and either HAI GMT or HAI titer fold change. We found no association between moderate or greater reaction severity and either longitudinal HAI GMT (Table 9) or HAI titer fold change (Table 10). We also found no association between a "prolonged" reaction duration greater than 48 continuous hours and either longitudinal HAI GMT (Table 11) or HAI titer fold change (Table 12).

Using modified Poisson GEEs, we then generated unadjusted and adjusted longitudinal estimates for the secondary outcome of HAI seroprotection (Table 13). There were no significant associations between any reaction score and HAI seroprotection, and models generating timespecific estimates frequently failed to converge (data not shown) due to the high proportion of participants with HAI seroprotection, including at baseline (Table 14). There were also no significant associations between any reaction score and HAI seroprotection after changing the dichotomization cut points of the scores (Table 15A-C).

Similarly, we generated unadjusted and adjusted longitudinal estimates for the secondary outcome of HAI seroconversion (Table 16), noting only an association between the Systemic reaction score and H3N2 HAI seroconversion in the MNP group after adjustment for confounding variables. When we generated adjusted time-specific estimates (Table 17A-C), we found that those with a high Local score had 1.4 (95% CI 1.0, 1.9) times the risk of day 28 H3N2 HAI seroconversion compared to those with a low Local score among the IM group (Table 17B). Those with a high Systemic score had 2.9 (95% CI 1.4, 6.0) times the risk of day 180 H3N2 HAI seroconversion compared to those with a low Systemic score. These findings were not robust to changes in the dichotomization cut points of the reaction scores (Table 18A-C, Table 19A-C).

Discussion

In this secondary analysis of a phase 1 clinical trial comparing the same seasonal IIV when delivered by IM injection or MNP, we found evidence of a relationship between reactogenicity and immunogenicity in the IM group. Our results suggest that heightened reactogenicity to IM IIV is related to low baseline HAI antibody titers to included antigens, in this case, H3N2. Participants with greater reactogenicity developed greater H3N2 HAI titer fold change after 4 weeks, though the overall magnitude of HAI response was similar or lower compared with lowreactogenicity participants. For those with relatively greater local reactogenicity, who appear to have had the lowest baseline H3N2 HAI titers, the greater titer fold change was associated with a greater probability of seroconversion.

These findings appear to conflict with the SARS-CoV-2 vaccine literature, in which participants without prior infection reported greater reactogenicity and developed most of their immunity from the second (boost) dose of a prime-boost vaccine series. As a result, we had hypothesized that those with greater reactogenicity would have greater resulting immunity. Our results instead show that those with greater reactogenicity had lower baseline markers of humoral immunity. The antibody response of these participants in the short term was higher in the relative sense and similar or lower in the absolute sense than their low-reactogenicity counterparts. The reasons behind these differences between our expectations and our findings are not clear, though there are multiple plausible explanations.

Regarding why those with lower baseline HAI titers might have greater reactogenicity, one possibility is that those with greater preexisting immunity do not undergo the same inflammatory cascade as those with less preexisting immunity, perhaps due to antibody-mediated precipitation and destruction of injected antigen without resulting presentation by APCs in the draining lymph node. This could also explain why the difference was only noted in our study in the IM group, though the MNP group was larger and had more statistical power – the muscle has a sparse distribution of APCs and lymphatic vessels compared to the dermis. Instead of rapid delivery from the skin to the draining lymph node, either by an APC or free within lymphatic vessels, antigen may move more slowly to the draining lymph node from the muscle, increasing the chances that it is bound along the way by IgG, the primary immunoglobulin isotype found in the extracellular space within body tissues.^{91–93}

A complicating factor in this analysis that might explain why participants had differing levels of baseline H3N2 HAI titers is that we are unable to accurately discern each participant's personal history of antigen exposure. We do not have granular data about trial participants' seasonal IIV and influenza infection histories, including what antigens and strains they have been exposed to, how many times, in what order, and how recently. All these pieces of information may affect a person's baseline HAI titer to specific antigens, such as the A/Texas/50/2012, NYMC X-223 used as the H3N2 component in the TIV-MNP 2015 study. We adjusted for receipt of influenza vaccine within the prior 2 years, as recent vaccination may affect responses to a new influenza vaccine via imprinting, but we were unable to adjust for most influenza antigen exposure in the participants' lifetimes. The mismatch of H3N2 vaccine antigen and influenza strain from the preceding influenza season may also have been important here, as having had undiagnosed

influenza A/H3N2 during the prior winter would not have primed participants perfectly for the H3N2 antigen they were exposed to in the trial.

This may also help to explain why these models only generated consistent and significant findings with the H3N2 HAI titer outcome. The H1N1 antigen used in influenza vaccines over the previous 5 years, including in a standalone pandemic influenza vaccine, had not changed. Individuals exposed to pandemic A/H1N1 influenza in 2009, vaccinated against this strain specifically during the pandemic emergency, or vaccinated against seasonal influenza between 2010 and 2014 were effectively primed with the same H1N1 antigen used in the TIV-MNP 2015 trial. This is supported by our findings that median baseline H1N1 HAI titers were higher than H3N2 or B HAI titers and that over 75% of participants had baseline seroprotection against A/H1N1 influenza.

Why we did not find a consistent association between B HAI titer and reactogenicity is harder to rationalize. The median baseline HAI titer was the same against H3N2 and B antigens, and the percentage of participants with baseline seroprotection was lower for the B antigen than for the H3N2 antigen. However, the IM group had higher baseline seroprotection against influenza B than against influenza A/H3N2, and the MNP group had lower baseline seroprotection against influenza B. Consistent with this, among the MNP group, a high Global reaction score was associated with lower baseline B HAI titer and a higher day 28 B HAI titer fold change, mirroring the more durable findings we focused on throughout the study. Despite this signal, we must exercise caution when interpreting influenza B HAI results from a study of a trivalent vaccine for the same reasons discussed above. Many participants most likely had unique personal

histories of influenza B antigen and virus exposure, leading to heterogeneity in their baseline HAI titers and their anamnestic responses to vaccination with an influenza B antigen from one of two co-circulating lineages.

Taken together, it is challenging to compare IIV data to SARS-CoV-2 vaccine data. The COVID-19 pandemic resulted in a novel pathogen exposure and novel vaccines. SARS-CoV-2 vaccine study participants did not have long histories of intermittent exposure to several sequences of antigen at varying ages and periods of immune system development. The observation that people have greater reactogenicity and greater immunity after an initial pair of sequential exposures to a similar antigen is not specific to SARS-CoV-2, but whether this holds true for influenza remains to be seen.

Moreover, our assessment of the relationship between HAI and reactogenicity does not account for other measures of humoral immunity, such as neuraminidase inhibition (NAI) antibody titers, and components of the cellular immune compartment, such as T cells. This is a key limitation of our study (and many others in this space). Several studies suggest that, although it is not the gold standard measurement, NAI functions as an independent correlate of protection against influenza.^{94–98} Similarly, a vast literature has shown that while most approved influenza vaccines do not elicit effective cross-reactive T cell immunity, intranasal live-attenuated influenza vaccine and influenza infection generate such cellular immune responses to a broad range of influenza antigens.^{99–101} Thus, we cannot assume that participants with unknown exposure histories and lower baseline HAI titers have lower baseline NAI titers or lower numbers of preexisting central memory T cells, for example. Further, since symptoms like fever and headache are well accepted to be driven by cytokines, which are predominantly secreted by the cellular immune compartment, attempts to correlate HAI titers with reactogenicity, particularly systemic reactogenicity, may be utilizing a measure of humoral immunity as a surrogate for measures of cellular immunity. Though HAI titer remains a standard correlate of protection for FDA approval of vaccines against influenza, in part due to the simplicity and reproducibility of testing peripheral blood, this metric cannot entirely explain vaccine immunogenicity or reactogenicity. Whenever possible, future investigations should incorporate measurements of NAI and various cell populations, particularly from tissues such as draining lymph nodes, to evaluate additional associations with reactogenicity. Ultimately, one goal of this line of investigation is to find reactogenicity correlates of vaccine efficacy, a relationship that we expect may be mediated by multiple variables that collectively describe immunogenicity.

Our study has some additional limitations. We took an a priori approach to defining reactogenicity variables as composite scores. These unweighted scores assume that each type of adverse event has the same impact on immunogenicity (e.g., fatigue and myalgia carry the same amount of meaning). However, we also recognize that adverse event grades are highly subjective. Summing grades of events instead of numbers of events could have introduced additional subjectivity to our models. Moreover, using sums of grades injects another assumption: that individual event grades (e.g., headache grades 1, 2, and 3) are equal distances apart. Ultimately, we chose our primary measure of reactogenicity because it is generalizable to future research, interpretable, and can be implemented clinically. Creating a bespoke, weighted composite reactogenicity score in future studies, for example by using LASSO regression techniques or unsupervised learning methods, to find a smaller number of more important

adverse event types would be instructive in determining if certain events matter more than others in the generation of immunity. However, given the small sample size in this study, such an approach would likely forfeit generalizability to larger databases if it were feasible at all. Our approach is supported by multiple publications in the SARS-CoV-2 literature, in which numbers of adverse events were added together to define groups of participants or variables for modeling. Though there is no metric to confirm the construct validity of our score definition or our dichotomization cut points, the fact that the findings were consistent and robust to at least some sensitivity analyses lends credence to our chosen approach.

The small sample size, large number of variables, and multitude of model outputs collectively represent another limitation of the study. Incorporating longitudinal data into mixed models ANOVA and GEEs mitigated the statistical power limitations (and prevented additional statistical comparisons) to some extent, but it remained challenging to prioritize and interpret model outputs. In some cases, particularly those involving the relative variables of titer fold change and seroconversion, using longitudinal models may have reduced the clarity of the findings. As an example, day 180 seroconversion carries less meaning than day 28 seroconversion, and its inclusion in GEEs obscured the association between local reactogenicity and seroconversion in the longitudinal estimates. At the same time, this study accomplished the function of a screening experiment successfully. We used advanced biostatistical methods on a small sample to identify signals for future investigation and generate hypotheses for future testing.

One more limitation is the low generalizability of our sample, which only included young and middle-aged adults without a wide range of self-reported racial and ethnic diversity. Though race and ethnicity should not have biologic significance regarding vaccine responses, we still adjusted for these variables due to associations we found in our preliminary bivariable comparisons and because they may act as proxies for unmeasured confounding variables that do affect vaccine biology. Future analyses of the influenza immunogenicity-reactogenicity relationship will need to include a more generalizable sample of healthy participants.

To address the intriguing findings and limitations of this pilot study, our group has acquired data from 12 completed phase 1 and 2 multicenter clinical trials of vaccines designed against pandemic strains of influenza. These vaccines delivered novel antigens to previously unexposed participants via intramuscular injection on a prime-boost schedule. With a larger database, broader source population, and sequential vaccine schedule, free of variation in participant exposure history, we will be able to meta-analyze these studies with better control of confounding to bridge the knowledge gap between our understanding of the immunogenicityreactogenicity relationships of SARS-CoV-2 vaccines and influenza vaccines. Beyond this, we also plan to investigate the molecular underpinnings of reactogenicity, a pursuit that may help us to ultimately answer several critical outstanding questions, including questions about significant vaccine-associated adverse events such as Guillain-Barré syndrome and myocarditis.

In conclusion, greater reactogenicity of intramuscular seasonal inactivated influenza vaccine was associated with lower baseline H3N2 immunity and greater antibody titer fold change after 28 days, though the overall magnitude of the response was similar or lower compared with low-

reactogenicity participants. Future analyses of influenza vaccine trials will further elucidate their immunogenicity-reactogenicity relationship, which has wide-ranging implications for patient care, public health, and vaccine development.

<u>Tables</u>

Table 1. Baseline Participant Characteristic	Table 1	. Baseline	Participant	Characteristics
--	---------	------------	--------------------	-----------------

Table 1. Dasenne I ar tierpa							
Characteristic*	IM	MNP	All				
Characteristic	(n = 25)	(n = 50)	(n = 75)				
Age, year	28.0 (7.0)	26.0 (10.0)	27.0 (9.0)				
BMI, kg/m ²	24.8 (6.9)	24.8 (5.3)	24.8 (6.0)				
≤ 25	14 (56.0%)	26 (52.0%)	40 (53.3%)				
25-30, not inclusive	9 (36.0%)	17 (34.0%)	26 (34.7%)				
\geq 30	2 (8.0%)	7 (14.0%)	9 (12.0%)				
Sex							
Female	11 (44.0%)	24 (48.0%)	35 (46.7%)				
Male	14 (56.0%)	26 (52.0%)	40 (53.3%)				
Race							
White	12 (48.0%)	25 (50.0%)	37 (49.3%)				
Black	8 (32.0%)	16 (32.0%)	24 (32.0%)				
Other	5 (20.0%)	9 (18.0%)	14 (18.7%)				
Ethnicity							
Not LatinX	22 (88.0%)	48 (96.0%)	70 (93.3%)				
LatinX	3 (12.0%)	2 (4.0%)	5 (6.7%)				
IIV in prior 2 seasons	· · ·						
No	17 (68.0%)	34 (68.0%)	51 (68.0%)				
Yes	8 (32.0%)	16 (32.0%)	24 (32.0%)				
Baseline HAI titer^	· · ·						
H1N1	80.0 (80.0)	80.0 (140.0)	80.0 (120.0)				
H3N2	40.0 (60.0)	40.0 (65.0)	40.0 (60.0)				
В	40.0 (40.0)	20.0 (30.0)	40.0 (70.0)				
Missing	Ò	2 (4%)	2 (2.7%)				
DML hadrenses in den HW. in a timeted influence and include how a shoting time in hit iden							

BMI: body mass index; IIV: inactivated influenza vaccine; HAI: hemagglutination inhibition

*continuous variables: median (IQR); categorical variables: N (%)

