

## Distribution Agreement

In presenting this thesis or dissertation as a partial fulfillment of the requirements for an advanced degree from Emory University, I hereby grant to Emory University and its agents the non-exclusive license to archive, make accessible, and display my thesis or dissertation in whole or in part in all forms of media, now or hereafter known, including display on the world wide web. I understand that I may select some access restrictions as part of the online submission of this thesis or dissertation. I retain all ownership rights to the copyright of the thesis or dissertation. I also retain the right to use in future works (such as articles or books) all or part of this thesis or dissertation.

Signature:

---

Patricia A. Woodall

Date

Approval Sheet

Association of Schistosomiasis with Impaired Fertility in East Africa

By

Patricia Woodall

Master of Public Health

Global Epidemiology

---

Michael Kramer

Faculty Thesis Advisor

Abstract Cover Page

Association of Schistosomiasis with Impaired Fertility in East Africa

By

Patricia Woodall

BS, Birmingham Southern College, 1975

MD, Vanderbilt University School of Medicine, 1979

MPH, University of Hawaii, 1996

MPHTM, Tulane University School of Public Health and Tropical Medicine, 2009

Faculty Thesis Advisor: Michael Kramer, MMSc, PhD

An abstract of

A thesis submitted to the Faculty of the Rollins School of Public Health of Emory  
University

in partial fulfillment of the requirements for the degree of

Master of Public Health in Global Epidemiology

2016

## Abstract

### Association of Schistosomiasis with Impaired Fertility in East Africa

By

Patricia Woodall

Many case reports and pathology series have suggested associations of Female Genital Schistosomiasis of the Fallopian tubes with infertility and ectopic pregnancy. Geographic distribution of infertility (which in Africa is most commonly due to tubal disease) has been reported but not explained. In this cross-sectional study, interpolated prevalence maps for *S. haematobium* and *S. mansoni* in East Africa were created using data from two open-access Neglected Tropical Diseases databases. Prevalence was extracted to georeferenced survey sample points for Demographic and Health Surveys for Ethiopia, Kenya, Tanzania and Uganda for 2009-2011 and 1999-2001. Outcomes included primary and secondary infertility (no births) and infecundity (no pregnancies) and history of pregnancy loss. Exploratory spatial analyses of outcomes (Moran's I, univariate and bivariate Local Indices of Spatial Autocorrelation) showed that outcomes were not spatially random and mapped clustering, hotspots, and areas of co-location of outcomes and exposures. Weighted multilevel logistic regression analysis found that women living in high compared to absent *S. haematobium* locations had significantly higher odds of secondary infertility (1999-2001: OR 1.8 [CI<sub>95</sub> 1.4, 2.3]; 2009-2011: OR 1.23 [1.1, 1.5]) and of primary infertility (1999-2001: OR 1.8 [1.3, 2.7]; 2001-2011: OR 1.58 [1.1, 2.3]). Living in high compared to absent *S. mansoni* locations did not affect the odds of any outcome. Women living in high *S. haematobium* compared to high *S. mansoni* locations had significantly higher odds of secondary infertility (1999-2001: OR 1.7 [1.3, 2.3]; 2009-2011: OR 1.6 [1.1, 2.0]), and of primary infertility (2010: OR 2.7 [1.5, 4.9]). For 1999-2001, history of pregnancy loss was significantly associated with high compared to absent *S. haematobium* (OR 1.3 [1.1, 1.6]) and with high *S. haematobium* compared to high *S. mansoni* (OR 1.4 [1.0, 1.8]). There is increasing evidence of the clinical and public health consequences of schistosomiasis to women's health and the importance of inclusion of girls and women in control strategies.

Cover Page

Association of Schistosomiasis with Impaired Fertility in East Africa

By

Patricia Woodall

BS, Birmingham Southern College, 1975

MD, Vanderbilt University School of Medicine, 1979

MPH, University of Hawaii, 1996

MPHTM, Tulane University School of Public Health and Tropical Medicine, 2009

Faculty Thesis Advisor: Michael Kramer, MMSc, PhD

A thesis submitted to the Faculty of the  
Rollins School of Public Health of Emory University  
in partial fulfillment of the requirements for the degree of  
Master of Public Health in Global Epidemiology

2016

## TABLE OF CONTENTS

| Page |   |
|------|---|
| 1    | Chapter I: Background/Literature Review   |
| 11   | Chapter II: Manuscript: Abstract  |
| 13   | Introduction  |
| 18   | Methods   |
| 25   | Results   |
| 31   | Discussion  |
| 34   | Chapter III: Future Directions  |
| 35   | Table 1. Published reports of infertility associated with schistosomiasis.  |
| 36   | Table 2. Published reports of ectopic pregnancy associated with schistosomiasis.  |
| 37   | Table 3. Population description, DHS East Africa 1999-2001  |
| 38   | Table 4. Population description, DHS East Africa 2009-2011  |
| 39   | Table 5. Measures of autocorrelation of outcome variables, Moran's I East Africa 1999-2001 & 2009-2011                        |
| 40   | Table 6. Association of impaired fertility with schistosomiasis prevalence, East Africa 1999-2001                             |
| 41   | Table 7. Association of impaired fertility with schistosomiasis prevalence, East Africa 2009-2011                             |
| 42   | Table 8. Association of impaired fertility with schistosomiasis prevalence, East Africa 1999-2001. Sensitivity analysis.      |
| 44   | Table 9. Association of impaired fertility with schistosomiasis prevalence, East Africa 2009-2011. Sensitivity analysis.      |
| 46   | Figure 1: Map of schistosomiasis distribution in East Africa.<br>Figure 2: Potential effects of schistosomiasis in pregnancy. |
| 48   | Figure 3. Predicted (interpolated) schistosomiasis distribution in East Africa,   |
| 49   | Figure 4. Local clustering of impaired fertility outcomes, Getis-Ord $G_i^*$  |
| 50   | Figure 5. Bivariate LISA, impaired fertility outcomes and schistosomiasis prevalence  |
| 52   | References  |
| 64   | Appendix: additional maps<br>Figure 6. Local clustering of impaired fertility outcomes, Local Moran's I                       |
| 65   | Figure 7. Measures of impaired fertility, box maps.   |
| 66   | Figure 8. Measures of impaired fertility, univariate LISA   |

# ASSOCIATION OF SCHISTOSOMIASIS WITH IMPAIRED FERTILITY IN EAST AFRICA

Patricia Woodall MD, MPH, MPH&TM

Emory University Rollins School of Public Health

## SECTION I: LITERATURE REVIEW

The intersection of the tropical infectious diseases with women's reproductive health has drawn limited attention. Increasing recognition that schistosomiasis is a reproductive as well as a urinary pathogen led to the renaming of urinary to urogenital schistosomiasis. This study will contribute to the knowledge base of these reproductive system effects.

### SCHISTOSOMIASIS

Schistosomiasis is a group of parasitic diseases of humans characterized by the presence of paired long-lived (up to 25 years) trematode worms in pelvic and mesenteric vascular plexuses, resulting in pathology caused by deposition of eggs in genitourinary (in the case of *Schistosoma haematobium*) and digestive organs (primarily *Schistosoma mansoni* and *S. japonicum*) and in transmission by passage of eggs in human excreta. After cycling through an intermediate snail host, the parasite is acquired by humans via skin penetration in fresh water. *S. haematobium* occurs throughout Africa, *S. mansoni* in Africa and northern South America, and *S. japonicum* in parts of Southeast Asia and China (1). At least 200 to 600 million people are infected, over 90% of these in Africa (2). Distribution is focal, related to water sources and snail populations, with local prevalence extremely high (50-80%) in affected areas (1). Although historically the resulting human morbidities have been considered to be diarrhea, hepatic cirrhosis, and bladder cancer, recently the much broader spectrum of disease has been recognized, encompassing other chronic urinary tract, genital, and hepatic pathology as well as more subtle effects on anemia and

chronic inflammation (3, 4). Clinical diagnosis is by presence of eggs on microscopy of urine or feces (depending on species) or in pathologic specimens. Recently serologic diagnosis of *S. mansoni* has become accepted, but serologic test characteristics remain unproven for *S. haematobium* (5). Exposure occurs where humans have skin contact with fresh water inhabited by the appropriate host snail and in which eggs have been deposited through urine or feces of infected humans. The popular image of young boys exposed during recreational swimming in irrigation ditches is a limited picture. Exposure occurs among all age groups and both sexes during washing and agricultural work in infested lakes, rivers, and canals. Immunologic factors are thought to reduce new infection rates in adults, after death of worms acquired in childhood; however, re-infection does occur (1).

Prevalence surveys measure egg shedding in urine or feces (measuring active presence of reproducing worm pairs), but not disease (organ damage). Although liver damage may regress after antihelminthic treatment or natural worm death, more advanced liver and genitourinary damage do not. Prevalence (of egg shedding) is typically highest among school-age children, but occurs in all ages (1).

Geographic distribution of the *Schistosoma* species is determined by the distribution of the specific snail host species. One species typically predominates in a given geographic region, although species may co-exist. Typical snail habitat is along freshwater bodies in warm, low-altitude, high-rainfall areas (6). Dams and irrigation schemes may increase snail habitat. Although the *Bulinus* snail host of *S. haematobium* tolerates higher temperature and lower or seasonal rainfall than does the *Biomphalaria* snail host of *S. mansoni*, no specific habitat differences explain the overall difference in geographic distribution of the two species (7, 8).

Older environmental methods of disease control by wetland management or snail control gave way to Mass Drug Administration (MDA) with Praziquantel (PZ) in the 1980s. Due to highest prevalence in school-age children, convenience of school-based MDA, limited availability of PZQ, and (until 2003) lack of approval of PZQ in pregnancy (9), control programs



(where they exist) have typically provided annual or semi-annual PZQ, often integrated with other deworming, to school children. Targeting has been based on prevalence surveys of children (10). Expectations that this approach would lead to eventual local elimination of infection have not been fulfilled (11). The school-based approach misses younger children, girls, adolescents, and adults, thus allowing transmission as well as disease to persist. In 2009 it was estimated that only 5% of the infected population were receiving PZQ (12). Recent evaluations have questioned the impact of MDA with PZQ on prevalence, transmission, and long-term pathology (13).

Of the countries addressed in this study, neither Ethiopia (14) nor Kenya (15) had implemented schistosomiasis control programs prior to the relevant DHS surveys. Tanzania (16) and Uganda (17) partially implemented school-based MDA beginning in 2004. Zanzibar and Pemba islands (in Tanzania) implemented MDA with PZQ in the mid-1980s, but with limited coverage until 2004 (18).

The World Health Organization mapped the known distribution of schistosomiasis in 1987 (19). Recently, detailed mapping has been carried out in many affected countries to facilitate planning of control programs, often integrated with mapping for control of other NTDs such as soil-transmitted helminths and lymphatic giardiasis (20-22). *S. mansoni* is distributed widely in Ethiopia and Uganda and more focally in western and southern Kenya, especially along the shores of Lake Victoria. *S. haematobium* is widely distributed in Tanzania, especially coastal areas, south of Lake Victoria, and Zanzibar islands (estimated national prevalence 50%); in coastal Kenya; and focally in Ethiopia (23). (Figure 1).

### **Schistosomiasis in women**

In addition to the hepatic and urinary effects common to both sexes, schistosomiasis has effects unique to the female reproductive system and reproductive functions. Deposition of eggs leads to granulomas, ulceration, and distortion in ureters, bladder, Fallopian tubes, ovaries, cervix

and vagina (24-26). Although documented for *S. mansoni* (27), this predominantly occurs from *S. haematobium*.

Female Genital Schistosomiasis (FGS) has been recognized for over a century by Egyptian physicians, especially as a major cause of vesicovaginal fistula (28). As the reproductive system disease is chronic and not clinically apparent, it received little attention until mounting evidence of its impact and probable association with HIV (29) led to renaming of *S. haematobium* from urinary to urogenital schistosomiasis. Population-based studies in Madagascar (30), Tanzania (31, 32), Malawi (33), and Egypt (34) have confirmed that vaginal/cervical schistosomiasis (sandy patches, ulcers, fungating lesions identified on vaginal/cervical visualization) occurs in 30-75% of women who have urinary schistosomiasis. Among Tanzanian women of reproductive age with 40% prevalence of urinary and 32% prevalence of cervical schistosomiasis, 23% had FGS with no urinary egg excretion (35). Thus urinary egg counts underestimate schistosomiasis in women.

FGS is associated with a variety of gynecologic symptoms. In Madagascar, significantly more women in an endemic village reported miscarriage, irregular menses, pelvic pain and vaginal discharge than in a control village (30). In Tanzania, post-coital bleeding and genital ulceration were more frequently reported in the endemic site, and FGS cases reported more irregular menses and hypermenorrhea than non-cases (32). 35% of pre-pubertal 10 to 12 year old girls in an *S. haematobium*-endemic area of South Africa reported genital symptoms (discharge, itching, sores) (36). The prevalence of upper tract (Fallopian tube) disease is unknown due to difficulty of assessment, but pelvic ultrasound abnormalities (ovarian cysts, uterine mass, hydrosalpinx) were seen in 9% of women in one affected community in Madagascar (37). The effects of schistosomiasis on the male reproductive system have been even less well studied, but pathology studies report anatomic effects and there are case reports of hemospermia and infertility (38).

Schistosomiasis may potentially affect female reproductive function by several mechanisms (Fig. 2). Little has been published regarding effects of schistosomiasis on pregnancy outcomes. Studies reporting prevalence of schistosomiasis in pregnant women in nine African countries were summarized by Salawu (39). Animal studies and small descriptive studies have on crude analysis suggested increased risk of prematurity (40) and of low birth weight (41-43) in offspring of women with schistosomiasis, thought to be associated with increased anemia or with placental infection (44). A meta-analysis of 20 published studies showed a mean hemoglobin deficit of 4 g/L associated with schistosomiasis (primarily *S. mansoni*) (45). Four descriptive studies of anemia in pregnant women with schistosomiasis found no association for *S. mansoni* in Tanzania and Uganda (46, 47) and a 1.2 - 2.4 OR for anemia in *S. haematobium* in Nigeria and Ghana (48, 49). In Uganda, Praziquantel treatment during pregnancy had no effect on maternal anemia, birthweight, perinatal mortality, or congenital anomalies (50).

Although the major causes of maternal mortality (hemorrhage, hypertensive disorder, sepsis) can each hypothetically be linked to schistosomiasis, published information on actual contribution is lacking. Coagulopathy occurs in young adults with hepatic schistosomiasis (51), suggesting possible contribution to maternal hemorrhage. A systematic review of kidney disease in sub-Saharan Africa summarized high prevalence of renal dysfunction and hydronephrosis/hydroureter in children in *S. haematobium*-endemic areas (52), suggesting possible contribution to eclampsia and to prolepsis. Bacteriuria prevalence of 30-50% has been reported in Nigerian children with urinary schistosomiasis (53, 54). Ossai found OR=31 for bacteriuria in children with/without urinary schistosomiasis (55). Bacteriuria in pregnancy is associated with preterm birth and with maternal sepsis. Three case reports of pregnancies in women with hepatic schistosomiasis report significant thrombocytopenia (56-58). One case report describes pre-eclampsia and fetal death in a woman with hepatic schistosomiasis (59).

Fifteen published case reports were identified linking FGS with ectopic pregnancy (Table 1). The extent of death of women from ruptured ectopic pregnancy in Africa is unknown (60), but

it is likely that most affected women die without being diagnosed, treated, or counted as maternal deaths. From surveillance data and verbal autopsy, Olack estimated that 25% of maternal deaths in an urban slum in Nairobi, Kenya 2009-2013 were due to ectopic pregnancy (61). Hospital series in Ghana and South Africa have reported 1-4% of pregnancies as ectopic with 53-61% in hypovolemic shock on arrival (62, 63). 61% of South African women with ectopic pregnancy had a known history of infertility (63). 73% of 145 Gabon women surviving ectopic pregnancy were subsequently infertile, and 10% had a repeat ectopic pregnancy (64). In Mozambique, 3.9% of operated ectopic pregnancies had associated schistosomiasis (65).

Schistosomiasis could be a primary etiology or a co-factor in occurrence of obstetric vesicovaginal fistula. Egyptian writers have long considered schistosomiasis to be the primary etiology of fistulas (28). One small case-control study did not find an association but found that schistosomiasis deterred healing of obstetric fistula (66).

## **INFERTILITY**

Infertility may be studied as demographic variable (affecting overall fertility rates), as a socio-cultural factor (leading to stigma, divorce, abandonment, and socio-economic burden) (67) and as a marker of its underlying biomedical determinants. Infertility may be a rough surrogate for ectopic pregnancy, otherwise difficult to measure in resource-limited settings where it likely results in unattended death.

Terms used (often inconsistently) include *infertility*, *sub-fertility*, *sterility*, and *infecundity*, with the latter two terms typically considering a permanent state (unmeasurable until after a woman has completed her childbearing years) and the former (infertility) referring to a measureable time period (68). *Fecundity* may refer to conception and *fertility* to birth, or vice-versa (69). *Sub-fertility* or *sub-fecundity* may include both failure to conceive and pregnancy losses (miscarriage, stillbirth, ectopic pregnancy), or may refer to not obtaining the desired number of children (70).

*Clinical definitions* (for individual diagnosis and treatment) typically measure conception and use short intervals (failure to conceive by 12 months). The *epidemiologic definition* (for monitoring and surveillance) of prevalence of infertility is the "percentage of women of reproductive age (15–49) at risk of pregnancy (not pregnant, sexually active, non-contracepting and non-lactating) who report trying for a pregnancy for two years or more" (71) (note that neither conception nor birth is specified - just "a pregnancy"). The *demographic definition* is failure of a sexually active, non-contracepting woman who desires a child to give birth within a specified time period (variously 3 to 7 years, to account for lactational amenorrhea and socio-cultural factors relating to resumption of intercourse and of childbearing) (70). As contraceptive coverage has increased globally, contraceptive use and desire for pregnancy have become more important factors in definitions.

At clinical level, the unit of analysis is the *couple* and infertility is classified as being due to male factor, female factor (tubal, endocrine, other), or unknown. Epidemiologically or demographically, the unit of analysis is the *woman* (for whom data are measured). *Primary infertility* is failure to produce a live birth (thus including women with early pregnancy loss or stillbirth). *Secondary infertility* is failure of a subsequent birth after having given birth. Bongaarts differentiates *natural sterility* (estimated at ~3% globally) and *pathologic sterility*, that due to a disease process (72). The preferred *denominator* is women ever married or in union. Using all Women of Reproductive Age (WRA) is too inclusive of those who are not sexually active. Counting only those currently married or in union excludes those who have been divorced or abandoned due to the infertility (69).

*Operationalizing* these definitions for analysis is problematic. Demographic surveys were designed to obtain information about fertility and contraceptive practices; few specifically ask about infertility or desire for pregnancy (73). Data are self-reported and variables must be derived from directly reported variables. Various formulas have been used, especially to measure secondary infertility, based on "self-reported infecundity", "behavioral infecundity", the "open

birth interval" or the "subsequently infertile estimator" (73, 74) and various modifications of it (75). These require complex longitudinal analysis methods to provide age-specific estimates. Operational definitions have been proposed for use with DHS data (75-77), and DHS includes an infertility variable designed to estimate population-level infertility risk.

Using data from 47 Demographic and Health Surveys, Rutstein estimated that over 186 million (25%) women in developing countries were infertile, with prevalence of 4% for primary infertility (5% if pregnancy losses are included) and secondary infertility rates increasing sharply with age (from 5% to 62%) (69). From a 2010 analysis of 277 demographic and reproductive health surveys, Mascarenas estimated that 48.5 million couples globally are infertile, with global prevalence of 1.9% for primary and 10% for secondary infertility (78). Both authors found wide variations among countries, with highest rates in sub-Saharan Africa. From 1978-1984 a WHO-sponsored multi-center worldwide study of 5800 infertile couples was carried out. Africa showed a different pattern from other regions; tubal causes predominated, with 3 times the rate of tubal occlusion as other regions, even though tubal disease was likely underestimated due to use of less-sensitive radiographic rather than surgical diagnostic methods (and thus no bacteriologic or histologic tests) (79).

### **Determinants of Tubal infertility**

Where medical investigation has occurred, tubal factors account for a much higher proportion of infertility in Africa than in other regions (79-82). Although tubal-factor infertility is usually attributed to sexually transmitted infections (gonorrhea and chlamydia), this is generally an assumption, as microbiologic or pathologic testing is rarely done, the diagnosis of tubal distortion or obstruction is made by radiologic imaging (hysterosalpingogram, HSG), and other possible etiologies are not considered. Where serologic testing has been done or pathologic specimens obtained during surgical procedures, only 2-20% of tubal disease was due to

chlamydia (83, 84). Six to 21% of tubal infertility cases in South African clinics were due to genito-urinary tuberculosis (85, 86).