^represented as inverse titers

Reaction Action Study Unadjusted GMR* Adjusted GMR*						
Туре	Antigen	Study Group	(95% CI)	p-value	(95% CI)	p-value
Type		Oloup	1.07		0.98	
		IM	(0.53, 2.16)	0.85	(0.58, 1.66)	0.95
	H1N1		0.87		0.85	
		MNP	(0.50, 1.52)	0.62	(0.58, 1.24)	0.38
			0.83		0.74	
		IM	(0.83) (0.47, 1.49)	0.54	(0.45, 1.22)	0.23
Global	H3N2		0.81		0.78	
		MNP	(0.50, 1.29)	0.37	(0.54, 1.12)	0.17
		D (1.21	0.00	0.95	0.50
	D	IM	(0.80, 1.83)	0.36	(0.63, 1.42)	0.79
	В	MNID	0.80	0.20	0.81	0.22
		MNP	(0.53, 1.22)	0.30	(0.58, 1.14)	0.22
		IM	0.73	0.40	0.65	0.12
	H1N1	1101	(0.35, 1.52)	0.40	(0.38, 1.12)	0.12
		MNP	0.91	0.83	0.85	0.61
		(0.36, 2.27)	(0.45, 1.60)	0.01		
	H3N2 B	IM	0.64 0.15	0.15	0.54	0.02
Systemic		1111	(0.35, 1.18)	0.15	(0.33, 0.89)	0.02
Systemic		MNP	1.43	0.36	1.42	0.26
		(0.66, 3.10)	0.50	(0.77, 2.60)	0.20	
		IM	1.07	0.77	0.81	0.33 0.38
		1101	(0.69, 1.65)		(0.53, 1.24)	
		MNP	0.84		0.78	
			(0.42, 1.66)	0101	(0.45, 1.36)	
	H1N1	IM	0.59	0.27	0.6	0.15
			(0.22, 1.54)		(0.30, 1.21)	
Local		MNP	1.53		1.54	0.03
			(0.90, 2.61)		(1.05, 2.25)	
	H3N2	MNP 0.72 0.15		0.05	0.37	<0.01
			(0.21, 1.01)		(0.19, 0.72)	
			0.15	0.63	0.01	
	В	IM (0.46, 1	(0.46, 1.13)	0.85	(0.44, 0.90)	0.68
					0.89	
			(0.60, 1.87)		(0.51, 1.55)	
		MNP	0.97		1.00	
TTAT 1		1.1.1.2	(0.65, 1.45)	CLUD	(0.71, 1.41)	

Table 2. Longitudinal Associations Between Reaction Score and HAI GMT

HAI: hemagglutination inhibition; GMT: geometric mean titer; GMR: geometric mean titer ratio; CI: confidence interval; IM: intramuscular; MNP: microneedle patch;

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

^Adjusted for body mass index category, sex, prior inactivated influenza vaccination, race, ethnicity, and study day

Reaction	A	Study	Unadjusted GMR*		Adjusted GMR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
		IM	0.96	0.90	0.93	0.77
	H1N1	1101	(0.51, 1.81)		(0.58, 1.51)	
		MNP	0.94	0.81	0.91	0.59
			(0.56, 1.58)	0.01	(0.64, 1.29)	0.09
~1.1.1		IM	0.97	0.90	0.84	0.45
Global	H3N2		(0.57, 1.64)		(0.53, 1.33)	
$(\geq 3 \text{ vs} \leq 2)$	-	MNP	0.81	0.35	0.78	0.15
			(0.53, 1.26)		(0.56, 1.10)	
		IM	1.47	0.04	1.16	0.42
	В		(1.02, 2.12)		(0.80, 1.69)	
		MNP	1.04	0.82	1.05	0.77
	H1N1		(0.71, 1.53)		(0.76, 1.44)	
		IM	0.85	0.68	0.81	0.47
		MNP	(0.39, 1.87)	0.91	(0.45, 1.45)	0.82
			1.03		0.95 (0.64, 1.43)	
			(0.57, 1.86) 0.82	0.55	0.73	
Global $(\geq 5 \text{ vs} \leq 4)$	H3N2	IM	(0.82) (0.43, 1.58)		(0.42, 1.26)	0.25
		3N2 MNP	1.00	1.0	0.94	0.76
			(0.61, 1.64)		(0.64, 1.39)	
	В		0.91		0.69	0.10
		IM	(0.57, 1.45)	0.69	(0.44, 1.08)	
		B MNP	0.91	0.68	0.90	0.54
			(0.59, 1.42)		(0.63, 1.28)	
	l		(0.57, 1.72)		(0.05, 1.20)	

 Table 3A. Longitudinal Associations Between Alternative Cut Point Global Reaction

 Score and HAI GMT

HAI: hemagglutination inhibition; GMT: geometric mean titer; GMR: geometric mean titer ratio; CI: confidence interval; IM: intramuscular; MNP: microneedle patch;

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

^Adjusted for body mass index category, sex, prior inactivated influenza vaccination, race, ethnicity, and study day
Reaction	Antion	Study	Unadjusted GMR*		Adjusted GMR*^		
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value	
		IM	1.15	0.67	1.12	0.63	
	H1N1		(0.60, 2.18)		(0.70, 1.81)		
		MNP	0.81 (0.49, 1.35)	0.42	0.8 (0.57, 1.13)	0.21	
		IM	0.95	0.96	0.84	0.44	
Systemic	H3N2	IM	(0.56, 1.63)	0.86	(0.53, 1.32)	0.44	
(≥ 1 vs 0)	1131\2	MNP	1.02	0.94	1.03	0.87	
		101111	(0.66, 1.56)	0.91	(0.74, 1.44)	0.07	
	В	IM	1.54	0.02	1.22	0.29	
			(1.07, 2.22)		(0.84, 1.76)		
		MNP	0.87	0.48	0.89	0.47	
			(0.60, 1.28)		(0.66, 1.21)		
		IM	1.03 (0.55, 1.93)	0.94	1.0 (0.63, 1.59)	0.99	
	H1N1		1.04	0.00	1.12	0.56	
		MNP	(0.59, 1.84)	0.89	(0.76, 1.65)		
		IM	0.79	0.37	0.72	0.13	
Systemic	H3N2	11VI	(0.47, 1.33)	0.57	(0.46, 1.11)	0.13	
$(\geq 2 \text{ vs} \leq 1)$	113112	MNP	0.99	0.95	0.99	0.94	
			(0.61, 1.60)	0.75	(0.68, 1.44)	0.74	
	В	IM	1.42	0.06	1.17	0.39	
			(0.99, 2.05)	0.00	(0.82, 1.67)	0.57	
		MNP	0.86	0.50	0.87	0.43	
			(0.56, 1.33)		(0.62, 1.23)		

 Table 3B. Longitudinal Associations Between Alternative Cut Point Systemic Reaction

 Score and HAI GMT

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

Reaction	A	Study	Unadjusted GMR*		Adjusted GMR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
	H1N1	IM	1.40 (0.71, 2.73)	0.32	1.34 (0.81, 2.21)	0.25
		MNP	$ 1.18 \\ (0.51, 2.71) $	0.69	1.00 (0.57, 1.77)	1.0
Local	H3N2	IM	1.01 (0.58, 1.77)	0.97	0.92 (0.57, 1.49)	0.73
(≥ 1 vs 0)	113112	MNP	1.00 (0.49, 2.01)	0.99	0.94 (0.54, 1.63)	0.81
	В	IM	1.18 (0.80, 1.76)	0.40	0.95 (0.64, 1.40)	0.79
		MNP	1.19 (0.64, 2.21)	0.59	1.13 (0.68, 1.87)	0.64
	H1N1	IM	1.10 (0.58, 2.09)	0.78	1.02 (0.63, 1.66)	0.92
		MNP	$ 1.19 \\ (0.69, 2.05) $	0.53	1.05 (0.72, 1.55)	0.79
Local	H3N2	IM	1.22 (0.72, 2.08)	0.46	1.05 (0.66, 1.67)	0.83
$(\geq 2 vs \leq 1)$	113112	MNP	0.79 (0.50, 1.26)	0.32	0.73 (0.50, 1.05)	0.09
	В	IM	1.20 (0.83, 1.75)	0.33	0.92 (0.63, 1.34)	0.66
		MNP	1.30 (0.87, 1.96)	0.20	1.25 (0.89, 1.76)	0.19

 Table 3C. Longitudinal Associations Between Alternative Cut Point Local Reaction

 Score and HAI GMT

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

Reaction	Study	Day 0 GMR*	p-	Day 28 GMR*	p-	Day 180 GMR*	p-
Туре	Group	(95% CI)	value	(95% CI)	value	(95% CI)	value
			Unadj	usted			
Global	IM	1.0 (0.4, 2.4) 1.2	0.99	1.1 (0.5, 2.5)	0.87	1.1 (0.5, 2.7)	0.77
Giobai	MNP	1.2 (0.6, 2.3)	0.59	0.7 (0.4, 1.4)	0.36	0.7 (0.4, 1.4)	0.35
Systemic	IM	0.6 (0.2, 1.4)	0.21	0.9 (0.4, 2.1)	0.78	0.8 (0.3, 1.9)	0.58
Systemic	MNP	1.5 (0.5, 4.2)	0.48	0.7 (0.2, 2.0)	0.50	0.7 (0.3, 2.1)	0.56
Local	IM	1.0 (0.3, 3.3)	0.94	0.5 (0.1, 1.5) 1.5	0.19	0.4 (0.1, 1.3) 1.5	0.14
Local	MNP	1.6 (0.9, 2.9)	0.14	(0.8, 2.8)	0.18	1.5 (0.8, 2.8)	0.18
			Adjus	sted^			
Global	IM	0.9 (0.4, 2.2)	0.86	1.0 (0.4, 2.3)	0.98	1.0 (0.4, 2.4)	0.92
Giotai	MNP	1.2 (0.6, 2.2)	0.64	0.7 (0.4, 1.4)	0.32	0.7 (0.4, 1.4)	0.31
Systemic	IM	0.5 (0.2, 1.2)	0.13	0.8 (0.3, 1.9)	0.59	0.7 (0.3, 1.7)	0.41
Systemic	MNP	1.4 (0.5, 3.9)	0.55	0.7 (0.2, 1.9) 0.5	0.42	0.7 (0.2, 2.0)	0.48
Local	IM	$ \begin{array}{r} 1.1 \\ (0.3, 3.3) \end{array} $	0.91	0.5 (0.2, 1.5) 1.5	0.20	0.4 (0.1, 1.3)	0.14
	MNP	1.6 (0.9, 2.9)	0.14	(0.8, 2.8)	0.19	1.5 (0.8, 2.8)	0.19

Table 4A. Reaction Score and H1N1 HAI GMT by Study Day

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Study Group	Day 0 GMR* (95% CI)	p- value	Day 28 GMR* (95% CI)	p- value	Day 180 GMR* (95% CI)	p- value
		1	Unadj			1	
Global	IM	$ \begin{array}{c} 0.5 \\ (0.2, 1.1) \end{array} $	0.09	1.1 (0.5, 2.4)	0.82	1.0 (0.5, 2.2)	0.93
Giobai	MNP	0.8 (0.4, 1.5)	0.53	0.7 (0.4, 1.4)	0.32	0.9 (0.5, 1.6)	0.69
Sustamia	IM	0.4 (0.2, 0.9)	0.03	1.0 (0.5, 2.3)	0.96	0.6 (0.3, 1.4)	0.25
Systemic	MNP	1.3 (0.5, 3.6)	0.63	1.5 (0.5, 4.1)	0.47	1.6 (0.6, 4.4)	0.40
Local	IM	0.3 (0.1, 0.9)	0.03	0.7 (0.2, 1.9)	0.48	0.5 (0.2, 1.3)	0.13
Local	MNP	0.6 (0.4, 1.2)	0.16	0.7 (0.4, 1.3)	0.29	0.8 (0.4, 1.4)	0.43
			Adjus	sted^			
Global	IM	0.5 (0.2, 1.0)	0.06	1.0 (0.4, 2.2)	0.95	0.9 (0.4, 2.1)	0.85
Global	MNP	0.8 (0.4, 1.5)	0.45	0.7 (0.4, 1.3)	0.26	0.8 (0.5, 1.6)	0.60
Systemic	IM	0.4 (0.2, 0.8)	0.01	0.9 (0.4, 1.9)	0.7	0.5 (0.2, 1.2)	0.12
Systemic	MNP	1.3 (0.5, 3.5)	0.63	1.4 (0.5, 4.0)	0.47	1.5 (0.6, 4.3)	0.40
Local	IM	0.3 (0.1, 0.8)	0.01	0.6 (0.2, 1.6)	0.28	0.4 (0.1, 1.1)	0.07
Local	MNP	0.6 (0.3, 1.0)	0.06	0.6 (0.4, 1.1)	0.13	0.7 (0.4, 1.2)	0.22

Table 4B. Reaction Score and H3N2 HAI GMT by Study Day

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Study Group	Day 0 GMR* (95% CI)	p- value	Day 28 GMR* (95% CI)	p- value	Day 180 GMR* (95% CI)	p- value
		n	Unadj			n	
Global	IM	1.2 (0.6, 2.3)	0.54	1.2 (0.6, 2.3) 1.1	0.58	1.2 (0.6, 2.3)	0.55
Giobai	MNP	0.6 (0.3, 1.0)	0.05	(0.6, 1.9)	0.84	0.9 (0.5, 1.6)	0.66
Systemia	IM	0.9 (0.5, 1.8)	0.83	1.1 (0.6, 2.1)	0.80	1.2 (0.6, 2.4)	0.60
Systemic	MNP	1.0 (0.4, 2.5) 1.2	0.92	0.9 (0.4, 2.5)	0.90	0.7 (0.2, 1.7) 1.2	0.39
Local	IM	(0.5, 3.0)	0.66	0.8 (0.3, 2.0) 1.1	0.68	1.2 (0.5, 2.9) 1.1	0.73
Local	MNP	0.7 (0.4, 1.3)	0.26	1.1 (0.6, 2.0)	0.71	1.1 (0.6, 2.0)	0.65
			Adjus	sted^			
Global	IM	1.0 (0.5, 1.8)	0.89	0.9 (0.5, 1.8)	0.85	0.9 (0.5, 1.8)	0.87
Giobal	MNP	0.6 (0.3, 1.0)	0.04	1.1 (0.6, 1.9)	0.80	0.9 (0.5, 1.5)	0.67
Systemic	IM	0.7 (0.4, 1.4)	0.32	0.8 (0.4, 1.6)	0.59	0.9 (0.5, 1.8)	0.80
Systemic	MNP	0.9 (0.4, 2.2)	0.81	0.9 (0.3, 2.2)	0.78	0.6 (0.2, 1.5)	0.29
Local	IM	1.0 (0.4, 2.5)	0.95	0.7 (0.3, 1.7)	0.42	1.0 (0.4, 2.4)	0.98
	MNP	$\begin{array}{c} 0.7 \\ (0.4, 1.3) \end{array}$	0.29	1.1 (0.7, 2.0)	0.62	1.2 (0.7, 2.0)	0.57