Many case reports, case series, and pathology series have identified Fallopian tube distortion and obstruction due to schistosomiasis in association with infertility (Figure 3). Two small community-based studies have addressed the possible association of schistosomiasis with infertility. In two *S. haematobium*-affected communities in Zimbabwe, 15% of women were infertile, with OR 3.6 for FGS among the infertile (87). Sub-fertility was found in 44% of women in a highly schistosomiasis-endemic area of Coast Province, Tanzania (88).

### **Geographic distribution of infertility**

Differential geographic distribution of infertility is noted, usually speculatively attributed to differing marriage patterns or sexual behaviors leading to sexually transmitted infections.

Portions of central Africa (Congo, Central African Republic, Congo, Cameroon, Gabon) have for decades been known as the "infertility belt" with over 20% of women older than 40 childless (89). Romaniuk noted high prevalence of infertility along the Congo and Uele Rivers in Congo, close to Lake Victoria and Lake Tanganyika, on the coastal regions of Tanzania and Kenya, on the Zanzibar islands, and in Tabora district, Tanzania (90). (Note that these Kenya and Tanzania locations reflect the distribution of *S. haematobium*). He noted a pattern of proximity to water, but attributed this to tribal territorial patterns. He also reviewed two clinical and radiographic studies which identified genital tract lesions in 60% and 75% of sterile Congo women (91, 92). These were presumed due to gonorrhoea, although this could be identified in only 25%. In 1996, Ericksen estimated infertility across 27 African countries from DHS or World Fertility Surveys, finding prevalence varying from 1-14% for primary and 5-17% for secondary infertility (73). She found associations with urban residence, early sexual debut, and multiple partners. She also found large variation among cultural groups across Africa; suggested that this may be due to different marriage, sexual, and female circumcision customs; but did not consider

geographic distribution as a possible variable (73). In Zambia, regional variation in infertility prevalence was not explained by STI risk (93).

In 2000 Larsen found wide variations in infertility rates by ethnic groups in Kenya and in Cameroon, presuming these to be due to cultural sexual practices affecting STI risk(94). A 1980-81 survey of infertility in rural Ethiopia found highest levels in Ilubabor region in the west (now Gambela), and suggested that ecological factors could be more responsible than ethnicity (95). (Note that this is one of the few *S. haematobium*-endemic areas in Ethiopia.) Larsen analyzed infertility data from DHS surveys for 29 African countries for the 1980s and 1990s to determine regional patterns. For rural Kenya, rates were highest in the Coast and Central regions; for Tanzania, Coast, Tabora Plateau, and South regions. She investigated ethnicity, age-cohort effects and different patterns of marital sex without successfully explaining the geographic differences in infertility (96). From 1991-92 DHS data, Erickson found overall infertility in Tanzania of 11.4%, primary infertility of 4.1% (73). (These regional patterns roughly coincide with *S. haematobium* endemicity). Blacker found highest Uganda infertility prevalence in Teso region (97).



## SECTION II: MANUSCRIPT

### ABSTRACT

Many case reports and pathology series have suggested associations of Female Genital Schistosomiasis of the Fallopian tubes with infertility and ectopic pregnancy. Geographic distribution of infertility (which in Africa is most commonly due to tubal disease) has been reported but not explained. In this cross-sectional study, interpolated prevalence maps for *S. haematobium* and *S. mansoni* in East Africa were created using data from two open-access Neglected Tropical Diseases databases. Prevalence was extracted to georeferenced survey sample points for Demographic and Health Surveys for Ethiopia, Kenya, Tanzania and Uganda for 2009-2011 and 1999-2001. Outcomes included primary and secondary infertility (no births) and infecundity (no pregnancies) and history of 1pregnancy loss. Exploratory spatial analyses of outcomes (Moran's I, univariate and bivariate Local Indices of Spatial Autocorrelation) showed that outcomes were not spatially random and mapped clustering, hotspots, and areas of co-location of outcomes and exposures. Weighted multilevel logistic regression analysis found that women living in high compared to absent *S. haematobium* locations had significantly higher odds of secondary infertility (1999-2001: OR 1.8 [CI<sub>95</sub> 1.4, 2.3]; 2009-2011: OR 1.23 [1.1, 1.5]) and of primary infertility (1999-2001: OR 1.8 [1.3, 2.7]; 2001-2011: OR 1.58 [1.1, 2.3]). Living in high compared to absent *S. mansoni* locations did not affect the odds of any outcome. Women living in high *S. haematobium* compared to high *S. mansoni* locations had significantly higher odds of secondary infertility (1999-2001: OR 1.7 [1.3, 2.3]; 2009-2011: OR 1.6 [1.1, 2.0]), and of primary infertility (2010: OR 2.7 [1.5, 4.9]). For 1999-2001, history of pregnancy loss was significantly associated with high compared to absent *S. haematobium* (OR 1.3 [1.1, 1.6]) and with high *S. haematobium* compared to

high *S. mansoni* (OR 1.4 [1.0, 1.8]). There is increasing evidence of the clinical and public health consequences of schistosomiasis to women's health and the importance of inclusion of girls and women in control strategies.

## INTRODUCTION

Schistosomiasis is a group of parasitic diseases of humans characterized by the presence of paired long-lived (up to 25 years) trematode worms in pelvic and mesenteric vascular plexuses, resulting in pathology caused by deposition of eggs in genitourinary (in the case of *Schistosoma haematobium*) and digestive organs (primarily *Schistosoma mansoni* and *S. japonicum*) and in transmission by passage of eggs in human excreta. After cycling through an intermediate snail host, the parasite is acquired by humans via skin penetration in fresh water. Exposure occurs among all age groups and both sexes during washing and agricultural work in infested freshwater bodies. *S. haematobium* occurs throughout Africa, *S. mansoni* in Africa and northern South America, and *S. japonicum* in parts of Southeast Asia and China (1). Distribution is focal, related to water sources and snail populations, with local prevalence extremely high (50-80%) in affected areas (1). At least 200 to 600 million people are infected, over 90% of these in Africa (2). Although historically the resulting human morbidities have been considered to be diarrhea, hepatic cirrhosis, and bladder cancer, recently the much broader spectrum of disease has been recognized, encompassing other chronic urinary tract, genital, and hepatic pathology as well as more subtle effects on anemia and chronic inflammation (3, 4). Advanced liver and genitourinary damage do not regress after antihelminthic treatment or natural worm death.

Clinical diagnosis is by presence of eggs on microscopy of urine or feces (depending on species) or in pathologic specimens. Prevalence surveys measure egg shedding in urine or feces (measuring active presence of reproducing worm pairs), but not disease (organ damage). Prevalence (of egg shedding) is highest among school-age children (1). One host snail species typically predominates in a given geographic region, with typical snail habitat along freshwater bodies in warm, low-altitude, high-rainfall areas (6). No specific habitat differences explain the overall difference in geographic distribution between the two species (7, 8).

Since the mid 1980s, control has focused on Mass Drug Administration (MDA) with Praziquantel (PZQ). Due to highest prevalence in school-age children, convenience of school-

based MDA, limited availability of PZQ, and (until 2003) lack of approval of PZQ in pregnancy (9), control programs (where they exist) have typically provided annual or semi-annual PZQ, often integrated with other deworming, to school children. In 2009 it was estimated that only 5% of the infected population were receiving PZQ (12). Recent evaluations have questioned the impact of MDA with PZQ on prevalence, transmission, and long-term pathology (13). Of the countries addressed in this study, neither Ethiopia (14) nor Kenya (15) had implemented schistosomiasis control programs prior to the relevant DHS surveys. Tanzania (16) and Uganda (17) implemented school-based MDA in certain locations in 2004 and beginning in the mid 1980s in Zanzibar.

Female Genital Schistosomiasis (FGS) occurs when deposition of (primarily) *S. haematobium* eggs leads to granulomas, ulceration, and distortion in ureters, Fallopian tubes, and other genitourinary organs (24-26). As the reproductive system disease is chronic and not clinically apparent, it received little attention until mounting evidence of its impact and probable association with HIV (29) led to renaming of *S. haematobium* from urinary to urogenital schistosomiasis. Population-based studies in Madagascar (30), Tanzania (31, 32), Malawi (33), and Egypt (34) have confirmed that vaginal/cervical schistosomiasis occurs in 30-75% of women who have urinary schistosomiasis. The prevalence of upper tract (Fallopian tube) disease is unknown due to difficulty of assessment, but pelvic ultrasound abnormalities (ovarian cysts, uterine mass, hydrosalpinx) were seen in 9% of women in one affected community in Madagascar (37). Pathology studies and case reports suggest that schistosomiasis also affects the male reproductive system (38).

Published reports of effects of schistosomiasis on reproductive outcomes are limited to case reports and descriptive series suggesting increased risk of prematurity (40) and of low birth weight (41-43); reports of schistosomiasis prevalence in pregnant women (39); and many case reports of FGS of the Fallopian tubes associated with infertility and ectopic pregnancy.

Impaired fertility is a common consequence of upper reproductive tract disease, often stigmatizing the woman due to the presumption of sexually transmitted infections as causal (67). The same tubal pathology may lead to ectopic pregnancy. 25% of maternal deaths in an urban slum in Nairobi, Kenya 2009-2013 were estimated to be due to ectopic pregnancy (61). Hospital series in Ghana and South Africa have reported 1-4% of pregnancies as ectopic with 53-61% in hypovolemic shock on arrival (62, 63). It is likely that most affected African women die without being diagnosed, treated, or counted as maternal deaths.