Table 4C. Reaction Score and B HAI GMT by Study Day

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Day 0 Adjusted GMR*^ (95% CI)	p- value	Day 28 Adjusted GMR*^ (95% CI)	p- value	Day 180 Adjusted GMR*^ (95% CI)	p- value
Global	0.5 (0.2, 1.0)	0.06	1.0 (0.4, 2.2)	0.95	0.9 (0.4, 2.1)	0.85
Systemic	0.4 (0.2, 0.8)	0.01	0.9 (0.4, 1.9)	0.7	0.5 (0.2, 1.2)	0.12
Local	0.3 (0.1, 0.8)	0.01	0.6 (0.2, 1.6)	0.28	0.4 (0.1, 1.1)	0.07

Table 5. IM Study Group Reaction Score and H3N2 HAI GMT by Study Day

IM: intramuscular; HAI: hemagglutination inhibition; GMT: geometric mean titer; GMR: geometric mean titer ratio; CI: confidence interval;

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Study Group	Day 28 FCR* (95% CI)	p- value	Day 180 FCR* (95% CI)	p- value
		Unadjus	ted	1	
Global	IM	$ \begin{array}{c} 1.1 \\ (0.3, 3.7) \end{array} $	0.92	1.1 (0.3, 3.9)	0.85
Giobai	MNP	0.6 (0.3, 1.4)	0.26	0.6 (0.3, 1.4)	0.25
Sustamia	IM	1.5 (0.4, 5.6)	0.51	1.4 (0.4, 5.0)	0.63
Systemic	MNP	0.5 (0.1, 1.8)	0.28	0.5 (0.1, 1.9)	0.31
Local	IM	0.4 (0.1, 2.4)	0.34	0.4 (0.1, 2.2)	0.28
Local	MNP	1.0 (0.4, 2.1)	0.91	1.0 (0.4, 2.1)	0.91
		Adjuste	d^		
Global	IM	1.0 (0.3, 3.1)	0.94	1.0 (0.3, 3.3)	0.99
Giobai	MNP	0.7 (0.3, 1.4)	0.30	0.7 (0.3, 1.4)	0.29
Sustamia	IM	1.1 (0.3, 4.0)	0.84	1.0 (0.3, 3.5)	1.0
Systemic	MNP	0.6 (0.2, 2.0)	0.41	0.6 (0.2, 2.1)	0.45
Local	IM	0.4 (0.1, 2.2)	0.32	0.4 (0.1, 2.0)	0.27
Local	MNP	$ \begin{array}{c} 1.0 \\ (0.5, 2.2) \end{array} $	0.90	1.0 (0.5, 2.2)	0.90

Table 6A. Reaction Score and H1N1 HAI Titer FoldChange by Study Day

HAI: hemagglutination inhibition; FCR: fold change ratio; CI: confidence interval; IM: intramuscular; MNP: microneedle patch; *Estimated fold change ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Study Group	Day 28 FCR* (95% CI)	p- value	Day 180 FCR* (95% CI)	p- value
		Unadjus	ted		
Global	IM	2.1 (1.1, 4.1)	0.03	2 (1.0, 3.9)	0.04
	MNP	$ \begin{array}{c} 0.9 \\ (0.5, 1.6) \end{array} $	0.68	1.1 (0.6, 1.9)	0.81
Systemia	IM	2.4 (1.2, 5.0)	0.02	1.5 (0.7, 3.1)	0.27
Systemic	MNP	$ 1.1 \\ (0.4, 2.9) $	0.79	1.2 (0.5, 3.1)	0.69
Local	IM	2.2 (0.8, 5.7)	0.12	1.4 (0.5, 3.8)	0.48
Local	MNP	1.1 (0.6, 1.9)	0.69	1.2 (0.7, 2.1)	0.48
		Adjuste	d^		
Global	IM	2 (1.0, 3.7)	0.04	1.9 (1.0, 3.5) 1.1	0.05
Giobai	MNP	0.9 (0.6, 1.6)	0.83	1.1 (0.7, 1.9)	0.61
Systemic	IM	1.9 (1.0, 3.8)	0.07	1.2 (0.6, 2.4)	0.64
Systemic	MNP	1.3 (0.6, 3.1)	0.49	1.4 (0.6, 3.3)	0.40
Local	IM	2 (0.8, 4.9) 1.1	0.15	1.3 (0.5, 3.2)	0.58
Local	MNP	1.1 (0.7, 1.9)	0.66	1.2 (0.7, 2.0)	0.44

Table 6B. Reaction Score and H3N2 HAI Titer FoldChange by Study Day

HAI: hemagglutination inhibition; FCR: fold change ratio; CI: confidence interval; IM: intramuscular; MNP: microneedle patch; *Estimated fold change ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Study Group	Day 28 FCR* (95% CI)	p- value	Day 180 FCR* (95% CI)	p- value
		Unadjus	ted		
Global	IM	1.0 (0.4, 2.3)	0.97	1.0 (0.4, 2.4)	0.99
Giobal	MNP	1.9 (1.0, 3.6)	0.05	1.6 (0.8, 3.0)	0.16
Systemic	IM	1.2 (0.5, 2.9)	0.73	1.3 (0.5, 3.2)	0.58
Systemic	MNP	$ \begin{array}{c} 1.0 \\ (0.3, 2.9) \end{array} $	0.98	0.7 (0.2, 2.0)	0.49
Local	IM	0.7 (0.2, 2.2) 1.5	0.52	1.0 (0.3, 3.1)	0.94
Local	MNP	1.5 (0.8, 2.9)	0.17	1.6 (0.8, 2.9)	0.15
		Adjuste	d^		
Global	IM	0.8 (0.4, 1.6)	0.46	0.8 (0.4, 1.6)	0.48
Giobai	MNP	1.8 (1.1, 3.2)	0.03	1.5 (0.9, 2.6)	0.14
Systemia	IM	0.9 (0.4, 1.9)	0.70	0.9 (0.4, 2.1)	0.88
Systemic	MNP	1.1 (0.4, 2.9)	0.79	0.8 (0.3, 2.0)	0.61
Local	IM	0.5 (0.2, 1.5) 1.3	0.22	0.7 (0.3, 2.1) 1.3	0.58
LUCAI	MNP	$ \begin{array}{c c} 1.3 \\ (0.7, 2.2) \\ \hline \end{array} $	0.39	1.3 (0.7, 2.3)	0.35

 Table 6C. Reaction Score and B HAI Titer Fold Change

 by Study Day______

HAI: hemagglutination inhibition; FCR: fold change ratio; CI: confidence interval; IM: intramuscular; MNP: microneedle patch; *Estimated fold change ratio for those in the higher score category compared with those in the lower score category

		care a second	cores and m		i by Study			
Reaction Type	Antigen	Study Group	Day 0 GMR* (95% CI)	p- value	Day 28 GMR* (95% CI)	p- value	Day 180 GMR* (95% CI)	p- value
	H1N1	IM	0.9 (0.3, 2.8)	0.87	0.8 (0.3, 2.5)	0.74	0.9 (0.3, 2.8)	0.87
	HINI	MNP	$ 1.7 \\ (0.5, 5.3) $	0.35	1.6 (0.5, 4.8)	0.44	1.4 (0.4, 4.2)	0.60
Global	H3N2	IM	0.6 (0.2, 1.6)	0.28	1.2 (0.4, 3.4) 0.7	0.73	1.1 (0.4, 3.2) 0.9	0.84
Giobai	113112	MNP	$ \begin{array}{c} 0.5 \\ (0.2, 1.5) \end{array} $	0.24	0.7 (0.3, 1.9) 1.2	0.47	0.9 (0.3, 2.4) 1.5	0.75
	В	IM	0.9 (0.4, 1.8)	0.68	(0.5, 2.5)	0.68	1.5 (0.7, 3.2) 1.1	0.29
	В	MNP	$ \begin{array}{c} 0.8 \\ (0.4, 1.8) \end{array} $	0.60	1.0 (0.5, 2.3)	0.95	(0.5, 2.4)	0.86
	H1N1	IM	0.5 (0.2, 1.7) 1.5	0.30	0.7 (0.2, 2.3)	0.55	0.5 (0.2, 1.6)	0.24
		MNP	$ \begin{array}{r} 1.5 \\ (0.5, 4.5) \\ 0.4 \end{array} $	0.44	0.8 (0.3, 2.3)	0.63	0.6 (0.2, 1.8) 0.8	0.35
Systemic	H3N2	IM	0.4 (0.1, 1.3) 0.5	0.14	1.2 (0.4, 3.8) 0.6	0.76	0.8 (0.2, 2.4) 0.7	0.65
Systemic		MNP	(0.2, 1.1)	0.08	0.6 (0.3, 1.4) 1.5	0.23	0.7 (0.3, 1.5) 1.5	0.38
	В	IM	1.0 (0.4, 2.6) 1.2	1.0	1.5 (0.6, 4.0) 0.7	0.36	$ \begin{array}{r} 1.5 \\ (0.6, 4.0) \\ 0.6 \end{array} $	0.36
	D	MNP	1.2 (0.6, 2.4) 4.4	0.69	0.7 (0.4, 1.5) 0.3	0.41	$ \begin{array}{r} 0.6 \\ (0.3, 1.3) \\ 0.6 \end{array} $	0.20
	H1N1	IM	(1.1, 17.7)	0.04	(0.1, 1.1)	0.07	0.6 (0.2, 2.6) 1.4	0.54
	111111	MNP	1.2 (0.5, 2.9)	0.61	1.2 (0.5, 2.7)	0.73	(0.6, 3.1)	0.47
Local	H3N2	IM	0.2 (0.1, 0.8)	0.02	$ \begin{array}{c} 0.3 \\ (0.1, 1.1) \end{array} $	0.06	0.2 (0.1, 0.8)	0.02
LUCAI	113112	MNP	0.5 (0.2, 1.4)	0.17	0.6 (0.2, 1.8)	0.40	0.7 (0.2, 1.8)	0.42
	В	IM	2.0 (0.9, 4.3)	0.08	0.9 (0.4, 2.0)	0.82	1.4 (0.7, 3.1)	0.37
	U	MNP	0.8 (0.4, 1.9)	0.63	0.9 (0.4, 2.1)	0.87	1.3 (0.5, 2.9)	0.58

Table 7. Extreme Reaction Scores and HAI GMT by Study Day

*Estimated geometric mean titer ratio for those in the higher score category compared with those in the lower score category

Study Da	<u>y</u>			1		1
Reaction Type	Antigen	Study Group	Day 0 FCR* (95% CI)	p- value	Day 28 FCR * (95% CI)	p- value
	H1N1	IM	0.9 (0.2, 5.3) 0.9	0.92	1.0 (0.2, 5.9) 0.8	1.0
	IIINI	MNP	0.9 (0.3, 2.8) 2.1	0.87	0.8 (0.3, 2.5) 2	0.68
Global	H3N2	IM	2.1 (1.0, 4.7) 1.3	0.06	2 (0.9, 4.4) 1.6	0.09
Giobai	113112	MNP	$ \begin{array}{r} 1.3 \\ (0.5, 3.1) \\ 1.4 \end{array} $	0.61	1.6 (0.6, 3.8) 1.7	0.33
	В	IM	1.4 (0.8, 2.4) 1.3	0.29	1.7 (1.0, 3.1) 1.3	0.06
	В	MNP	1.3 (0.5, 3.0) 1.3	0.59	(0.5, 3.2)	0.52
	H1N1	IM	$ \begin{array}{r} 1.3 \\ (0.2, 8.4) \\ 0.5 \end{array} $	0.78	0.9 (0.1, 6.0) 0.4	0.92
		MNP	0.5 (0.2, 1.4) 2.8	0.18	0.4 (0.1, 1.1) 1.8	0.07
Sustamia	H3N2	IM		0.02	1.8 (0.8, 4.2) 1.4	0.15
Systemic		MNP	(1.2, 6.5) 1.3 (0.6, 2.6) 1.5	0.54	1.4 (0.7, 3.0) 1.5	0.34
	В	IM	1.5 (0.7, 3.3)	0.25	1.5 (0.7, 3.3) 0.5	0.25
	D	MNP	0.6 (0.3, 1.5)	0.31	0.5 (0.2, 1.3) 0.1	0.17
	H1N1	IM	0.1 (0.0, 0.5)	0.01	0.1 (0.0, 1.2)	0.07
	111111	MNP	0.9 (0.3, 3.1)	0.91	$ 1.1 \\ (0.3, 3.7) $	0.88
Local	H3N2	IM	1.4 (0.5, 4.2) 1.3	0.52	1.1 (0.4, 3.2) 1.4	0.87
Local	ПЭIN2	MNP	$ \begin{array}{r} 1.3 \\ (0.6, 3.2) \\ 0.5 \end{array} $	0.52	1.4 (0.6, 3.3) 0.7	0.48
	B -	IM	0.5 (0.2, 1.2) 1.1	0.11	0.7 (0.3, 1.9) 1.5	0.47
		MNP	(0.4, 3.0)	0.79	(0.6, 4.1)	0.38
TTAT. Is a second	1.1.1.1.1.1.1.1.1.1.1.1		"D. fold ahang	CI CI	<u>c 1</u>	4 · · · · · · 1 ·

 Table 8. Extreme Reaction Scores and HAI Titer Fold Change by

 Study Day

*Estimated fold change ratio for those in the higher score category compared with those in the lower score category

Reaction	Antigen	Study Group	Unadjusted GMR* (95% CI)	p-value	Adjusted GMR*^ (95% CI)	p-value
	H1N1	IM	1.1 (0.6, 2.2)	0.69	1.1 (0.6, 2.2)	0.75
	HINI	MNP	1.3 (0.7, 2.3)	0.42	1.4 (0.8, 2.5)	0.30
Severe	H3N2	IM	0.9 (0.5, 1.6)	0.80	0.8 (0.5, 1.5)	0.52
(Yes vs. No)		MNP	0.8 (0.5, 1.3)	0.41	0.8 (0.5, 1.3)	0.38
	В	IM	1.4 (0.9, 2.0)	0.11	1.1 (0.8, 1.7)	0.58
		MNP	1.1 (0.7, 1.7)	0.77	1.2 (0.8, 1.8)	0.47

Table 9. Reaction Severity and Longitudinal HAI GMT

*Estimated geometric mean titer ratio for those in the higher severity category compared with those in the lower severity category

Reaction	Antigen	Study Group	Day 28 FCR* (95% CI)	p- value	Day 180 FCR* (95% CI)	p- value			
Unadjusted									
	H1N1	IM	0.7 (0.2, 2.0) 0.5	0.46	0.7 (0.2, 2.3) 0.6	0.60			
	111111	MNP	(0.2, 1.2)	0.12	0.6 (0.3, 1.5) 1.5	0.28			
Severe	H3N2	IM	1.7 (0.9, 3.2) 0.9	0.12	1.5 (0.8, 2.8) 1.5	0.23			
(Yes vs. No)	H3N2	MNP	0.9 (0.5, 1.7)	0.80	1.5 (0.8, 2.7) 1.1	0.20			
	В	IM	$ \begin{array}{r} 1.0 \\ (0.5, 2.3) \\ \overline{1.0} \end{array} $	0.91	1.1 (0.5, 2.4) 1.2	0.82			
	В	MNP	1.0 (0.5, 2.0)	0.96	1.2 (0.6, 2.3)	0.66			
		A	Adjusted^						
	H1N1	IM	0.6 (0.2, 1.9)	0.40	0.7 (0.2, 2.1) 0.7	0.53			
	111111	MNP	0.6 (0.3, 1.3)	0.17	0.7 (0.3, 1.6)	0.41			
Severe	H3N2	IM	1.5 (0.8, 2.8)	0.17	1.4 (0.7, 2.5)	0.32			
(Yes vs. No)	113112	MNP	1.0 (0.6, 1.7)	0.99	1.6 (0.9, 2.7)	0.09			
	в	IM	1.0 (0.5, 2.1)	0.91	1.1 (0.5, 2.2)	0.80			
	В	MNP	1.1 (0.6, 1.9)	0.87	1.2 (0.7, 2.3)	0.46			

Table 10. Reaction Severity and HAI Titer Fold Change by Study Day

*Estimated fold change ratio for those in the higher severity category compared with those in the lower severity category

Reaction	Antigen	Study Group	Unadjusted GMR* (95% CI)	p-value	Adjusted GMR*^ (95% CI)	p-value
	H1N1	IM	1.0 (0.6, 2.0)	0.90	1.0 (0.5, 2.0)	0.94
		MNP	1.4 (0.8, 2.3)	0.26	1.3 (0.8, 2.3)	0.34
Prolonged	H3N2	IM	1.2 (0.7, 2.0)	0.58	1.1 (0.6, 1.9)	0.85
(Yes vs. No)		MNP	1.0 (0.6, 1.5)	0.89	1.0 (0.6, 1.6)	1.0
	В	IM	1.3 (0.9, 1.9)	0.18	1.0 (0.7, 1.5)	0.99
		MNP	0.9 (0.6, 1.4)	0.77	1.0 (0.7, 1.5)	0.94

Table 11. Reaction Duration and Longitudinal HAI GMT

*Estimated geometric mean titer ratio for those in the prolonged category compared with those not in the prolonged category

Reaction	Antigen	Study Group	Day 28 FCR* (95% CI)	p- value	Day 180 FCR* (95% CI)	p- value			
Unadjusted									
	H1N1	IM	0.7 (0.2, 2.2)	0.55	0.8 (0.3, 2.5)	0.72			
	111111	MNP	0.9 (0.4, 2.0)	0.83	0.8 (0.4, 1.7)	0.58			
Prolonged	H3N2	IM	1.1 (0.6, 2.1) 1.1	0.72	1.5 (0.8, 2.9) 1.4	0.19			
(Yes vs. No)	H3N2	MNP	(0.6, 1.8)	0.82	1.4 (0.8, 2.3) 1.3	0.25			
	В	IM	$ \begin{array}{r} 1.1 \\ (0.5, 2.5) \\ 0.9 \end{array} $	0.72	(0.6, 2.8)	0.55			
		MNP	0.9 (0.5, 1.7)	0.82	1.0 (0.5, 1.8)	0.92			
		A	Adjusted^						
	H1N1	IM	0.9 (0.3, 2.8)	0.87	1.0 (0.3, 3.2)	0.93			
	111111	MNP	0.7 (0.4, 1.4)	0.33	0.6 (0.3, 1.2)	0.18			
Prolonged	H3N2	IM	1.2 (0.6, 2.2)	0.62	1.6 (0.9, 3.0)	0.14			
(Yes vs. No)	113112	MNP	0.9 (0.5, 1.5)	0.66	1.1 (0.7, 1.9)	0.58			
	В	IM	1.3 (0.6, 2.6)	0.48	1.4 (0.7, 2.9)	0.34			
	В	MNP	0.6 (0.4, 1.1)	0.08	0.6 (0.4, 1.1)	0.10			

Table 12. Reaction Duration and HAI Titer Fold Change by StudyDay

*Estimated fold change ratio for those in the prolonged category compared with those not in the prolonged category

Reaction		Study	Unadjusted RR*		Adjusted RR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
J 1			1.01	0.00	0.99	0.00
	11111	IM	(0.90, 1.12)	0.88	(0.89, 1.12)	0.92
	H1N1	MNP	1.03	0.55	1.02	0.66
		IVIINE	(0.94, 1.13)	0.55	(0.93, 1.12)	0.00
		IM	0.88	0.41	0.87	0.33
Global	H3N2	1101	(0.64, 1.20)	0111	(0.65, 1.15)	0.55
		MNP	0.94	0.58	0.93	0.47
			(0.74, 1.18) 1.02		(0.75, 1.14) 0.96	
		IM	(0.87, 1.19)	0.83	(0.81, 1.15)	0.67
	В		0.98		1.00	
		MNP	(0.80, 1.20)	0.83	(0.82, 1.22)	0.99
		П.	0.92	0.20	0.92	0.20
	H1N1	IM	(0.79, 1.07)	0.28	(0.80, 1.07)	0.28
	IIINI	MNP	1.01	0.92	0.99	0.86
		IVIINI	(0.86, 1.19)	0.72	(0.84, 1.15)	0.00
		IM	0.75	0.16	0.74	0.11
Systemic	H3N2		(0.49, 1.13)		(0.51, 1.07)	
-		MNP	MNP 1.16 0.11 0.11	(0.97, 1.33)	0.12	
			0.99		0.92	
	D	IM	(0.84, 1.18)	0.94	(0.78, 1.10)	0.37
	В		1.00	1.0	0.98	0.02
		MNP	(0.69, 1.46)	1.0	(0.65, 1.48)	0.92
		IM	0.93	0.50	0.91	0.32
	H1N1	1101	(0.76, 1.14)	0.50	(0.76, 1.09)	0.32
	111111	MNP	1.08	0.06	1.08	0.10
			(1.00, 1.18)	0.00	(0.98, 1.19)	0110
		IM	0.63	0.17	0.61	0.10
Local	H3N2		(0.33, 1.21) 0.84		(0.35, 1.09) 0.81	
		MNP	(0.66, 1.06)	0.15	(0.65, 1.02)	0.07
			0.99		0.95	
			(0.80, 1.24)	0.96	(0.73, 1.23)	0.70
	В		1.00	1.0	1.02	0.01
		MNP	(0.82, 1.22)	1.0	(0.84, 1.25)	0.81
TT A T 1	1	1 '1 '.' T	D. might motion CI. comfide	• . 1		、

Table 13. Longitudinal Associations Between Reaction Score and HAI Seroprotection

Antigon	Study	IM	MNP^	All
Antigen	Day	(n = 25)	(n = 48)	(n = 73)
	0	21 (84.0%)	35 (72.9%)	56 (76.7%)
H1N1	28	25 (100.0%)	48 (100.0%)	73 (100.0%)
	180	25 (100.0%)	48 (100.0%)	73 (100.0%)
	0	18 (72.0%)	29 (60.4%)	47 (64.4%)
H3N2	28	25 (100.0%)	47 (97.9%)	72 (98.6%)
	180	20 (80.0%)	39 (81.25%)	59 (80.8%)
	0	19 (76.0%)	23 (47.9%)	42 (57.5%)
В	28	25 (100.0%)	47 (97.9%)	72 (98.6%)
	180	23 (92.0%)	38 (79.2%)	61 (83.6%)

Table 14. HAI Seroprotection* by Vaccine Route and Day

*Number (%) of participants with hemagglutination inhibition antibody titer \geq 1:40 ^2 participants had missing outcome data

Reaction	A	Study	Unadjusted RR*		Adjusted RR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
		IM	1.01	0.79	0.99	0.79
	H1N1		(0.91, 1.12)		(0.88, 1.10)	
		MNP	1.05 (0.96, 1.16)	0.31	1.04 (0.94, 1.15)	0.47
			0.98	0.00	0.94	0.60
Global	H3N2	IM	(0.76, 1.26)	0.89	(0.73, 1.20)	0.60
$(\geq 3 vs \leq 2)$	II JINZ	MNP	0.96	0.70	0.95	0.57
		1711 11	(0.79, 1.17)	0.70	(0.79, 1.14)	0.57
		IM	1.09	0.30	0.99	0.87
	В		(0.92, 1.30)		(0.83, 1.17)	
		MNP	1.12 (0.90, 1.39)	0.30	1.15 (0.94, 1.41)	0.18
			0.98		0.98	
	111311	IM	(0.86, 1.13)	0.80	(0.85, 1.13)	0.77
	H1N1	MNP	1.01	0.85	1.01	0.89
		IVIINI	(0.91, 1.12)	0.85	(0.91, 1.12)	0.89
		IM	0.85	0.42	0.83	0.33
Global	H3N2		(0.56, 1.27)	0.12	(0.58, 1.21)	0.55
$(\geq 5 vs \leq 4)$	1151 (2	MNP	0.97	0.77	0.95	0.66
			(0.76, 1.22)		(0.77, 1.18)	
		IM	0.96	0.69	0.90	0.28
	В		(0.80, 1.16)		(0.75, 1.09)	
	_	MNP	1.00	1.0	1.02	0.86
	1	11 DD	(0.82, 1.22)	1.04	(0.82, 1.26)	

 Table 15A. Longitudinal Associations Between Alternative Cut Point Global Reaction

 Score and HAI Seroprotection

Reaction	A	Study	Unadjusted RR*		Adjusted RR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
		IM	1.02	0.67	1.01 (0.90, 1.13)	0.91
	H1N1	MNP	$ \begin{array}{r} (0.92, 1.14) \\ 1.00 \\ (0.91, 1.10) \end{array} $	0.98	$\frac{(0.90, 1.13)}{1.00}$ (0.91, 1.10)	0.99
Systemic	H3N2	IM	1.01 (0.78, 1.31)	0.92	0.97 (0.76, 1.24)	0.81
$(\geq 1 \text{ vs } 0)$	115112	MNP	1.03 (0.84, 1.25)	0.81	1.03 (0.86, 1.23)	0.78
	В	IM	1.12 (0.93, 1.34)	0.22	1.02 (0.85, 1.21)	0.86
		MNP	1.02 (0.83, 1.25)	0.86	1.05 (0.87, 1.28)	0.59
	H1N1	IM	1.00 (0.90, 1.10)	0.93	0.98 (0.88, 1.08)	0.68
	111111	MNP	1.06 (0.97, 1.16)	0.20	1.04 (0.95, 1.13)	0.37
Systemic	H3N2	IM	0.92 (0.72, 1.19)	0.54	$\begin{array}{c} 0.90 \\ (0.71, 1.14) \end{array}$	0.37
$(\geq 2 \text{ vs} \leq 1)$	1131\2	MNP	0.99 (0.80, 1.24)	0.95	0.97 (0.79, 1.19)	0.77
	В	IM	1.12 (0.96, 1.30)	0.16	1.04 (0.89, 1.21)	0.64
		MNP	1.08 (0.88, 1.34)	0.45	1.08 (0.88, 1.31)	0.47

 Table 15B. Longitudinal Associations Between Alternative Cut Point Systemic Reaction

 Score and HAI Seroprotection

Reaction	Autican	Study	Unadjusted RR*		Adjusted RR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
	H1N1	IM	1.05 (0.93, 1.18)	0.45	1.03 (0.91, 1.17)	0.63
	111111	MNP	1.06 (0.89, 1.25)	0.54	1.08 (0.92, 1.27)	0.32
Local	H3N2	IM	0.94 (0.74, 1.20)	0.63	0.91 (0.71, 1.17)	0.47
(≥ 1 vs 0)	113112	MNP	1.00 (0.73, 1.36)	0.99	1.00 (0.77, 1.29)	0.98
	В	IM	0.96 (0.83, 1.12)	0.62	0.86 (0.73, 1.01)	0.06
		MNP	1.14 (0.76, 1.71)	0.53	$ 1.16 \\ (0.83, 1.62) $	0.39
	H1N1	IM	0.98 (0.88, 1.09)	0.67	0.97 (0.87, 1.08)	0.57
		MNP	1.03 (0.93, 1.15)	0.53	1.05 (0.94, 1.17)	0.42
Local	H3N2	IM	0.99 (0.76, 1.28)	0.92	0.96 (0.75, 1.23)	0.74
$(\geq 2 vs \leq 1)$	115112	MNP	0.89 (0.74, 1.06)	0.18	0.89 (0.74, 1.07)	0.21
	В	IM	1.01 (0.87, 1.18)	0.87	0.92 (0.78, 1.09)	0.35
		MNP	1.13 (0.87, 1.47)	0.37	1.15 (0.88, 1.51)	0.31