Terms used often inconsistently to describe impaired fertility include *infertility*, *sub-fertility*, *sterility*, and *infecundity* (68). *Fecundity* may refer to conception and *fertility* to birth, or vice-versa (69). *Clinical definitions* (for individual diagnosis and treatment) typically measure conception and use short intervals (failure to conceive by 12 months). The *epidemiologic definition* (for monitoring and surveillance) of prevalence of infertility is the "percentage of women of reproductive age (15–49) at risk of pregnancy (not pregnant, sexually active, non-contracepting and non-lactating) who report trying for a pregnancy for two years or more" (71) (note that neither conception nor birth is specified - just "a pregnancy"). The *demographic definition* is failure of a sexually active, non-contracepting woman who desires a child to give birth within a specified time period (variously 3 to 7 years, to account for lactational amenorrhea and socio-cultural factors relating to resumption of intercourse and of childbearing) (70). At clinical level, the unit of analysis is the *couple* and infertility is classified as being due to male factor, female factor (tubal, endocrine, other), or unknown. Epidemiologically or demographically, the unit of analysis is the *woman* (for whom data are measured). *Primary infertility* is failure to produce a live birth (thus including women with early pregnancy loss or stillbirth). *Secondary infertility* is failure of a subsequent birth after having given birth. The preferred *denominator* is women ever married or in union. Using all Women of Reproductive Age (WRA) is too inclusive of those who are not sexually active. Counting only those currently married or in union excludes those who have been divorced or abandoned due to the infertility

(69). Various formulas to operationalize these definitions for analysis have been proposed for use with demographic survey data (75-77).

Using data from 47 Demographic and Health Surveys, Rutstein estimated that over 186 million (25%) women in developing countries were infertile, with prevalence of 4% for primary infertility (5% if pregnancy losses are included) and secondary infertility rates increasing sharply with age (from 5% to 62%) (69). From a 2010 analysis of 277 demographic and reproductive health surveys, Mascarenas estimated that 48.5 million couples globally are infertile, with global prevalence of 1.9% for primary and 10% for secondary infertility (78). Both authors found wide variations among countries, with highest rates in sub-Saharan Africa. Where medical investigation has occurred, tubal factors account for a much higher proportion of infertility in Africa than in other regions (79-82). Although tubal-factor infertility is usually attributed to sexually transmitted infections, this is generally an assumption, as microbiologic or pathologic testing is rarely done, the diagnosis of tubal distortion or obstruction is made by radiologic imaging (hysterosalpingogram, HSG), and other possible etiologies are not considered. Where serologic testing has been done or pathologic specimens obtained during surgical procedures, only 2-20% of tubal disease was due to chlamydia (83, 84). Six to 21% of tubal infertility cases in South African clinics were due to genito-urinary tuberculosis (85, 86). Two small community-based studies have addressed the possible association of FGS with infertility. In two *S. haematobium*-affected communities in Zimbabwe, 15% of women were infertile, with OR 3.6 for FGS among the infertile (87). Sub-fertility was found in 44% of women in a highly schistosomiasis-endemic area of Coast Province, Tanzania (88).

Differential geographic distribution of infertility has been noted, usually speculatively attributed to differing marriage patterns or sexual behaviors leading to sexually transmitted infections. Portions of central Africa have for decades been known as the "infertility belt" with over 20% of women older than 40 childless (89). Romaniuk noted high prevalence of infertility along the Congo and Uele Rivers in Congo, close to Lake Victoria and Lake Tanganyika, on the

coastal regions of Tanzania and Kenya, on the Zanzibar islands, and in Tabora district, Tanzania, noting a pattern of proximity to water but attributing this to tribal territorial patterns (90). (Note that these Kenya and Tanzania locations reflect the distribution of *S. haematobium*). Clinical and radiographic studies in the 1950s identified genital tract lesions in 60% and 75% of sterile Congo women (91, 92). These were presumed due to gonorrhoea, although this could be identified in only 25%. In 1996, Ericksen estimated infertility across 27 African countries from DHS or World Fertility Surveys, finding prevalence varying from 1-14% for primary and 5-17% for secondary infertility (73). She found associations with urban residence, early sexual debut, and multiple partners. She also found large variation among cultural groups across Africa; suggested that this may be due to different marriage and sexual customs but did not consider geographic distribution as a possible variable (73). In Zambia, regional variation in infertility prevalence was not explained by STI risk (93). A 1980-81 survey of infertility in rural Ethiopia found highest levels in Ilubabor region in the west (now Gambela), and suggested that ecological factors could be more responsible than ethnicity (95). (Note that this is one of the few *S. haematobium*-endemic areas in Ethiopia.) Larsen analyzed infertility data from DHS surveys for 29 African countries for the 1980s and 1990s to determine regional patterns. For rural Kenya, rates were highest in the Coast and Central regions; for Tanzania, Coast, Tabora Plateau, and South regions. She investigated ethnicity, age-cohort effects and different patterns of marital sex without successfully explaining the geographic differences in infertility (96).

## METHODS

This cross-sectional multi-level study explored associations between impaired fertility in East Africa with the geographic distribution and prevalence of *S. haematobium* (associated with Fallopian tubal damage) and of *S. mansoni* (not commonly associated with Fallopian tubal damage). Ethiopia, Kenya, Tanzania and Uganda were included in this study as representing a contiguous geographic region with clearly delineated distribution of the two *Schistosoma* species of interest and with established DHS programs using the same methodology for collection of outcome data within a two-year period. Hypotheses are that risks of infertility, infecundity, and subfertility are associated with residence in high versus low *S. haematobium*-endemic locations; are associated with residence in high *S. haematobium*-endemic versus high *S. mansoni*-endemic locations; and do not differ with residence in high versus low *S. mansoni*-endemic locations.

### Data Sources

#### Exposure

The primary exposure data source was the Global Neglected Tropical Diseases Database ([www.gntd.org](http://www.gntd.org)), a georeferenced, open-access database of schistosomiasis prevalence data established within the EU-funded CONTRAST project based in the Swiss Tropical Institute. For this database, prevalence and location data dating from 1900 were abstracted from published scientific articles, institutional reports (WHO, government, control programs), and direct communication with researchers and health staff (98). Most studies determined point prevalence based on egg counts using the Kato-Katz method for *S. mansoni* and urine filtration for *S. haematobium*. Presence data predominated, although absence data were included for some locations. A few studies reported data aggregated at administrative levels.

For this study, additional exposure data were obtained from the Global Atlas of Helminth Infection ([thiswormyworld.org](http://thiswormyworld.org) and [ntdmap.org](http://ntdmap.org)), a similar database project of the London School of Hygiene and Tropical Medicine, which included some more recent data but lacked open access



to some of the earlier data and did not differentiate by species in the ntdmap.org platform (99, 100). Finally, to address areas with sparse data, additional data points were obtained from the WHO 1987 Global Atlas of Schistosomiasis (19) and geocoded (using Google Earth) by name of village and approximate map location. Where prevalence was reported only as quantiles, midpoints of the quantile were used. Although schistosomiasis distribution is heterogeneous at local and seasonal scale, existing maps show no apparent change in regional distribution over the 60-year time scale included in the database or the 50-year time scale of the measured outcomes. Several studies have used these databases to produce national and regional schistosomiasis prevalence and risk maps (6, 22, 23)

Data for the four countries of interest (Ethiopia, Kenya, Tanzania, and Uganda) were compiled into a single data file. To address border issues in interpolation, data for surrounding countries were also included, to cover an area from 16° North to -12° South latitude and from 28° West to 8° East longitude. In exploratory analysis, prevalence for each of the two *Schistosoma* species was mapped in ArcGIS as point prevalence, administrative-level prevalence, and kernel density.

A prevalence map for each of the two species was produced by interpolation using ArcGIS Geostatistical Analyst extension (Figure 3). Various kriging methods and models were tested to map the predicted distribution and prediction error, using half the randomized dataset for model calibration and half for model testing. The Empiric Bayesian Kriging model produced the lowest prediction error (RMS standardized error 0.95 for *S. haematobium* and 0.97 for *S. mansoni*). It is noted that geographic areas of high standard error coincided with areas of low human population (as indicated by few DHS datapoints).

Interpolated schistosomiasis prevalence was extracted to georeferenced DHS datapoints. DHS coordinates are geomasked by displacement of urban clusters up to 2 km and rural clusters up to 5 km, with a randomly selected 1% of rural clusters displaced up to 10 km (101). DHS guidance indicates that point extraction (rather than buffering to account for this displacement) is

acceptable as it has been shown to produce unbiased estimates for interpolated surfaces having moderate to high spatial autocorrelation (in this data, Moran's I = 0.49 for *S. haematobium*, 0.80 for *S. mansoni*) (101). Exposure was categorized as high (>25%), moderate (5-25%) or low (<5%) for each of the two *Schistosoma* species, based on commonly used programmatic categorizations, apparent natural breaks in histograms of the spatial data and sample size considerations, and confirmed in sensitivity analysis using other levels of categorization. Inclusion of Lake Victoria in maps addresses the very high schistosomiasis prevalence among people living on islands within the lake.

### **Outcomes and Covariates: Demographic and Health Surveys**

The Demographic and Health Surveys, a program of the US Agency for International Development (USAID), support a country's collection and analysis of nationally representative data about women's and children's health, fertility, contraception, social factors, malaria, and HIV, using standard questionnaires and methodologies (102, 103). Surveys use a two-stage sampling method with nationally representative selection of DHS clusters (modified for rural/urban and regional stratification) followed by PPS selection of households. All Women of Reproductive Age (15-49) within each selected household are surveyed. Surveys are typically carried out every 5 years. For this analysis and to explore temporal changes, surveys were combined for each of 2 time periods for which data were available: 2009-11 for Ethiopia, Kenya, Tanzania, and Uganda, and 1998-2000 for Ethiopia, Tanzania, and Uganda. (Georeferenced data were not available for the 2000 Kenya survey).

The reproductive factors measurable at individual level from DHS data include infertility and pregnancy loss, although pregnancy loss can only be assessed as a dichotomous variable of limited accuracy due to under-reporting (104). Differentiation of early miscarriage from stillbirth could not be accurately assessed due to large amount of missing data. Other outcomes of interest were not measurable. As women who have died are not sampled, DHS measures maternal

mortality through the sisterhood method, which would not correlate with the individual-level geographic exposure variable. Birth weight data is non-representative (over 50% missing). Urinary incontinence as a proxy for fistula was measured in the 2005 Ethiopia and Uganda DHS, but not in the *S. haematobium*-endemic countries.

Outcome measures used were *total infertility* (married, non-contracepting women who had not given birth within the past 5 years and were not currently pregnant), classified as *primary infertility* (never having given birth) and *secondary infertility* (ever having given birth).

Conception rather than livebirth was measured as *total infecundity* (married, non-contracepting women who had neither given birth nor reported a miscarriage or stillbirth within the past 5 years and were not currently pregnant), classified as *primary infecundity* (never having reported a birth, miscarriage or stillbirth) and *secondary infertility* (ever having reported a birth, miscarriage or stillbirth). *Infecundity* measures were only available for the 2009-2011 data, as calendar year of pregnancy losses (miscarriage or stillbirth) was not reported in the 2000 Tanzania survey.