Table 15C. Longitudinal Associations Between Alternative Cut Point Local Reaction Score and HAI Seroprotection

Reaction	Antigen	Study	Unadjusted RR*	p-value	Adjusted RR*^	p-value
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
		IM	1.05	0.86	1.05	0.85
	H1N1		(0.59, 1.87)	0.00	(0.62, 1.78)	0.00
		MNP	0.96	0.77	0.98	0.86
			(0.75, 1.24) 1.65		(0.78, 1.24)	
C1 1 1	110310	IM		0.05	,	0.07
Global	H3N2		(0.99, 2.76) 0.97	0.00	(0.96, 2.63) 0.97	0.05
		MNP	(0.65, 1.45)	0.89	(0.71, 1.33)	0.85
		IM	1.29	0.69	1.20	0.73
	В	1101	(0.37, 4.41) 1.53	0.07	(0.43, 3.31)	0.75
	D	MNP		0.07	1.38	0.15
			(0.96, 2.43)		(0.89, 2.14)	
		IM	1.10	0.76	1.06	0.84
	H1N1		(0.59, 2.05) 0.97		(0.59, 1.91) 1.04	
		MNP	(0.69, 1.37)	0.87		0.83
			1.69		(0.71, 1.52) 1.53	0.11
Genter	TIONO	IM	(1.08, 2.65)	0.02	(0.91, 2.58) 1.50	0.11
Systemic	H3N2	MNP	1.25	0.52	1.50	<0.01
		IVIINI	(0.64, 2.45)		(1.21, 1.85)	~0.01
		IM	2.26	0.18	1.83	0.22
	В	1101	(0.69, 7.39)	0.10	(0.70, 4.79) 1.26	0.22
		MNP	1.10	0.80		0.41
			(0.53, 2.30) 0.51		(0.72, 2.20) 0.52	
		IM		0.41	(0.52) (0.11, 2.53)	0.41
	H1N1		(0.10, 2.54) 1.02		1.00	
		MNP		0.87	(0.79, 1.26)	0.99
			(0.81, 1.29) 1.54	0.00	1.57	0.10
T 1		IM		0.09	(0.88, 2.80)	0.13
Local	H3N2	MNP	(0.94, 2.55) 1.16	0.40	1.06	0.70
		MINP	(0.82, 1.64) 1.47	0.40	(0.80, 1.40)	0.70
		IM		0.66	1.35	0.71
	В	1171	(0.26, 8.29)	0.00	(0.28, 6.55)	
		MNP	1.52	0.08	1.14	0.61
		1.11.1.1	(0.95, 2.43)		(0.69, 1.90)	0.01

 Table 16. Longitudinal Associations Between Reaction Score and HAI

 Seroconversion

	ci sion by	Day 28		Day 180	
Reaction	Study	RR*	p-	RR*	p-
Туре	Group	(95% CI)	value	(95% CI)	value
		Unadjus	ted	())/(01)	
	IM	0.9	0.56	1.5	0.38
Global	IM	(0.5, 1.4)	0.36	(0.6, 3.5)	0.38
Giobal	MNP	-	0.87	0.9	0.77
	1011 (1	(0.8, 1.2)	0.07	(0.5, 1.6)	0.77
	IM	0.8	0.44	1.8	0.15
Systemic		(0.4, 1.4)		(0.8, 4.1) 0.8	
	MNP	(1.0, 1.2)	0.08	(0.3, 2.2)	0.69
		0.4		0.7	
T 1	IM MNP		0.25		0.71
Local		(0.1, 1.9) 1.0	1.0	(0.1, 3.9) 1.1	0.83
	MINP	(0.9, 1.2)		(0.7, 1.7)	0.05
		Adjuste	d^		
	IM	0.9	0.59	1.5	0.33
Global		(0.5, 1.4)	0.57	(0.7, 3.3)	0.55
	MNP	1.0	0.98	0.9	0.81
		(0.8, 1.2)		(0.6, 1.5)	
	IM	0.8	0.41	1.8 (0.8, 3.9)	0.14
Systemic		(0.4, 1.4) 1.1		0.9	
	MNP	(0.9, 1.4)	0.18	(0.3, 2.4)	0.78
	IM	0.4	0.26	0.8	0.75
Local	IM	(0.1, 2.0)	0.26	(0.2, 3.9)	0.75
Local	MNP	1.0	0.84	1.0	0.89
	1011.01	(0.8, 1.2)	0.04	(0.7, 1.6)	0.07

Table 17A. Reaction Score and H1N1 HAISeroconversion by Study Day

*Estimated risk ratio for those in the higher score category compared with those in the lower score category

	ci sion og	Day 28		Day 180	
Reaction	Study		p-		p-
Туре	Group	RR*	value	RR*	value
	oronp	(95% CI)		(95% CI)	
		Unadjus	ted		
	IM	1.2	0.42	7.7	0.06
Global	11V1	(0.8, 1.8)	0.42	(1.0, 62.2) 1.4	0.00
Giobai	MAID	0.9	0.45	1.4	0.54
	MNP	(0.6, 1.3)	0.45	(0.5, 4.0) 3.2	0.54
	IM	1.5	0.01	3.2	0.19
Gratania	1101	(1.1, 2.0)	0.01	(0.6, 17.9)	0.19
Systemic	MNID	0.9	0.94	(0.6, 17.9) 2.4	0.12
	MNP	(0.5, 1.7)	0.84	(0.8, 7.6)	0.12
	IM	1.4	0.01	2.4	0.26
T a sal		(1.1, 1.8)	0.01	(0.4, 16.6)	0.30
Local	MNP	1	0.8	1.7	0.33
	MINP	(0.8, 1.4)		(0.6, 4.6)	
		Adjuste	d^		
	П.	1.1	0.50	7.3	0.00
C1-1-1	IM	(0.8, 1.7)	0.56	(0.9, 56.5)	0.06
Global		0.9	0.27	1.4	0.50
	MNP	(0.6, 1.2)	0.37	(0.5, 3.6)	0.52
	T) (1.3	0.11	2.8	0.04
G ()	IM	(0.9, 1.8)	0.11	(0.5, 16.5)	0.24
Systemic		1.1	0.57	2.9	-0.01
	MNP	(0.8, 1.6)	0.57	(1.4, 6.0)	<0.01
	1) (1.4	0.03	2.5	0.27
T 1	IM	(1.0, 1.9)	0.03		0.37
Local		0.9	0.71	(0.3, 17.8) 1.5	0.20
	MNP	(0.7, 1.3)	0.71	(0.6, 3.8)	0.38
				(l

Table 17B. Reaction Score and H3N2 HAISeroconversion by Study Day

*Estimated risk ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Study Group	Day 28 RR* (95% CI)	p- value	Day 180 RR* (95% CI)	p- value
		Unadjus	ted		
Global	IM	1.5 (0.5, 4.8)	0.45	0.9 (0.1, 6.9)	0.88
Giobal	MNP	1.3 (0.9, 2.0)	0.15	2.1 (0.9, 5.1)	0.11
Sustamia	IM	1.9 (0.6, 5.7)	0.25	3.2 (0.6, 17.9)	0.19
Systemic	MNP	1.2 (0.6, 2.2)	0.60	0.9 (0.2, 5.4)	0.92
Local	IM	1.0 (0.2, 5.8) 1.3	0.96	2.4 (0.4, 16.6) 2.3	0.36
Local	MNP	1.3 (0.8, 1.9)	0.26	2.3 (0.9, 5.8)	0.07
		Adjuste	d^		
Global	IM	1.3 (0.5, 3.3)	0.52	0.7 (0.1, 6.3)	0.79
Giobal	MNP	1.3 (0.8, 2.0)	0.32	2.0 (0.9, 4.4)	0.11
Systemic	IM	1.6 (0.7, 3.9)	0.26	2.7 (0.5, 15.1)	0.25
Systemic	MNP	1.3 (0.7, 2.7)	0.43	1.0 (0.3, 3.6)	0.96
т 1	IM	1.1 (0.2, 5.1)	0.92	2.5 (0.4, 15.6)	0.32
Local	MNP	$ \begin{array}{c} 1.0 \\ (0.6, 1.7) \end{array} $	0.96	1.9 (0.7, 4.7)	0.18

Table 17C. Reaction Score and B HAI Seroconversion by Study Day

HAI: hemagglutination inhibition; RR: risk ratio; CI: confidence

interval; IM: intramuscular; MNP: microneedle patch; *Estimated risk ratio for those in the higher score category compared with those in the lower score category

Reaction	Aution	Study	Unadjusted RR*		Adjusted RR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
		IM	1.09	0.73	1.21	0.43
	H1N1		(0.67, 1.76)		(0.76, 1.93)	0.16
		MNP	0.84 (0.68, 1.04)	0.11	0.86 (0.69, 1.06)	
		IM	1.02	0.04	0.98	0.05
Global	H3N2	IM	(0.61, 1.72)	0.94	(0.59, 1.64)	0.95
$(\geq 3 \text{ vs} \leq 2)$	113112	MNP	0.88	0.46	0.87	0.36
		1011 (1	(0.62, 1.24)	0.10	(0.64, 1.18)	0.50
		IM	1.57	0.49	1.61	0.40
	В		(0.43, 5.69)		(0.53, 4.88)	
		MNP	0.94 (0.58, 1.52)	0.80	0.84 (0.53, 1.32)	
		0.96		0.97		
	H1N1	IM	(0.45, 2.05)	0.92	(0.47, 1.98)	0.93
	HINI	MNP	1.04	0.79	1.02	0.87
		IVIINI	(0.80, 1.35)	0.79	(0.79, 1.31)	0.07
		IM	1.75	0.01	1.63	0.09
Global	H3N2	1101	(1.12, 2.74)	0.01	(0.93, 2.86)	0.09
$(\geq 5 \text{ vs} \leq 4)$	1151(2	MNP	1.08	0.69	1.04	0.78
			(0.73, 1.60)		(0.77, 1.41)	
		IM	1.33	0.68	1.21	0.74
	В		(0.34, 5.26)		(0.38, 3.81)	
		MNP	1.71	0.02	1.42	0.13
			(1.10, 2.68)		(0.91, 2.22)	

 Table 18A. Longitudinal Associations Between Alternative Cut Point Global Reaction

 Score and HAI Seroconversion

Reaction	Antion	Study	Unadjusted RR*		Adjusted RR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
		IM	1.06	0.83	1.15	0.57
	H1N1	1101	(0.64, 1.73)	0.05	(0.71, 1.87)	0.57
		MNP	0.94	0.60	0.95	0.60
			(0.75, 1.18)		(0.77, 1.16)	
Sustamia		IM	1.04	0.89	1.02	0.93
Systemic $(\geq 1 \text{ vs } 0)$	H3N2		(0.61, 1.75) 1.04		(0.61, 1.71) 1.03	
$(\geq 1 \vee S \circ)$		MNP	(0.73, 1.47)	0.83	(0.77, 1.37)	0.84
		n (2.00	0.00	2.16	0.25 0.47
	р	IM	(0.45, 8.83)	0.36	(0.58, 8.05)	
	В	MNP	0.93	0.76	0.86	
		101101	(0.57, 1.50)	0.70	(0.58, 1.29)	0.47
		IM	0.89	0.65	0.95	0.81
	H1N1	1101	(0.55, 1.46)	0.05	(0.60, 1.50)	0.01
		MNP	0.93	0.60	1.04	0.79
			(0.71, 1.22)		(0.80, 1.35)	
		IM	0.99	0.98	1.06	0.81
Systemic $(> 2 + 1)$	H3N2		(0.58, 1.71)		(0.65, 1.74) 0.98	
$(\geq 2 \text{ vs} \leq 1)$		MNP	0.87 (0.58, 1.32)	0.52	(0.71, 1.35)	0.90
			1.52		1.8	
		IM	(0.42, 5.44)	0.52	(0.62, 5.23)	0.28
	В		1.01		1.38	
		MNP	(0.60, 1.70)	0.97	(0.87, 2.20)	0.17
TTAT 1	4. 4. 1	11 DD	i 1 d' CI d' 1	· · · · · · 1. TM		

 Table 18B. Longitudinal Associations Between Alternative Cut Point Systemic Reaction

 Score and HAI Seroconversion

Reaction	Aution	Study	Unadjusted RR*		Adjusted RR*^	
Туре	Antigen	Group	(95% CI)	p-value	(95% CI)	p-value
		IM	0.75	0.17	0.74	0.15
	H1N1		(0.49, 1.14) 0.96		(0.50, 1.11) 0.87	
		MNP	(0.61, 1.51)	0.86	(0.56, 1.34)	0.52
		IM	0.88	0.63	1.02	0.95
Local	H3N2	1101	(0.53, 1.46)	0.05	(0.65, 1.60)	0.75
$(\geq 1 \text{ vs } 0)$	1101 (2	MNP	1.02	0.94	0.93	
			(0.57, 1.83)		(0.61, 1.44)	
		IM	0.94 (0.27, 3.30)	0.92	1.01 (0.37, 2.77)	0.98
	В		0.74		0.42	<0.01
		MNP	(0.49, 1.10)	0.14	(0.24, 0.72)	
		IM	1.24	0.39	1.28	0.20
	H1N1	11V1	(0.77, 1.99)	0.39	(0.81, 2.03)	0.50
	111111	MNP	1.07	0.61	0.98	0.88
		1011 (1	(0.82, 1.41)	0.01	(0.77, 1.25)	0.00
		IM	1.64	0.05	1.54	0.07
Local	H3N2		(0.99, 2.71)		(0.97, 2.46)	
$(\geq 2 \text{ vs} \leq 1)$		MNP	0.94	0.72	0.87	0.35
			(0.66, 1.33)		(0.64, 1.17)	
		IM	2.10 (0.63, 7.01)	0.23	1.75 (0.65, 4.69)	0.27
	В		1.36		0.81	
		MNP	(0.83, 2.23)	0.22	(0.81) $(0.47, 1.40)$	0.44
TTAT. 1		11.1. DD	(0.05, 2.25)	1. TN /		