Georeferenced data for other known risk factors for infertility (STIs, TB), which are not known to be spatially distributed, were not available.

For spatial analysis, individual outcome (case) data were aggregated as point prevalence at the geocoded DHS cluster locations and as areal prevalence at second administrative district level. Potential covariates derived from the DHS data included age (measured at subject level) and rural-urban status (measured at DHS cluster level). DHS defines "urban" as cities and small towns and "rural" as countryside, based on each country's governmental designation of the cluster location. Mismatch may be high. Comparison to satellite-imagery methods (GRUMP, MODIS) and population-density methods (WorldPop Project) found that most Tanzania DHS urban-defined clusters were classified as rural by other methods, less so for Uganda (106). However, the DHS classification was used in this study.

Guidance for spatial and regression modeling methods is provided by DHS, as well as examples of studies which have used similar methods to analyze associations between DHS variables and environmental exposures including malaria (105, 106).

### **Ethical review**

Review was waived by the Emory University Institutional Review Board. Necessary data use agreements were executed for MeasureDHS (USAID), the Global Neglected Tropical Diseases Database, and the Global Atlas of Helminth Infections.

### **Analysis**

Analyses were carried out using SAS 9.4 (Cary, NC), ArcGIS 10.3 (ESRI, Redland, CA), and GeoDa 1.6.6 (Tempe, AZ) (107). In addition to development of interpolated spatial models to quantify exposure (schistosomiasis prevalence), spatial methods (assessment of autocorrelation, cluster analysis) were used to assess spatial distribution of outcome variables (108, 109). Spatial analysis addresses the questions of whether the data are autocorrelated (not randomly spatially distributed) and where any clustering/aggregation occurs, and suggests spatial factors associated with this clustering. Spatial distribution of outcomes (measured as point prevalence) was explored analytically (global and local Moran's I) and visually (cluster and hotspot mapping). Possible associations of outcomes with exposures were explored visually with map overlay and analytically (as areal prevalence in GeoDa) with bivariate LISA (Local Indices of Spatial Autocorrelation). Mapping of urban sample points is more affected by displacement (in DHS data), and the inherent high population density tends to mask visualization of prevalence data (109). For these reasons and as both urban status and advanced age are strongly associated with the outcomes in this study, mapping of outcomes for rural women under 40 years of age provided the best visualization of the unconfounded spatial distribution of the outcomes.

Using pseudo-likelihood methods and empirical standard error estimation, odds ratios were estimated in a weighted multilevel logistic regression model. This mixed model included random-intercept effects for country/survey round (third level) and for DHS cluster (second level) and fixed effects at first (individual) and second level. As DHS second-stage sampling is by household with all eligible women in the household included (mean 1.05 women per household for the study population), inclusion of household as a level was considered, but produced no change in model fit or in estimates.

Indicator (dummy) variables for the two levels of comparison for each of the two *Schistosoma* species were measured at second (DHS cluster) level. Covariates included age as a continuous variable (first level) and rural/urban status as a dichotomous variable (second level). Estimates were weighted by DHS sample weights adjusted for differing country-level sampling fractions, as recommended in DHS guidance (110). Total infertility/infecundity and history of pregnancy loss were estimated as dichotomous outcome variables. Polytomous estimation methods were used for 3-level outcomes of primary and secondary (which sum to total) and no infertility/infecundity. Analyses were carried out in SAS proc GLIMMIX using SAS-recommended procedures for multilevel analysis (111, 112). Models included the two *Schistosoma* species simultaneously, controlling for possible co-endemicity, but were also estimated separately for each of the two *Schistosoma* species with minimal difference in results. Associations were estimated separately for each of the two survey rounds (1999-2001 and 2009-2011). As sensitivity analysis, associations were estimated for various age restrictions, exposure cut-off points, and combinations of countries.

$$\text{Model equation: } \text{logit}(Y_{ijk}) = \delta_{000} + (\delta_{100} * X_{ijk}) + \sum(\delta_{0n0} * Z_{njk}) + \mu_{0jk} + e_{00k}$$

where i = individual (first level), j = DHS cluster (second level), k=country (third level); X=first-level variable (age), Z=second-level variables (n=5; 4 dummy variables for schistosomiasis

exposure and 1 for rural-urban status),  $\mu_{0jk}$  = random effect (intercept) for cluster, and  $e_{00k}$  = random effect (intercept) for country.

## RESULTS

### 2009-2011 surveys (Table 3)

Of 43,772 women surveyed in the 2009-2011 Ethiopia, Kenya, Tanzania and Uganda DHS, 18,052 (41%) met inclusion criteria of married or in union at least 5 years and not using contraception. By t-test, neither marriage duration nor contraceptive use (selection criteria) was associated with schistosomiasis prevalence (exposure). Of the 18,052 at risk of pregnancy, 505 (2.8%) were excluded due to non-geocoded cluster locations (thus no exposure measurement). 16,861 households remained, with a mean of 1.04 women per household,

Of 17,547 women analyzed, 13.6% were currently pregnant, 3.2% had never given birth, and 64.8% had given birth within the past 5 years (Table 3). The inclusion criteria limited the total number under 20 years of age to 138; 92 of these were in Ethiopia, reflecting young marriages in that country. This number has decreased since the 1999-2001 survey. 19.7% reported ever have experienced a pregnancy loss (miscarriage or stillbirth). As over half of the data for gestational month of the loss were missing, differentiation by trimester could not be accurately determined.

35.2% of women were classified as infertile and 30.6% as infecund. Secondary infertility increased with age for all countries (7.5% of age 15-29, 22.7% of age 30-39, and 67.2% of age 40-49). Infecundity followed the same pattern at 6.6%, 20.7% and 63.6% by age category. This is the expected age pattern noted by other writers, reflecting natural declines in fertility and in frequency of intercourse. 3.2% of women had never given birth and 2.4% had never been pregnant (primary infertility and infecundity). Risk did not vary widely by age, as is expected of this measure of biological sterility.

81% lived in rural locations. The odds of infertility were higher for urban residents (OR 2.5 [95% CI 2.1, 3.0]), highest for Ethiopia (OR 3.4 [2.5, 4.7]).

### 1999-2001 surveys (Table 4)

Of 26,642 women surveyed in the 1999-2001 Ethiopia, Tanzania and Uganda DHS, 13,437 (50%) met inclusion criteria of married or in union at least 5 years and not using contraception. By t-test, neither marriage duration nor contraceptive use (selection criteria) was associated with schistosomiasis prevalence (exposure). Of the 13,437 at risk of pregnancy, 493 (3.7%) were excluded due to non-geocoded cluster locations (thus no exposure measurement). 12,197 households remained, with a mean of 1.06 study-eligible women per household.

Of 12,944 women analyzed, 12.6% were currently pregnant, 4.5% had never given birth, 65.0% had given birth within the past 5 years (Table 4). The inclusion criteria limited the total number under 20 years of age to 200; 168 of these were in Ethiopia. 21.9% reported ever have experienced a pregnancy loss (miscarriage or stillbirth). As over half of the data for gestational month of the loss were missing, differentiation by trimester could not be accurately determined.

35.0% of women were classified as infertile. Secondary infertility increased with age for all countries (7.2% of age 15-29, 23.6% of age 30-39, and 65.5% of age 40-49). This is the expected age pattern noted by other writers, reflecting natural declines in fertility and in frequency of intercourse. 4.5% of women had never given birth. Risk did not vary widely by age, as is expected of this measure of biological sterility. Infecundity was not measured in the 1999-2001 survey.

80% lived in rural locations. The odds of infertility were higher for urban residents (OR 3.1 [95% CI 2.5, 3.8]), highest for Ethiopia (OR 3.1 [2.4, 4.1]). Duration of residence was measured in the 1999-2001 surveys. 85.2% of subjects had lived at their current residence for at least five years, 91.9% for Ethiopia, 83.3% for Tanzania, and 66.7% for Uganda. Duration of residence was not associated with prevalence of either of the exposures of interest.

Age was not associated with either *S. haematobium* or *S. mansoni* prevalence for either survey set ( $R^2=0.001$ ). For each of the two survey sets, 14% of women lived in areas of high *S.*



*haematobium* prevalence, 48% in moderate, and 38% in low prevalence. 12% lived in areas of high *S. mansoni* prevalence, 50% in moderate, and 37% in low prevalence. Schistosomiasis prevalence varied somewhat with rural/urban classification, with high and low classifications of both species higher in urban sites. Age and rural/urban status were considered as potential confounders.

### **Spatial analysis of outcomes**

For 2009-2010, Moran's I (inverse distance weighting) for total (0.54), primary (0.24), and secondary (0.39) infertility and for total (0.53), primary (0.24), and secondary (0.42) infecundity indicated that these outcomes were autocorrelated (not randomly spatially distributed). Ever having experienced a pregnancy loss was weakly autocorrelated (Moran's I = 0.13). For 1999-2001, Moran's I for total (1.00), primary (0.30), and secondary (0.78) infertility indicated that these outcomes were autocorrelated (not randomly spatially distributed). Ever having experienced a pregnancy loss was very weakly autocorrelated (Moran's I = 0.07) (Table 5).

Visually, secondary infertility among younger rural women appeared to cluster in western and central Ethiopia; southwestern and south central Kenya; and southwestern, southeastern and island Tanzania. Primary infertility appeared to cluster in western and northern Ethiopia and in southwestern and island Tanzania. Ever having experienced a pregnancy loss appeared to cluster throughout Uganda, in central Ethiopia, and in central, lake, and island Tanzania (Figure 4). Additional maps illustrate the spatial distribution of outcomes as Local Moran's I Cluster and Outlier Analysis (Appendix, Table 6), as quartile or box maps (Appendix, Table 7), and as Univariate LISAs (Appendix, Table 8).

The Bivariate LISA method allows visualization of areas of high or low prevalence of one variable in relation to another. High secondary infertility is co-located with high *S. haematobium* prevalence in southwestern, southeastern, central and coastal Tanzania and coastal

Kenya; and with *S. mansoni* in central and southwestern Tanzania and western Uganda. Primary infertility is co-located with high *S. haematobium* prevalence in lake, southwest, and coastal Tanzania, coastal Kenya, and northeastern Ethiopia; and with *S. mansoni* in lake and southwest Tanzania and western Uganda. History of pregnancy loss is co-located with high *S. haematobium* prevalence in lake, south, and coastal Tanzania and coastal Kenya, and with *S. mansoni* in lake and southwest Tanzania and western Uganda (Figure 5).

### Regression analysis

Results are presented in Tables 6 and 7. For brevity, survey rounds 1999-2001 are referred to as "2000" and rounds 2009-2011 as "2010".

For total infertility for 2010, the intraclass correlation coefficient (ICC) for DHS cluster was 0.108 and for country 0.003, indicating that 10.8% of variability in infertility existed between clusters, 0.3% between countries and 88.9% between individuals. For 2000, the intraclass correlation coefficient (ICC) for DHS cluster was 0.073 and for country 0.012, indicating that 7.3% of variability in infertility existed between clusters, 1.2% between countries and 91.5% between individuals.

Women living in high compared to absent *S. haematobium* locations had significantly higher odds of total infertility (2000: [OR 1.53, 95%ci 1.27, 1.85], 2010: [OR 1.24, 95%ci 1.06, 1.47]), of secondary infertility (2000: [OR 1.77, 95%ci 1.43, 2.18], 2010: OR 1.27, 95%ci 1.07, 1.52]), and of primary infertility (2000: [OR 1.61, 95%ci 1.10, 2.37], 2010: [OR 1.53, 95%ci 1.05, 2.25]) for both survey rounds. Women living in moderate compared to absent *S. haematobium* locations had significantly higher odds of total infertility [OR 1.31, 95%ci 1.15, 1.49]), and of secondary infertility [OR 1.27, 95%ci 1.07, 1.52]for 2010 and of primary infertility for 2000: [OR 1.56, 95%ci 1.14, 2.14]. Living in high or moderate *S. mansoni* locations was not associated with the odds of any infertility outcome.

To control for any unidentified confounders relevant to presence or absence of any schistosomiasis (for example, low, warm, humid location compared to high, cooler, dryer location), residence in high *S. haematobium* compared to high *S. mansoni* locations was analyzed, as no relevant variables determining which species is present have been identified (see Introduction). Women living in high *S. haematobium* compared to high *S. mansoni* locations had significantly higher odds of total infertility (2000: [OR 1.41, 95% ci 1.07, 1.85], 2010: [OR 1.44, 95% ci 1.14, 1.81]), of secondary infertility (2000: [OR 1.60, 95% ci 1.17, 2.17], 2010: [OR 1.52, 95% ci 1.18, 1.94]), and of primary infertility for 2010: [OR 2.83, 95% ci 1.60, 5.00]).

Ever having experienced a pregnancy loss was less consistently associated with schistosomiasis exposure status. Significant associations were with high compared to absent *S. haematobium* in 2000 (OR 1.23, 95% ci 1.04, 1.45) and with high *S. haematobium* compared to high *S. mansoni* in 2000 (OR 1.32, 95% ci 1.03, 1.69).

As duration of residence at current location was measured in the 1999-2001 surveys, some assessment of temporal factors was possible. Assuming a 5 year lag time between exposure to the parasite and occurrence of tubal disease and an additional lag time of the 5 years exposure to pregnancy necessary to declare infertility, associations were estimated for those locally resident for 5 years and for 10 years (Table 8). Women living in high compared to absent *S. haematobium* locations for at least 10 years had significantly increased odds of total infertility (OR 1.47, 95% ci 1.19, 1.81), of secondary infertility (OR 1.74, 95% ci 1.38, 2.19), and of pregnancy loss (OR 1.32, 95% ci 1.10, 1.59). Women living for at least 10 years in high *S. haematobium* compared to high *S. mansoni* locations had significantly higher odds of total infertility (OR 1.35, 95% ci 1.01, 1.83), secondary infertility (OR 1.55, 95% ci 1.11, 2.18), and of pregnancy loss (OR 1.35, 95% ci 1.03, 1.78).

As age at exposure to schistosomiasis may be a factor in disease occurrence, associations were also estimated for those residing locally since before age 10. Women living in high compared to absent *S. haematobium* locations since before age 10 had significantly increased

odds of total infertility (OR 1.69, 95% ci 1.31, 2.17), of secondary infertility (OR 2.11, 95% ci 1.59, 2.80), and of pregnancy loss (OR 1.38, 95% ci 1.11, 1.73). Women living for at least 10 years in high *S. haematobium* compared to high *S. mansoni* locations had significantly higher odds of total infertility (OR 1.66, 95% ci 1.15, 2.40) and of secondary infertility (OR 2.11, 95% ci 1.39, 3.22).

Other researchers have excluded the youngest (<20) or oldest (45-50) age groups in infertility classifications. Sensitivity analysis of various age restrictions and country groupings consistently found total and secondary infertility and (for 2009-2011) infecundity to be significantly associated with residence in high compared to absent *S. haematobium* locations and in high *S. haematobium* compared to high *S. mansoni* locations. Associations were less consistent for moderate compared to absent *S. haematobium* and for primary infertility, with confidence intervals for some comparisons including 1. (Table 8)

Population Attributable Fraction (PAF) is a measure of excess risk of an outcome in relation to an exposure; use of the term Etiologic Fraction for this measure is avoided due to the implication of causation. By PAF, for 2000, 7.0% [4.3, 9.4] of all infertility cases were associated with residence in high *S. haematobium* and 5.6% [0, 12.0] with residence in moderate *S. haematobium* areas. For 2010, 4.5% [1.2, 7.2] of all infertility cases were associated with residence in high *S. haematobium* and 11.0% [6.3, 15.4] with residence in moderate *S. haematobium* areas. For 2000, 7.8% [1.9, 11.8] of primary infertility cases were associated with residence in high *S. haematobium* and 15.2% [3.7, 23.8] with residence in moderate *S. haematobium* areas. For 2009-2011, 8.7% [1.1, 13.8] of primary infertility cases were associated with residence in high *S. haematobium* areas.

## DISCUSSION

Risks of primary, secondary, and total infertility (births) and infecundity (conceptions) were significantly associated with residence in areas of high *S. haematobium* prevalence, compared to both *S. haematobium* absence and to equivalent *S. mansoni* prevalence. Levels of *S. mansoni* prevalence were not associated with any impaired fertility outcomes. This suggests that the association is related not to unmeasured confounders of presence/absence of schistosomiasis, but to the differing clinical manifestations of the two *Schistosoma* species (i.e., the tubal or other urogenital damage of *S. haematobium* rather than the hepatic damage of *S. mansoni*).

There does not appear to be an ordinal effect of the exposure, as odds ratios did not differ proportionally for high and for moderate *S. haematobium* prevalence. When exposure was dichotomized at 5% (or present/absent), odds ratios for total infertility with any *S. haematobium* exposure did not differ markedly from those for the high and moderate categories (1999-2001: OR 1.25, 95%ci 1.09, 1.43; 2009-2011: OR 1.29, 95%ci 1.15, 1.46). This supports the argument that the usual prevalence measure (proportion of sampled schoolchildren excreting eggs on a given day) while useful as a measure of transmission, does not accurately measure prevalence of disease, which is cumulative and persistent.

History of pregnancy losses (stillbirth or miscarriage), which have many possible etiologies other than genital tract disease, was not consistently associated with schistosomiasis exposure status.

Strengths of the study were the regional approach, large sample size, and standardized data collection and analysis methods available by combining national Demographic and Health Surveys. Another strength was the public accessibility of large parasitic disease data compilations supporting generation of highly predictive interpolated spatial models of disease prevalence. The ability to use *S. mansoni* exposure as a negative control for *S. haematobium* exposure was useful, as there may have been unknown environmental factors confounding the comparison only of high to low prevalence within each species.

A limitation inherent to the cross-sectional study design is the difficulty assessing temporal factors. The 1999-2001 surveys included a variable for duration of current residence, allowing some temporal measures of exposure. Associations for those living in high-exposure regions for at least 5 years or 10 years (accounting for lag time from exposure to measured outcome) did not appear to differ meaningfully from those for the total survey population. This could be explained if previous residences were within the same exposure zone, such as neighboring villages. No data were available for exposure classifications of previous residence or of reasons for moves, such as possible urban migration of infertile women. Migration from a high-exposure to a low-exposure location could result in misclassification of exposed as unexposed, although the reverse would not occur, due to the cumulative nature of exposure. Age of exposure may be important; associations for total and secondary infertility were strengthened for those exposed before age 10. The 2009-2011 surveys lacked this temporal exposure measure, but included a variable for calendar time of previous pregnancy losses, allowing inclusion of all pregnancies, not just those resulting in birth, in outcome measures (infecundity as well as infertility).

Exposure was measured not individually but ecologically, based on current residence. Individual exposure to schistosomiasis within an endemic area varies with lifetime water exposure behaviors as well as focal distribution of infected snails within an ecological zone. Urogenital schistosomiasis lesions occur within a few years of exposure and persist even with anti-helminthic treatment. Thus disease (infertility due to tubal damage) may reflect either remote childhood exposure or recent migration into an endemic area.

As exposure was measured by predicted point prevalence using the same interpolation method for each *Schistosoma* species, any misclassification of exposure was unlikely to be differential by type of schistosomiasis. In sensitivity analysis, various categorization cut-points were tested, with variation in significance of results primarily related to smaller sample sizes (Tables 8 & 9). Misclassification of infertility was unlikely, as DHS collects detailed data on all

births and previous studies have assessed the accuracy of this measurement. Infertility, the major outcome assessed, measures livebirths only. Conceptions (livebirths, stillbirths, and miscarriages) would be a more specific indicator of Fallopian tubal disease, but infecundity was measurable in only the 2009-2011 survey round. Misclassification of history of pregnancy losses (miscarriage or stillbirth) or of infecundity (which incorporates pregnancy losses) may be high due to both reporting bias and recognition bias. Possible misclassification of the rural-urban co-variate was previously discussed. However, none of these is likely to be differential by exposure.

Survival bias may be a factor if ectopic pregnancy, the most severe form of tubal disease, is associated with *S. haematobium* exposure. As DHS surveys only living women, any women who have died from ruptured ectopic pregnancy, a highly likely outcome in the study setting, would not be counted as either cases or subjects, potentially biasing study results toward the null.

This study addresses infertility as a female outcome. Although *S. haematobium* may damage male as well as female reproductive organs, the role of infertility and exposure status of the of the male partner was not addressed.

The association with schistosomiasis increases the evidence that impaired fertility may have etiologies other than sexually transmitted infections and should not be equated with stigmatized sexual behaviors. The data sources and methods provide an approach to studying the intersection of two disparate fields: Neglected Tropical Diseases and Maternal-Child Health. These results contribute to the mounting evidence of the adverse effects of schistosomiasis on women's health, supporting the need to ensure inclusion of girls and women in disease control interventions.