 Table 18C. Longitudinal Associations Between Alternative Cut Point Local Reaction

 Score and HAI Seroconversion

Seroconversion by Study Day									
Reaction		Study	Day 28	n	Day 180	n			
	Antigen	Study	RR*	p-	RR*	p-			
Туре	-8	Group	(95% CI)	value	(95% CI)	value			
		T	Jnadjusted		())/(001)				
		IM	1.0	0.84	1.4	0.51			
	H1N1		(0.6, 1.4)		(0.5, 3.5)				
		MNP	1.0	0.76	0.7	0.08			
		1011 01	(0.8, 1.1)	0.70	(0.4, 1.0)	0.00			
		IM	0.9	0.54	2.4	0.43			
Global	TTONIO	1101	(0.6, 1.3)	0.54	(0.3, 19.7)	0.45			
$(\geq 3 \text{ vs} \leq 2)$	H3N2		0.9	0.20	0.9	0.77			
``´´´		MNP	(0.7, 1.2)	0.38	(0.3, 2.4)	0.77			
			1.3		2.4				
		IM		0.66		0.43			
	В		(0.4, 4.3) 0.9		(0.3, 19.7) 1.1				
		MNP		0.50	(0.4, 3.0)	0.78			
	l		(0.6, 1.3)		(0.4, 5.0)				
		1	Adjusted^		1.5				
		IM	1.1	0.74	1.5	0.34			
	H1N1		(0.7, 1.7)		(0.6, 3.7)				
		MNP	1.0	0.87	0.7	0.07			
		1011 01	(0.8, 1.2)	0.07	(0.4, 1.0)	0.07			
	112112	IM	0.8	0.43	2.3	0.44			
Global			(0.5, 1.3)		(0.3, 18.5)	0.44			
$(\geq 3 \text{ vs} \leq 2)$	H3N2		0.9	0.27	0.8	0.74			
, , ,		MNP	(0.6, 1.2)	0.37	(0.3, 2.2)	0.74			
			1.4	0.49	2.6				
		IM			(0.3, 21.5)	0.38			
	В		(0.5, 4.1) 0.8		1.0				
		MNP	(0.5, 1.3)	0.33	(0.4, 2.5)	0.92			
		T	Jnadjusted		(0.4, 2.3)				
			0.7		1.5				
		IM		0.36		0.37			
	H1N1		(0.3, 1.5)		(0.6, 3.7)				
		MNP		0.76	1.1	0.59			
			(0.8, 1.2)		(0.7, 1.9)				
~ 1 1 1		IM	1.4	0.01	4.0	0.11			
Global	H3N2		(1.1, 1.9)		(0.7, 21.8)				
$(\geq 5 vs \leq 4)$	1101.2	MNP	0.9	0.70	1.7	0.31			
		11111	(0.6, 1.3)	0.70	(0.6, 4.9)	0.51			
		IM	1.3	0.66	1.3	0.78			
	р	11VI	(0.4, 4.7)	0.00	(0.2, 10.3)	0.70			
	В		1.4	0.07	2.6	0.02			
		MNP	(1.0, 2.1)	0.06	(1.1, 6.2)	0.03			
Adjusted^									
			0.7		1.5				
Global		IM	(0.4, 1.5)	0.38	(0.7, 3.6)	0.31			
$(\geq 5 \text{ vs} \leq 4)$	H1N1		1.0		1.1				
		MNP	(0.8, 1.2)	0.72	(0.7, 1.8)	0.60			
	1		(0.0, 1.2)		(0.7, 1.8)				

Table 19A. Alternative Cut Point Global Reaction Score and HAISeroconversion by Study Day

	H3N2	IM	1.3 (0.9, 1.8)	0.11	3.7 (0.6, 21.2)	0.14
		MNP	0.9 (0.6, 1.2)	0.49	1.6 (0.6, 4.2)	0.31
	В	IM	1.2 (0.4, 3.3)	0.72	1.2 (0.2, 9.3)	0.86
		MNP	1.2 (0.8, 2.0)	0.34	2.2 (1.0, 5.1)	0.05

*Estimated risk ratio for those in the higher score category compared with those in the lower score category

Reaction Type	Antigen	Study Group	Day 28 RR* (95% CI)	p- value	Day 180 RR* (95% CI)	p- value
		l	Jnadjusted			
	11111	IM	1.0 (0.7, 1.5)	1.0	1.2 (0.5, 3.0) 0.9	0.75
	H1N1	MNP	1.0 (0.8, 1.1) 0.9	0.65	$ \begin{array}{r} 0.9 \\ (0.6, 1.4) \\ 2 \end{array} $	0.67
Systemic	H3N2	IM	(0.6, 1.4)	0.69	2 (0.2, 16.6) 1.5	0.52
$(\geq 1 \text{ vs } 0)$	115112	MNP	0.9 (0.7, 1.3)	0.67	$ \begin{array}{r} 1.5 \\ (0.5, 4.4) \\ 2.0 \end{array} $	0.48
	В	IM	2.0 (0.5, 8.0)	0.33	(0.2, 16.6)	0.52
		MNP	0.9 (0.6, 1.4)	0.63	1.0 (0.4, 2.5)	0.98
		1	Adjusted^	F		[
	H1N1	IM	1.1 (0.7, 1.7) 1.0	0.69	$ \begin{array}{r} 1.3 \\ (0.5, 3.1) \\ 0.9 \end{array} $	0.59
		MNP	1.0 (0.8, 1.1) 0.9	0.71	0.9 (0.6, 1.4) 2.0	0.66
Systemic	H3N2	IM	(0.6, 1.4)	0.66	2.0 (0.2, 16.0) 1.5	0.52
(≥1 vs 0)		MNP	0.9 (0.7, 1.2)	0.63	(0.5, 3.9)	0.45
	В	IM	2.2 (0.6, 7.6)	0.23	2.2 (0.3, 17.7)	0.48
		MNP	0.9 (0.6, 1.3)	0.44	0.9 (0.4, 2.1)	0.86
	1	l	Jnadjusted	r		r
	H1N1	IM	0.9 (0.6, 1.3)	0.56	0.9 (0.4, 2.2)	0.82
	HINI	MNP	1.0 (0.8, 1.2)	0.81	0.9 (0.5, 1.5)	0.59
Systemic	H3N2	IM	0.8 (0.5, 1.3)	0.31	3.2 (0.4, 27.2)	0.28
$(\geq 2 \text{ vs} \leq 1)$		MNP	1.0 (0.7, 1.4)	0.82	0.6 (0.1, 2.4)	0.47
	В	IM	1.8 (0.5, 6.0)	0.33	$ 1.1 \\ (0.2, 6.5) $	0.93
		MNP	1.1 (0.7, 1.7)	0.67	0.8 (0.3, 2.5)	0.71
	1	1	Adjusted^			
Systemic	H1N1	IM	0.9 (0.6, 1.4)	0.78	1.0 (0.4, 2.2)	0.92
$(\geq 2 vs \leq 1)$	HINI	MNP	1.1 (0.9, 1.3)	0.42	0.9 (0.5, 1.7)	0.86

Table 19B. Alternative Cut Point Systemic Reaction Score and HAISeroconversion by Study Day

	H3N2	IM	0.8 (0.5, 1.3)	0.42	3.5 (0.4, 27.5)	0.24
		MNP	1.1 (0.8, 1.5)	0.63	0.7 (0.2, 2.4)	0.54
	В	IM	2.0 (0.7, 5.5)	0.18	1.2 (0.2, 6.9)	0.84
		MNP	1.5 (0.9, 2.3)	0.12	1.1 (0.4, 3.0)	0.89

*Estimated risk ratio for those in the higher score category compared with those in the lower score category

D 20 D 100									
Reaction		Study	Day 28	p-	Day 180	p-			
Туре	Antigen	Group	RR*	value	RR*	value			
Type		Oroup	(95% CI)	value	(95% CI)	value			
		U	Inadjusted						
			0.7		0.8	0.67			
		IM	(0.5, 1.0)	0.03	(0.3, 2.0)	0.67			
	H1N1		1.2		0.7				
		MNP	(0.8, 1.9)	0.44		0.22			
			0.8		(0.4, 1.2) 1.4				
Local		IM	(0.5, 1.2)	0.30		0.75			
	H3N2				(0.2, 11.5) 1.2				
$(\geq 1 \text{ vs } 0)$		MNP	1.0	0.96		0.87			
			(0.6, 1.6)		(0.2, 7.3)				
		IM	0.8	0.68	1.4	0.75			
	В		(0.2, 2.5)		(0.2, 11.5)				
		MNP	0.6	<0.01		0.72			
			(0.5, 0.8)		(0.2, 8.6)				
	1	1	Adjusted^		1				
		IM	0.7	0.06	0.8	0.66			
	H1N1	1111	(0.5, 1.0)	0.00	(0.4, 1.9) 0.6	0.00			
	1111111	MND	1.1	0.79	0.6	0.08			
		MNP	(0.7, 1.6)	0.78	(0.4, 1.1)	0.08			
	112212	D (0.9	0.70	(0.4, 1.1) 1.6	0.64			
Local $(\geq 1 \text{ vs } 0)$		IM	(0.6, 1.4)	0.70	(0.2, 12.8)	0.64			
	H3N2	MNP	0.9	0.55	1.1				
()			(0.6, 1.4)	0.66	(0.2, 5.4)	0.94			
		IM	0.9	0.86	1.7				
			(0.4, 2.3)			0.64			
	В		0.4		(0.2, 13.9) 0.8				
		MNP	(0.2, 0.6)	<0.01	(0.1, 5.8)	0.86			
		T	Jnadjusted		(0.1, 5.0)				
			ř.		1.0				
		IM	1.0	1.0	1.8	0.19			
	H1N1		(0.7, 1.5)		(0.7, 4.3)				
	111111	MNP	1.1	0.29	1.0	0.97			
			(0.9, 1.4)		(0.6, 1.7)				
T 1		IM	1.3	0.15	4.5	0.16			
Local	H3N2		(0.9, 2.0)		(0.5, 37.4)				
$(\geq 2 \text{ vs} \leq 1)$	1101.2	MNP	0.9	0.34	1.2	0.75			
		1011 (1	(0.7, 1.2)	0.51	(0.4, 3.9)	0.75			
		IM	1.5	0.48	4.5	0.16			
	В	1111	(0.5, 4.7)	0.40	(0.5, 37.4)	0.10			
	Б	MND	1.0	0.84	5.5	0.09			
		MNP	(0.6, 1.5)	0.04	(0.8, 38.2)	0.09			
		1	Adjusted^						
		T) 4	1.0	0.02	1.9	0.12			
Local	1112.14	IM	(0.7, 1.6)	0.83	(0.8, 4.3)	0.13			
$(\geq 2 \text{ vs} \leq 1)$	H1N1		1.0	0.01	0.9	o - -			
		MNP	(0.8, 1.2)	0.81	(0.6, 1.5)	0.75			
L	1		(0.0, 1.2)		(0.0, 1.5)				

 Table 19C. Alternative Cut Point Local Reaction Score and HAI

 Seroconversion by Study Day

	H3N2	IM	1.3 (0.9, 1.9)	0.24	4.2 (0.5, 33.3)	0.17
		MNP	0.8 (0.6, 1.1)	0.11	1.1 (0.4, 3.4)	0.84
	В	IM	1.4 (0.6, 3.4)	0.45	4.2 (0.5, 35.8)	0.19
		MNP	0.7 (0.4, 1.1)	0.13	3.8 (0.5, 28.1)	0.20

*Estimated risk ratio for those in the higher score category compared with those in the lower score category

Figures



Figure 1. Conceptual Model. Directed acyclic graph (DAG) of the relationship between reactogenicity (green, exposure) and immunogenicity (blue, outcome) with confounding variables (salmon).



Figure 2. Parent Study CONSORT Diagram. CONSORT (flow) diagram from TIV-MNP 2015⁵⁴.



Figure 3. Adverse Events by Vaccine Route. Fraction of participants in the intramuscular (IM, red) and microneedle patch (MNP, teal) group who developed each type of adverse event.


Figure 4. Select Local Adverse Events by Time and Vaccine Route. Redness (A) and swelling (B) diameter in millimeters over time after vaccination for those in the intramuscular (IM, red) and microneedle patch (MNP, teal) groups.

Nausea														1
0.40	Malaise													- 0.8
0.20	0.40	Myalgia												- 0.6
0.01	0.41	0.28	Fatigue											
0.25	0.22	0.36	0.44	Sweating										- 0.4
0.08	0.07	0.03	0.02	-0.10	Arthralgia									- 0.2
0.10	0.26	0.00	0.20	-0.14	0.09	Headache								
-0.01	0.11	0.18	0.22	0.13	0.09	0.18	Tenderness							- 0
0.04	0.16	-0.02	0.24	0.14	0.10	0.23	0.24	Swelling						0.2
0.07	0.14	0.21	0.09	0.10	0.19	0.18	0.34	0.41	Pain					0.4
0.37	0.55	0.28	0.27	0.32	-0.05	0.04	0.01	0.40	0.18	Shivering				
-0.04	0.31	-0.05	0.15	-0.05	-0.03	0.20	0.09	0.36	0.19	0.57	Fever			0.6
-0.21	0.06	-0.18	0.11	-0.06	0.08	0.13	0.14	0.43	0.09	0.03	0.19	Redness		0.8
-0.07	-0.03	-0.16	0.05	-0.08	0.00	0.10	0.24	0.26	0.02	0.03	0.09	0.43	Pruritus	

Figure 5. Correlation Between Adverse Events. Correlation plot of all solicited adverse event types.



Figure 6. Reaction Score Univariable Distributions. Histograms of the Global (A), Systemic (B), and Local (C) reaction scores. Vertical red lines depict the dichotomization cut points chosen for the primary analyses.



Figure 7. Reaction Score Bivariable Distributions. Scatterplots of the Global and Systemic (A), Local and Systemic (B), and Global and Local (C) reaction scores. Red lines depict the dichotomization cut points chosen for the primary analyses.



Figure 8. H3N2 HAI Titer by Time and Reaction Score. Estimated adjusted geometric mean titer against the H3N2 antigen of intramuscular seasonal inactivated influenza vaccine by study day, comparing low and high score levels for systemic reactogenicity (A) and local reactogenicity (B). * p = 0.01.

References

- 1. Castrucci MR. Factors affecting immune responses to the influenza vaccine. *Hum Vaccines Immunother*. 2018;14(3):637-646. doi:10.1080/21645515.2017.1338547
- Pollard AJ, Bijker EM. A guide to vaccinology: from basic principles to new developments. Nat Rev Immunol. 2021;21(2):83-100. doi:10.1038/s41577-020-00479-7
- Cortese M, Sherman AC, Rouphael NG, Pulendran B. Systems Biological Analysis of Immune Response to Influenza Vaccination. *Cold Spring Harb Perspect Med.* 2021;11(6):a038596. doi:10.1101/cshperspect.a038596
- 4. Couch RB. Seasonal inactivated influenza virus vaccines. *Vaccine*. 2008;26 Suppl 4(Suppl 4):D5-9. doi:10.1016/j.vaccine.2008.05.076
- Denly L. The effect of sex on responses to influenza vaccines. *Hum Vaccines Immunother*. 2021;17(5):1396-1402. doi:10.1080/21645515.2020.1830685
- Harper A, Flanagan KL. Effect of sex on vaccination outcomes: important but frequently overlooked. *Curr Opin Pharmacol*. 2018;41:122-127. doi:10.1016/j.coph.2018.05.009
- Lin CJ, Martin JM, Cole KS, et al. Are children's vitamin D levels and BMI associated with antibody titers produced in response to 2014–2015 influenza vaccine? *Hum Vaccines Immunother*. 2017;13(7):1661-1665. doi:10.1080/21645515.2017.1299837
- Clarke M, Goodchild LM, Evans S, et al. Body mass index and vaccine responses following influenza vaccination during pregnancy. *Vaccine*. 2021;39(34):4864-4870. doi:10.1016/j.vaccine.2021.06.065

- Kainth MK, Fishbein JS, Aydillo T, et al. Obesity and Metabolic Dysregulation in Children Provide Protective Influenza Vaccine Responses. *Viruses*. 2022;14(1):124. doi:10.3390/v14010124
- Tagliabue C, Principi N, Giavoli C, Esposito S. Obesity: impact of infections and response to vaccines. *Eur J Clin Microbiol Infect Dis*. 2016;35(3):325-331. doi:10.1007/s10096-015-2558-8
- Talbot HK, Coleman LA, Crimin K, et al. Association between obesity and vulnerability and serologic response to influenza vaccination in older adults. *Vaccine*. 2012;30(26):3937-3943. doi:10.1016/j.vaccine.2012.03.071
- Sheridan PA, Paich HA, Handy J, et al. Obesity is associated with impaired immune response to influenza vaccination in humans. *Int J Obes*. 2012;36(8):1072-1077. doi:10.1038/ijo.2011.208
- Belongia EA, Skowronski DM, Mclean HQ, Chambers C, Sundaram ME, De Serres G.
 Repeated annual influenza vaccination and vaccine effectiveness: review of evidence. *Expert Rev Vaccines*. 2017;16(7):723-736. doi:10.1080/14760584.2017.1334554
- Sherman AC, Lai L, Bower M, et al. The Effects of Imprinting and Repeated Seasonal Influenza Vaccination on Adaptive Immunity after Influenza Vaccination. *Vaccines*. 2020;8(4):663. doi:10.3390/vaccines8040663
- O'Shea J, Prausnitz MR, Rouphael N. Dissolvable Microneedle Patches to Enable Increased Access to Vaccines against SARS-CoV-2 and Future Pandemic Outbreaks. *Vaccines*. 2021;9(4):320. doi:10.3390/vaccines9040320

- Arya J, Prausnitz MR. Microneedle patches for vaccination in developing countries. J Control Release. 2016;240:135-141. doi:10.1016/j.jconrel.2015.11.019
- 17. Reisinger KS, Holmes SJ, Pedotti P, Arora AK, Lattanzi M. A dose-ranging study of MF59(®)-adjuvanted and non-adjuvanted A/H1N1 pandemic influenza vaccine in young to middle-aged and older adult populations to assess safety, immunogenicity, and antibody persistence one year after vaccination. *Hum Vaccines Immunother*. 2014;10(8):2395-2407. doi:10.4161/hv.29393
- Goubau P, Van Gerven V, Safary A, et al. Effect of virus strain and antigen dose on immunogenicity and reactogenicity of an inactivated hepatitis A vaccine. *Vaccine*. 1992;10 Suppl 1:S114-118. doi:10.1016/0264-410x(92)90561-w
- Domnich A, Orsi A, Trombetta CS, Guarona G, Panatto D, Icardi G. COVID-19 and Seasonal Influenza Vaccination: Cross-Protection, Co-Administration, Combination Vaccines, and Hesitancy. *Pharm Basel Switz*. 2022;15(3):322. doi:10.3390/ph15030322
- Giambi C, Fabiani M, D'Ancona F, et al. Parental vaccine hesitancy in Italy Results from a national survey. *Vaccine*. 2018;36(6):779-787. doi:10.1016/j.vaccine.2017.12.074
- 21. Cook IF. Subcutaneous vaccine administration an outmoded practice. *Hum Vaccines Immunother*. 2021;17(5):1329-1341. doi:10.1080/21645515.2020.1814094
- 22. Gowda C, Dempsey AF. The rise (and fall?) of parental vaccine hesitancy. *Hum Vaccines Immunother*. 2013;9(8):1755-1762. doi:10.4161/hv.25085

- 23. Salmon DA, Moulton LH, Omer SB, DeHart MP, Stokley S, Halsey NA. Factors associated with refusal of childhood vaccines among parents of school-aged children: a case-control study. *Arch Pediatr Adolesc Med.* 2005;159(5):470-476. doi:10.1001/archpedi.159.5.470
- Siddiqui M, Salmon DA, Omer SB. Epidemiology of vaccine hesitancy in the United States. *Hum Vaccines Immunother*. 2013;9(12):2643-2648. doi:10.4161/hv.27243
- Schmid P, Rauber D, Betsch C, Lidolt G, Denker ML. Barriers of Influenza Vaccination Intention and Behavior - A Systematic Review of Influenza Vaccine Hesitancy, 2005 - 2016. *PloS One*. 2017;12(1):e0170550. doi:10.1371/journal.pone.0170550
- 26. Cascini F, Pantovic A, Al-Ajlouni Y, Failla G, Ricciardi W. Attitudes, acceptance and hesitancy among the general population worldwide to receive the COVID-19 vaccines and their contributing factors: A systematic review. *EClinicalMedicine*. 2021;40:101113. doi:10.1016/j.eclinm.2021.101113
- Cowling BJ, Thompson MG, Ng TWY, et al. Comparative Reactogenicity of Enhanced Influenza Vaccines in Older Adults. *J Infect Dis*. 2020;222(8):1383-1391. doi:10.1093/infdis/jiaa255
- Saleh E, Moody MA, Walter EB. Effect of antipyretic analgesics on immune responses to vaccination. *Hum Vaccines Immunother*. 2016;12(9):2391-2402. doi:10.1080/21645515.2016.1183077
- 29. What COVID vaccine side effects can and can't tell you about your body's immune response. PBS NewsHour. Published April 20, 2021. Accessed January 11, 2024.

https://www.pbs.org/newshour/health/what-covid-vaccine-side-effects-can-and-cant-tellyou-about-your-bodys-immune-response

- 30. Why do some people get side effects after COVID-19 vaccines? Yahoo Entertainment. Published June 10, 2021. Accessed January 7, 2024. https://www.yahoo.com/entertainment/why-people-side-effects-covid-070229275.html
- Did My Strong COVID Vax Reaction Give Me Better Immunity? Published November 1, 2022. Accessed January 11, 2024. https://www.medpagetoday.com/specialreports/exclusives/101522
- 32. Siangphoe U, Baden LR, El Sahly HM, et al. Associations of Immunogenicity and Reactogenicity After Severe Acute Respiratory Syndrome Coronavirus 2 mRNA-1273 Vaccine in the COVE and TeenCOVE Trials. *Clin Infect Dis Off Publ Infect Dis Soc Am*. 2023;76(2):271-280. doi:10.1093/cid/ciac780
- 33. Debes AK, Xiao S, Colantuoni E, et al. Association of Vaccine Type and Prior SARS-CoV-2 Infection With Symptoms and Antibody Measurements Following Vaccination Among Health Care Workers. *JAMA Intern Med.* 2021;181(12):1660-1662. doi:10.1001/jamainternmed.2021.4580
- 34. Michos A, Tatsi EB, Filippatos F, et al. Association of total and neutralizing SARS-CoV-2 spike -receptor binding domain antibodies with epidemiological and clinical characteristics after immunization with the 1st and 2nd doses of the BNT162b2 vaccine. *Vaccine*. 2021;39(40):5963-5967. doi:10.1016/j.vaccine.2021.07.067

- 35. Yoshida M, Kobashi Y, Kawamura T, et al. Association of systemic adverse reaction patterns with long-term dynamics of humoral and cellular immunity after coronavirus disease 2019 third vaccination. *Sci Rep.* 2023;13(1):9264. doi:10.1038/s41598-023-36429-1
- 36. Bannister WP, Raben D, Valentiner-Branth P, et al. Association of Self-reported Systemic Reactions Following SARS-CoV-2 Vaccination With Immunological Response in the Danish National Cohort Study of Effectiveness and Safety of SARS-CoV-2 Vaccines (ENFORCE). Open Forum Infect Dis. 2023;10(6):ofad248. doi:10.1093/ofid/ofad248
- 37. Braun E, Horowitz NA, Leiba R, et al. Association between IgG antibody levels and adverse events after first and second Bnt162b2 mRNA vaccine doses. *Clin Microbiol Infect Off Publ Eur Soc Clin Microbiol Infect Dis*. 2022;28(12):1644-1648. doi:10.1016/j.cmi.2022.07.002
- 38. Romero-Ibarguengoitia ME, González-Cantú A, Pozzi C, et al. Analysis of immunization time, amplitude, and adverse events of seven different vaccines against SARS-CoV-2 across four different countries. *Front Immunol.* 2022;13:894277. doi:10.3389/fimmu.2022.894277
- Romero-Ibarguengoitia ME, González-Cantú A, Rivera-Salinas D, et al. Analysis of Immunization, Adverse Events, and Efficacy of a Fourth Dose of BNT162b2 Vaccine in Health Workers in Mexico, a Pilot Study. *Vaccines*. 2022;10(7):1139. doi:10.3390/vaccines10071139
- 40. Yamamoto S, Fukunaga A, Tanaka A, et al. Association between reactogenicity and SARS-CoV-2 antibodies after the second dose of the BNT162b2 COVID-19 vaccine. *Vaccine*. 2022;40(13):1924-1927. doi:10.1016/j.vaccine.2022.02.052

- 41. Izak M, Stoyanov E, Dezuraev K, Shinar E. Correlation of Anti-SARS-CoV-2 S1-specificIgG antibody levels and adverse events following vaccination with BNT162b2 mRNA COVID-19 vaccine in healthcare workers. *Vaccine*. 2022;40(3):428-431. doi:10.1016/j.vaccine.2021.11.082
- 42. Cheng A, Hsieh MJ, Chang SY, et al. Correlation of adverse effects and antibody responses following homologous and heterologous COVID19 prime-boost vaccinations. *J Formos Med Assoc Taiwan Yi Zhi*. Published online December 15, 2022:S0929-6646(22)00441-7. doi:10.1016/j.jfma.2022.12.002
- 43. Tani N, Ikematsu H, Goto T, et al. Correlation of post-vaccination fever with specific antibody response to SARS-CoV-2 BNT162b2 booster and no significant influence of antipyretic medication. Published online 2022. doi:10.1101/2022.07.25.22277569
- 44. Tani N, Chong Y, Kurata Y, et al. Relation of fever intensity and antipyretic use with specific antibody response after two doses of the BNT162b2 mRNA vaccine. *Vaccine*. 2022;40(13):2062-2067. doi:10.1016/j.vaccine.2022.02.025
- 45. Heo JY, Seo YB, Kim EJ, et al. COVID-19 vaccine type-dependent differences in immunogenicity and inflammatory response: BNT162b2 and ChAdOx1 nCoV-19. *Front Immunol.* 2022;13:975363. doi:10.3389/fimmu.2022.975363
- 46. Takeuchi M, Higa Y, Esaki A, Nabeshima Y, Nakazono A. Does reactogenicity after a second injection of the BNT162b2 vaccine predict spike IgG antibody levels in healthy Japanese subjects? *PloS One*. 2021;16(9):e0257668. doi:10.1371/journal.pone.0257668

- 47. Levy I, Levin EG, Olmer L, et al. Correlation between Adverse Events and Antibody Titers among Healthcare Workers Vaccinated with BNT162b2 mRNA COVID-19 Vaccine. *Vaccines*. 2022;10(8):1220. doi:10.3390/vaccines10081220
- Lin TY, Hung NK, Hung SC. Association of Reactogenicity with Immunogenicity of the ChAdOx1 nCoV-19 Vaccine in Patients Undergoing Hemodialysis. *Vaccines*. 2022;10(8):1366. doi:10.3390/vaccines10081366
- 49. Daulagala P, Mann BR, Leung K, et al. Imprinted Anti-Hemagglutinin and Anti-Neuraminidase Antibody Responses after Childhood Infections of A(H1N1) and A(H1N1)pdm09 Influenza Viruses. *mBio*. 2023;14(3):e00084-23. doi:10.1128/mbio.00084-23
- 50. Perofsky AC, Huddleston J, Hansen C, et al. Antigenic drift and subtype interference shape A(H3N2) epidemic dynamics in the United States. *MedRxiv Prepr Serv Health Sci*.
 Published online October 3, 2023:2023.10.02.23296453. doi:10.1101/2023.10.02.23296453
- Tripp RA. Understanding immunity to influenza: implications for future vaccine development. *Expert Rev Vaccines*. 2023;22(1):871-875. doi:10.1080/14760584.2023.2266033
- 52. Carregaro RL, Roscani ANCP, Raimundo ACS, et al. Immunogenicity and safety of inactivated quadrivalent influenza vaccine compared with the trivalent vaccine for influenza infection: an overview of systematic reviews. *BMC Infect Dis.* 2023;23(1):563. doi:10.1186/s12879-023-08541-0