### **SECTION III: PUBLIC HEALTH IMPLICATIONS AND FUTURE DIRECTIONS**

These results may be confirmed by applying the same approach to a different region, perhaps West Africa or southern Africa. The approach of combining geo-referenced DHS data with large tropical infectious disease prevalence datasets may be applied to other possibly spatially-distributed DHS-measurable outcomes (anemia, other maternal and child health indicators) and other spatially-distributed infectious diseases (malaria, lymphatic giardiasis, other vector-borne diseases). Other methods should be explored for assessing possible associations of schistosomiasis with important women's reproductive outcomes of maternal mortality or intervening conditions, such as *S. haematobium* with prolepsis and *S. mansoni* with coagulopathy.



## TABLES

**Table 1. Published reports of infertility associated with schistosomiasis.**

| Author                    | Country                    | Type of study    | Species | Number  |
|---------------------------|----------------------------|------------------|---------|---------|
| Adenitis 2001 (113)       | Nigeria                    | pathology series | NS      | 2 of 7  |
| Bailey 2010 (114)         | UK, Africa travel          | case reports     | Sh      | 2       |
| Balasko 1995 (115)        | Spain ex Nigeria           | case report      | Sh      | 1       |
| Billy-Bris sac 1994 (116) | Guadeloupe                 | case report      | Sm      | 1       |
| Bland 1970 (117)          | Rhodesia                   | path series      | Sh      | 3 of 10 |
| Boulez 1964 (118)         | South Africa               | path series      | NS      | 14      |
| Furlough 1976 (119)       | Malawi                     | pathology series | Sh      | **      |
| Cornier 1981 (120)        | France ex Tunisia          | case report      | Sh      | 1       |
| Crump 2000 (121)          | New Zealand, Africa travel | case report      | NS      | 1       |
| DeMille 1995 (122)        | USA ex Liberia             | case report      | Sh      | 1       |
| Doug 1973 (123)           | Senegal                    | case series      | NS      | 4       |
| Ekoukou 1995 (124)        | France ex Senegal          | case report      | Sh      | 1       |
| El-Mahgoub 1982 (125)     | Egypt                      | case series      | NS      | 13      |
| El_maraghy 1982 (126)     | Egypt                      | case series      | NS      | 10      |
| Gilbert 1943 (127)        | Southern Rhodesia          | case series      | Sh      | 2       |
| Harouny 1988 (128)        | Kuwait                     | case report      | Sh      | 1       |
| Hoffmann 2004 (129)       | Germany ex Angola          | case report      | Sh      | 1       |
| Krolikowski 1995 (130)    | South Africa               | case report      | NS      | 1       |
| LeGuyader 1965 (131)      | France                     | case report      | Sh      | 1       |
| Morice 1996 (132)         | France ex Africa           | case reports     | NS      | 3       |
| Mouktar 1966 (133)        | Egypt                      | case reports     | NS      | 2       |
| Nouhou 1998 (134)         | Niger                      | case series      | Sh      | 6       |
| O'Leary 1994 (135)        | USA ex Zimbabwe            | case report      | Sh      | 1       |
| Ogunniyi 1994 (136)       | Nigeria                    | case report      | Sh      | 1       |
| Picaud 1990 (137)         | Gabon                      | case report      | Si      | 1       |
| Schanz 2013 (138)         | Germany ex Nigeria         | case report      | NS      | 1       |
| Schroers 1995 (139)       | Germany ex Togo            | case report      | Sh      | 1       |
| Sheorey 2004 (140)        | Australia, Africa ravel    | case report      | Sh      | 1       |
| Swai 2006 (141)           | Tanzania                   | pathology series | NS      | 4 of 7  |
| Vass 1982 (142)           | UK ex Malawi               | case report      | Sh      | 1       |

NS = not specified  
Sh = *Schistosoma haematobium*  
Si = *S. intercalatum*  
Sm = *S. mansoni*  
\*\* 57 of 138 infertile had Sh, 6 had Sm; 13 of 42 spontaneous abortions had schistosomiasis

acknowledgement: Eleanor Friedman, replication of search and data abstraction

**Table 2. Published reports of ectopic pregnancy associated with schistosomiasis**

| <b>Author</b>                         | <b>Country</b>     | <b>Type of study</b> | <b>Species</b> | <b>Number</b> |
|---------------------------------------|--------------------|----------------------|----------------|---------------|
| Aminu 2014 (143)                      | Nigeria            | case report          | NS             | 1             |
| Bahrami 2006 (144)                    | USA ex East Africa | case report          | Sh             | 1             |
| Bland 1970 (117)                      | Rhodesia           | pathology series     | NS             | 6 of 38       |
| Bugalho 1991 (145)                    | Mozambique         | case reports         | NS             | 4             |
| Doug 1973 (123)                       | Senegal            | case report          | NS             | 1             |
| Ekoukou 1995 (124)                    | France ex Senegal  | case report          | Sh             | 1             |
| El-Bedri 1958 (146)                   | Egypt              | case report          | Sh             | 1             |
| Eogan 2002 (147)                      | Ireland ex Nigeria | case report          | Sh             | 1             |
| Garba 2004 (148)                      | Niger              | case report          | Sh             | 1             |
| Gilbert 1943(127)                     | Southern Rhodesia  | case series          | Sh             | 1             |
| Hassim 1966 (149)                     | Zambia             | case report          | Sh             | 1             |
| Laxman 2008 (150)                     | UK ex Zambia       | case report          | Sh             | 1             |
| Mayat 1959 (151)                      | South Africa       | case report          | NS             | 1             |
| Mohammed 2004 (152)                   | Nigeria            | case report          | Sh             | 1             |
| Okonufua 1990 (153)                   | Nigeria            | case report          | Sh             | 1             |
| Owusu-Bempah 2013 (154)               | Ghana              | case report          | NS             | 1             |
| Sahu 2013 (155)                       | India ex Africa    | case report          | Sh             | 1             |
| Scheller 1974 (156)                   | Tanzania           | case series          | Sh             | 37 of 93      |
| Schneider 2000 (157)                  | South Africa       | case report          | Sh             | 1             |
| Ville 1991 (158)                      | Gabon              | case reports         | Sh             | 3             |
| NS = not specified                    |                    |                      |                |               |
| Sh = <i>Schistosoma haematobium</i>   |                    |                      |                |               |
| ***6 of 38 ectopic pregnancies had Sh |                    |                      |                |               |

**Table 3. Population description, Demographic and Health Surveys Ethiopia 2000, Tanzania 1999, Uganda 2001.**

| 1999-2001                            | Ethiopia       | Tanzania       | Uganda         | Total          |
|--------------------------------------|----------------|----------------|----------------|----------------|
| Number                               | 8357           | 1747           | 2840           | 12944          |
|                                      | N (%)          | N (%)          | N (%)          | N (%)          |
| Age 15-19                            | 168 (2.0)      | 6 (0.3)        | 24 (0.8)       | 200 (1.5)      |
| 20-29                                | 2736 (32.7)    | 610 (34.9)     | 1295 (45.6)    | 4647 (35.9)    |
| 30-39                                | 3087 (36.9)    | 685 (39.2)     | 1050 (37.0)    | 4822 (37.3)    |
| 40-49                                | 2366 (28.3)    | 457 (26.2)     | 652 (22.9)     | 3475 (26.8)    |
| Currently pregnant                   | 949 (11.4)     | 232 (13.3)     | 440 (15.8)     | 1679 (12.6)    |
| Never given birth                    | 424 (5.1)      | 71 (4.1)       | 94 (3.3)       | 589 (4.5)      |
| Ever had miscarriage or stillbirth   | 1562 (18.7)    | 506 (29.0)     | 762 (26.9)     | 2830 (21.9)    |
| Infertile (no birth in last 5 years) | 2828 (33.8)    | 684 (39.2)     | 875 (30.8)     | 4534 (35.0)    |
| Primary (never given birth)          | 392 (3.7)      | 66 (3.8)       | 93 (3.3)       | 551 (4.3)      |
| age 15-29                            | 236 (8.1)      | 27 (4.5)       | 24 (2.0)       | 287 (6.2)      |
| age 30-39                            | 94 (3.0)       | 32 (4.7)       | 37 (3.5)       | 163 (3.4)      |
| age 40-49                            | 62 (2.6)       | 7 (1.5)        | 32 (4.9)       | 101 (2.9)      |
| Secondary (previous birth)           | 2436 (29.1)    | 578 (33.0)     | 747 (26.3)     | 3761 (29.1)    |
| age 15-29                            | 192 (6.6)      | 65 (10.7)      | 81 (7.0)       | 338 (7.2)      |
| age 30-39                            | 707 (22.9)     | 196 (28.6)     | 240 (22.6)     | 1143 (23.6)    |
| age 40-49                            | 1537 (65.0)    | 317 (69.4)     | 426 (64.7)     | 2280 (65.5)    |
| Current residence > 5 years          | 7679 (91.9)    | 1456 (83.3)    | 1895 (66.73)   | 11030 (85.2)   |
| Rural                                | 6930 (82.4)    | 1293 (72.8)    | 2543 (78.1)    | 10766 (80.1)   |
|                                      |                |                |                |                |
|                                      | OR (95%CI)     | OR (95%CI)     | OR (95%CI)     | OR (95%CI)     |
| Odds of infertility, urban vs rural  | 3.6 (3.2, 4.1) | 2.3 (1.8, 2.8) | 2.4 (2.0, 2.8) | 3.0 (2.7, 3.3) |