- Grohskopf LA. Prevention and Control of Seasonal Influenza with Vaccines: Recommendations of the Advisory Committee on Immunization Practices — United States, 2023–24 Influenza Season. *MMWR Recomm Rep.* 2023;72. doi:10.15585/mmwr.rr7202a1
- Rouphael NG, Paine M, Mosley R, et al. The safety, immunogenicity, and acceptability of inactivated influenza vaccine delivered by microneedle patch (TIV-MNP 2015): a randomised, partly blinded, placebo-controlled, phase 1 trial. *Lancet*. 2017;390(10095):649-658. doi:10.1016/s0140-6736(17)30575-5
- 55. Wilder-Smith A, Osman S. Public health emergencies of international concern: a historic overview. J Travel Med. 2020;27(8):taaa227. doi:10.1093/jtm/taaa227
- 56. Roychoudhury S, Das A, Sengupta P, et al. Viral Pandemics of the Last Four Decades: Pathophysiology, Health Impacts and Perspectives. *Int J Environ Res Public Health*. 2020;17(24):9411. doi:10.3390/ijerph17249411
- 57. Bourrier MS, Deml MJ. The Legacy of the Pandemic Preparedness Regime: An Integrative Review. *Int J Public Health*. 2022;67:1604961. doi:10.3389/ijph.2022.1604961
- Friedman N, Drori Y, Pando R, et al. A(H1N1)pdm09 influenza infection: vaccine inefficiency. *Oncotarget*. 2017;8(20):32856-32863. doi:10.18632/oncotarget.16459
- Broberg E, Snacken R, Adlhoch C, et al. Start of the 2014/15 influenza season in Europe: drifted influenza A(H3N2) viruses circulate as dominant subtype. *Euro Surveill Bull Eur Sur Mal Transm Eur Commun Dis Bull*. 2015;20(4):21023. doi:10.2807/1560-7917.es2015.20.4.21023

- 60. Appiah GD, Blanton L, D'Mello T, et al. Influenza activity United States, 2014-15 season and composition of the 2015-16 influenza vaccine. *MMWR Morb Mortal Wkly Rep*. 2015;64(21):583-590.
- 61. Centers for Disease Control and Prevention (CDC). Influenza activity--United States, 2012-13 season and composition of the 2013-14 influenza vaccine. *MMWR Morb Mortal Wkly Rep.* 2013;62(23):473-479.
- 62. Fiore AE, Uyeki TM, Broder K, et al. Prevention and control of influenza with vaccines: recommendations of the Advisory Committee on Immunization Practices (ACIP), 2010. *MMWR Recomm Rep Morb Mortal Wkly Rep Recomm Rep.* 2010;59(RR-8):1-62.
- Kieninger D, Sheldon E, Lin WY, et al. Immunogenicity, reactogenicity and safety of an inactivated quadrivalent influenza vaccine candidate versus inactivated trivalent influenza vaccine: a phase III, randomized trial in adults aged ≥18 years. *BMC Infect Dis*. 2013;13:343. doi:10.1186/1471-2334-13-343
- 64. Belshe RB. The need for quadrivalent vaccine against seasonal influenza. *Vaccine*. 2010;28Suppl 4:D45-53. doi:10.1016/j.vaccine.2010.08.028
- 65. Langley JM, Carmona Martinez A, Chatterjee A, et al. Immunogenicity and safety of an inactivated quadrivalent influenza vaccine candidate: a phase III randomized controlled trial in children. *J Infect Dis.* 2013;208(4):544-553. doi:10.1093/infdis/jit263
- 66. Tinoco JC, Pavia-Ruz N, Cruz-Valdez A, et al. Immunogenicity, reactogenicity, and safety of inactivated quadrivalent influenza vaccine candidate versus inactivated trivalent influenza

vaccine in healthy adults aged ≥ 18 years: a phase III, randomized trial. *Vaccine*. 2014;32(13):1480-1487. doi:10.1016/j.vaccine.2014.01.022

- 67. Notarte KI, Ver AT, Velasco JV, et al. Effects of age, sex, serostatus, and underlying comorbidities on humoral response post-SARS-CoV-2 Pfizer-BioNTech mRNA vaccination: a systematic review. *Crit Rev Clin Lab Sci.* 2022;59(6):373-390. doi:10.1080/10408363.2022.2038539
- Urakawa R, Isomura ET, Matsunaga K, Kubota K. Young Age, Female Sex, and No Comorbidities Are Risk Factors for Adverse Reactions after the Third Dose of BNT162b2 COVID-19 Vaccine against SARS-CoV-2: A Prospective Cohort Study in Japan. *Vaccines*. 2022;10(8):1357. doi:10.3390/vaccines10081357
- 69. Research C for BE and. FLUAD QUADRIVALENT. FDA. Published July 3, 2023. Accessed December 3, 2023. https://www.fda.gov/vaccines-blood-biologics/fluadquadrivalent
- 70. Research C for BE and. Fluzone Quadrivalent, Fluzone High-Dose Quadrivalent, Fluzone, Intradermal Quadrivalent, Fluzone Quadrivalent Southern Hemisphere, Fluzone High-Dose Quadrivalent Southern Hemisphere. *FDA*. Published online February 15, 2024. Accessed March 6, 2024. https://www.fda.gov/vaccines-blood-biologics/vaccines/fluzonequadrivalent-fluzone-high-dose-quadrivalent-fluzone-intradermal-quadrivalent-fluzone
- 71. Research C for BE and. Flublok Quadrivalent. *FDA*. Published online July 12, 2023.
 Accessed March 6, 2024. https://www.fda.gov/vaccines-blood-biologics/vaccines/flublokquadrivalent

- 72. Badizadegan K, Goodson JL, Rota PA, Thompson KM. The potential role of using vaccine patches to induce immunity: platform and pathways to innovation and commercialization. *Expert Rev Vaccines*. 2020;19(2):175-194. doi:10.1080/14760584.2020.1732215
- 73. Mclenon J, Rogers MAM. The fear of needles: A systematic review and meta-analysis. J Adv Nurs. 2019;75(1):30-42. doi:10.1111/jan.13818
- 74. Hettinga J, Carlisle R. Vaccination into the Dermal Compartment: Techniques, Challenges, and Prospects. *Vaccines*. 2020;8(3):534. doi:10.3390/vaccines8030534
- 75. Creighton RL, Woodrow KA. Microneedle-Mediated Vaccine Delivery to the Oral Mucosa. *Adv Healthc Mater*. Published online December 10, 2018:1801180. doi:10.1002/adhm.201801180
- 76. Intradermal Delivery of Vaccines A Review of the Literature and the Potential for Development for Use in Low and Middle Income Countries. Program for Appropriate Technology in Health (PATH); 2009.
- 77. Norman JJ, Arya JM, Mcclain MA, Frew PM, Meltzer MI, Prausnitz MR. Microneedle patches: Usability and acceptability for self-vaccination against influenza. *Vaccine*. 2014;32(16):1856-1862. doi:10.1016/j.vaccine.2014.01.076
- 78. Vicente-Pérez EM, Quinn HL, Mcalister E, et al. The Use of a Pressure-Indicating Sensor Film to Provide Feedback upon Hydrogel-Forming Microneedle Array Self-Application In Vivo. *Pharm Res.* 2016;33(12):3072-3080. doi:10.1007/s11095-016-2032-z

- 79. Guillermet E, Alfa DA, Phuong Mai LT, et al. End-user acceptability study of the nanopatchTM; a microarray patch (MAP) for child immunization in low and middle-income countries. *Vaccine*. 2019;37(32):4435-4443. doi:10.1016/j.vaccine.2019.02.079
- 80. Arya J, Henry S, Kalluri H, Mcallister DV, Pewin WP, Prausnitz MR. Tolerability, usability and acceptability of dissolving microneedle patch administration in human subjects. *Biomaterials*. 2017;128:1-7. doi:10.1016/j.biomaterials.2017.02.040
- Korkmaz E, Balmert SC, Sumpter TL, Carey CD, Erdos G, Falo LD. Microarray patches enable the development of skin-targeted vaccines against COVID-19. *Adv Drug Deliv Rev*. 2021;171:164-186. doi:10.1016/j.addr.2021.01.022
- 82. Forster AH, Witham K, Depelsenaire ACI, et al. Safety, tolerability, and immunogenicity of influenza vaccination with a high-density microarray patch: Results from a randomized, controlled phase I clinical trial. *PLOS Med.* 2020;17(3):e1003024. doi:10.1371/journal.pmed.1003024
- 83. Iwata H, Kakita K, Imafuku K, et al. Safety and dose-sparing effect of Japanese encephalitis vaccine administered by microneedle patch in uninfected, healthy adults (MNA-J): a randomised, partly blinded, active-controlled, phase 1 trial. *Lancet Microbe*. 2022;3(2):e96-e104. doi:10.1016/s2666-5247(21)00269-x
- 84. Fernando GJP, Hickling J, Jayashi Flores CM, et al. Safety, tolerability, acceptability and immunogenicity of an influenza vaccine delivered to human skin by a novel high-density microprojection array patch (NanopatchTM). *Vaccine*. 2018;36(26):3779-3788. doi:10.1016/j.vaccine.2018.05.053

- 85. Center for Biologics Evaluation and Research. Toxicity Grading Scale for Healthy Adult and Adolescent Volunteers Enrolled in Preventive Vaccine Clinical Trials. U.S. Food and Drug Administration. Published May 17, 2019. Accessed February 23, 2023. https://www.fda.gov/regulatory-information/search-fda-guidance-documents/toxicitygrading-scale-healthy-adult-and-adolescent-volunteers-enrolled-preventive-vaccine-clinical
- 86. Rouphael NG, Lai L, Tandon S, et al. Immunologic mechanisms of seasonal influenza vaccination administered by microneedle patch from a randomized phase I trial. NPJ Vaccines. 2021;6(1):89. doi:10.1038/s41541-021-00353-0
- McNutt LA, Wu C, Xue X, Hafner JP. Estimating the relative risk in cohort studies and clinical trials of common outcomes. *Am J Epidemiol*. 2003;157(10):940-943. doi:10.1093/aje/kwg074
- Zou G. A modified poisson regression approach to prospective studies with binary data. Am J Epidemiol. 2004;159(7):702-706. doi:10.1093/aje/kwh090
- Zou GY, Donner A. Extension of the modified Poisson regression model to prospective studies with correlated binary data. *Stat Methods Med Res*. 2013;22(6):661-670. doi:10.1177/0962280211427759
- 90. Yelland LN, Salter AB, Ryan P. Performance of the Modified Poisson Regression Approach for Estimating Relative Risks From Clustered Prospective Data. *Am J Epidemiol.* 2011;174(8):984-992. doi:10.1093/aje/kwr183

- 91. Tozuka M, Oka T, Jounai N, et al. Efficient antigen delivery to the draining lymph nodes is a key component in the immunogenic pathway of the intradermal vaccine. *J Dermatol Sci*. 2016;82(1):38-45. doi:10.1016/j.jdermsci.2015.11.008
- 92. Charles A Janeway J, Travers P, Walport M, Shlomchik MJ. The distribution and functions of immunoglobulin isotypes. In: *Immunobiology: The Immune System in Health and Disease. 5th Edition*. Garland Science; 2001. Accessed March 28, 2024. https://www.ncbi.nlm.nih.gov/books/NBK27162/
- 93. Chen N, Wang W, Fauty S, et al. The effect of the neonatal Fc receptor on human IgG biodistribution in mice. *mAbs*. 2014;6(2):502-508. doi:10.4161/mabs.27765
- 94. Monto AS, Petrie JG, Cross RT, et al. Antibody to Influenza Virus Neuraminidase: An Independent Correlate of Protection. *J Infect Dis*. 2015;212(8):1191-1199. doi:10.1093/infdis/jiv195
- 95. Gilbert PB, Fong Y, Juraska M, et al. HAI and NAI titer correlates of inactivated and live attenuated influenza vaccine efficacy. *BMC Infect Dis*. 2019;19(1):453. doi:10.1186/s12879-019-4049-5
- 96. Weiss CD, Wang W, Lu Y, et al. Neutralizing and Neuraminidase Antibodies Correlate With Protection Against Influenza During a Late Season A/H3N2 Outbreak Among Unvaccinated Military Recruits. *Clin Infect Dis Off Publ Infect Dis Soc Am*. 2020;71(12):3096-3102. doi:10.1093/cid/ciz1198
- 97. Memoli MJ, Shaw PA, Han A, et al. Evaluation of Antihemagglutinin and Antineuraminidase Antibodies as Correlates of Protection in an Influenza A/H1N1 Virus

Healthy Human Challenge Model. *mBio*. 2016;7(2):e00417-00416. doi:10.1128/mBio.00417-16

- 98. Couch RB, Atmar RL, Franco LM, et al. Antibody correlates and predictors of immunity to naturally occurring influenza in humans and the importance of antibody to the neuraminidase. *J Infect Dis.* 2013;207(6):974-981. doi:10.1093/infdis/jis935
- Kastenschmidt JM, Sureshchandra S, Jain A, et al. Influenza vaccine format mediates distinct cellular and antibody responses in human immune organoids. *Immunity*. 2023;56(8):1910-1926.e7. doi:10.1016/j.immuni.2023.06.019
- 100. Schmidt A, Lapuente D. T Cell Immunity against Influenza: The Long Way from Animal Models Towards a Real-Life Universal Flu Vaccine. *Viruses*. 2021;13(2):199. doi:10.3390/v13020199
- 101. Korenkov D, Isakova-Sivak I, Rudenko L. Basics of CD8 T-cell immune responses after influenza infection and vaccination with inactivated or live attenuated influenza vaccine. *Expert Rev Vaccines*. 2018;17(11):977-987. doi:10.1080/14760584.2018.1541407