**Table 4. Population description, Demographic and Health Surveys Ethiopia 2011, Kenya 2009, Tanzania 2001, Uganda 2011.**

| 2009-2011                              | Ethiopia     | Kenya        | Tanzania     | Uganda       | Total        |
|--|--------------|--------------|--------------|--------------|--------------|
| Number                                 | 7314         | 2758         | 3945         | 3530         | 17547        |
|  | N (%)        | N (%)        | N (%)        | N (%)        | N (%)        |
| Age 15-19                              | 92 (1.3)     | 13 (0.5)     | 9 (0.2)      | 24 (0.7)     | 138 (0.7)    |
| 20-29                                  | 2497 (34.1)  | 867 (31.4)   | 1125 (28.6)  | 168 (35.9)   | 5757 (32.8)  |
| 30-39                                  | 2730 (37.3)  | 1045 (37.9)  | 1556 (39.4)  | 1292 (37.2)  | 6634 (37.9)  |
| 40-49                                  | 1995 (27.3)  | 833 (30.2)   | 1255 (31.8)  | 926 (26.2)   | 5009 (28.6)  |
| Currently pregnant                     | 895 (12.2)   | 347 (12.6)   | 581 (14.7)   | 566 (16.0)   | 2389 (13.6)  |
| Never given birth                      | 313 (4.3)    | 54 (20.0)    | 128 (3.2)    | 59 (1.7)     | 554 (3.2)    |
| Ever had miscarriage/stillbirth (loss) | 974 (13.3)   | 457 (16.6)   | 1032 (26.2)  | 997 (28.2)   | 3460 (19.7)  |
| Infertile (no birth in last 5 years)   | 2600 (35.6)  | 1107 (40.1)  | 1461 (37.0)  | 1015 (28.8)  | 6183 (35.2)  |
| Primary (never given birth)            | 285 (3.9)    | 50 (1.8)     | 124 (3.1)    | 54 (1.5)     | 513 (2.9)    |
| age 15-29                              | 152 (5.9)    | 14 (1.2)     | 27 (2.4)     | 14 (1.1)     | 207 (3.5)    |
| age 30-39                              | 79 (2.9)     | 20 (1.9)     | 59 (3.8)     | 21 (1.6)     | 179 (2.7)    |
| age 40-49                              | 54 (2.7)     | 16 (1.9)     | 38 (3.0)     | 19 (2.0)     | 127 (2.5)    |
| Secondary (previous birth)             | 2166 (29.6)  | 935 (33.9)   | 1247 (31.6)  | 917 (26.0)   | 5318 (30.3)  |
| age 15-29                              | 216 (8.3)    | 69 (7.8)     | 86 (7.6)     | 72 (5.6)     | 443 (7.5)    |
| age 30-39                              | 624 (22.9)   | 299 (28.6)   | 353 (22.7)   | 234 (17.8)   | 1510 (22.7)  |
| age 40-49                              | 1326 (66.5)  | 620 (74.4)   | 808 (64.4)   | 611 (66.0)   | 3365 (67.2)  |
| Infecund (no birth or loss in 5 years) | 2507 (34.3)  | 970 (35.2)   | 1232 (31.2)  | 855 (24.2)   | 5364 (30.6)  |
| Primary (never conceived)              | 251 (3.4)    | 32 (1.2)     | 88 (2.2)     | 42 (1.2)     | 413 (2.4)    |
| age 15-29                              | 136 (5.2)    | 7 (0.8)      | 21 (1.8)     | 9 (0.3)      | 173 (2.9)    |
| age 30-39                              | 71 (2.6)     | 14 (1.3)     | 40 (2.6)     | 18 (1.4)     | 143 (2.2)    |
| age 40-49                              | 44 (2.2)     | 11 (1.3)     | 27 (2.2)     | 15 (1.6)     | 97 (1.9)     |
| Secondary (previous conception)        | 2256 (28.1)  | 938 (34.0)   | 1144 (29.0)  | 813 (23.0)   | 4951 (28.2)  |
| age 15-29                              | 199 (7.5)    | 63 (7.2)     | 76 (6.7)     | 53 (4.1)     | 391 (6.6)    |
| age 30-39                              | 586 (21.5)   | 274 (26.2)   | 311 (20.0)   | 203 (15.5)   | 1374 (20.7)  |
| age 40-49                              | 1271 (63.7)  | 601 (72.2)   | 757 (60.3)   | 557 (60.2)   | 3186 (63.6)  |
| Rural                                  | 6024 (82.4)  | 2146 (77.8)  | 3319 (81.6)  | 2848 (80.1)  | 14237(81.1)  |
|  |              |              |              |              |              |
|  | OR (95%CI)   | OR (95%CI)   | OR (95%CI)   | OR (95%CI)   | OR (95%CI)   |
| Odds of infertility, urban vs rural    | 3.4(2.5,4.7) | 2.2(1.4,3.6) | 2.0(1.5,2.6) | 1.9(1.5,2.4) | 2.5(2.1,3.0) |

**Table 5. Measure of autocorrelation of outcome variables (impaired fertility), Moran's I (inverse distance weighting). Demographic and Health Surveys, Ethiopia 2000 & 2011, Kenya 2009, Tanzania 1999 & 2001, Uganda 2001 & 2011.**

|                         | Moran's I* |
|-------------------------|------------|
| 1999-2001               |            |
| Infertility (total)     | 1.0110     |
| Infertility (primary)   | 0.2966     |
| Infertility (secondary) | 0.7817     |
| Pregnancy loss          | 0.0674     |
|                         |            |
| 2009-2011               |            |
| Infertility (total)     | 0.5413     |
| Infertility (primary)   | 0.2412     |
| Infertility (secondary) | 0.3925     |
| Infecundity (total)     | 0.5251     |
| Infecundity (primary)   | 0.2438     |
| Infecundity (secondary) | 0.4178     |
| Pregnancy loss          | 0.1276     |
| *all $p < 0.01$         |            |

**Table 6. Association of impaired fertility with schistosomiasis endemicity of residence location, women age 15-50, Demographic and Health Surveys Ethiopia 2000, Tanzania 1999, Uganda 2001.**

| DHS 1999-2001   | OR   | 95% CI     | PAF   | 95% CI       |
|---|------|------------|-------|--------------|
| Total infertility   |      |            |       |              |
| <i>S. h</i> high (ref=absent)   | 1.53 | 1.27, 1.85 | 0.070 | 0.043, 0.093 |
| <i>S. h</i> moderate  | 1.14 | 0.99, 1.32 |       |              |
| <i>S. m</i> high  | 1.09 | 0.89, 1.33 |       |              |
| <i>S. m</i> moderate  | 1.01 | 0.87, 1.17 |       |              |
| <i>S. h</i> high (ref= <i>S. m</i> high)  | 1.43 | 1.08, 1.90 |       |              |
| Secondary infertility   |      |            |       |              |
| <i>S. h</i> high (ref=absent)   | 1.77 | 1.43, 2.18 | 0.088 | 0.061, 0.110 |
| <i>S. h</i> moderate  | 1.09 | 0.92, 1.28 |       |              |
| <i>S. m</i> high  | 1.11 | 0.87, 1.40 |       |              |
| <i>S. m</i> moderate  | 0.97 | 0.82, 1.15 |       |              |
| <i>S. h</i> high (ref= <i>S. m</i> high)  | 1.60 | 1.17, 2.17 |       |              |
| Primary infertility   |      |            |       |              |
| <i>S. h</i> high (ref=absent)   | 1.78 | 1.61, 2.37 | 0.078 | 0.019, 0.118 |
| <i>S. h</i> moderate  | 1.47 | 1.08, 2.00 | 0.152 | 0.037, 0.238 |
| <i>S. m</i> high  | 1.12 | 0.73, 1.73 |       |              |
| <i>S. m</i> moderate  | 1.25 | 0.92, 1.70 |       |              |
| <i>S. h</i> high (ref= <i>S. m</i> high)  | 1.44 | 0.82, 2.53 |       |              |
| Pregnancy loss  |      |            |       |              |
| <i>S. h</i> high (ref=absent)   | 1.23 | 1.04, 1.45 |       |              |
| <i>S. h</i> moderate  | 0.87 | 0.77, 1.00 |       |              |
| <i>S. m</i> high  | 0.93 | 0.81, 1.12 |       |              |
| <i>S. m</i> moderate  | 0.92 | 0.81, 1.05 |       |              |
| <i>S. h</i> high (ref= <i>S. m</i> high)  | 1.32 | 1.03, 1.69 |       |              |
| <p><i>S. h</i> = <i>Schistosoma haematobium</i><br/> <i>S. m</i> = <i>Schistosoma mansoni</i><br/> PAF = Population Attributable Fraction<br/> N = 12944 clusters N = 973 model AIC = 12420<br/> Method: weighted multi-level mixed logistic regression</p> |      |            |       |              |

**Table 7. Association of impaired fertility with schistosomiasis endemicity of residence, women age 15-50, Demographic and Health Surveys Ethiopia2011, Kenya2009, Tanzania2001, Uganda2011.**

| DHS 2009-2011  | OR   | 95% CI     | PAF   | 95% CI        |
|--|------|------------|-------|---------------|
| Total infertility  |      |            |       |               |
| <i>S. h</i> high (ref=absent)  | 1.24 | 1.06, 1.47 | 0.045 | 0.012, 0.072  |
| <i>S. h</i> moderate   | 1.31 | 1.15, 1.49 | 0.111 | 0.063, 0.154  |
| <i>S. m</i> high   | 0.86 | 0.73, 1.02 |       |               |
| <i>S. m</i> moderate   | 1.02 | 0.90, 1.16 |       |               |
| <i>S. h</i> high (ref= <i>S. m</i> high)   | 1.44 | 1.14, 1.81 |       |               |
| Secondary infertility  |      |            |       |               |
| <i>S. h</i> high (ref=absent)  | 1.27 | 1.07, 1.52 | 0.048 | 0.014, 0.077  |
| <i>S. h</i> moderate   | 1.35 | 1.18, 1.55 | 0.123 | 0.072, 0.167  |
| <i>S. m</i> high   | 0.84 | 0.70, 1.00 |       |               |
| <i>S. m</i> moderate   | 0.97 | 0.85, 1.11 |       |               |
| <i>S. h</i> high (ref= <i>S. m</i> high)   | 1.52 | 1.18, 1.94 |       |               |
| Primary infertility  |      |            |       |               |
| <i>S. h</i> high (ref=absent)  | 1.53 | 1.05, 2.25 | 0.087 | 0.011, 0.139  |
| <i>S. h</i> moderate   | 1.34 | 0.99, 1.81 |       |               |
| <i>S. m</i> high   | 0.54 | 0.35, 0.84 |       |               |
| <i>S. m</i> moderate   | 1.31 | 0.98, 1.75 |       |               |
| <i>S. h</i> high (ref= <i>S. m</i> high)   | 2.83 | 1.60, 5.00 |       |               |
| Total infecundity  |      |            |       |               |
| <i>S. h</i> high (ref=absent)  | 1.25 | 1.05, 1.47 | 0.045 | 0.011, 0.0673 |
| <i>S. h</i> moderate   | 1.35 | 1.19, 1.54 | 0.123 | 0.075, 0.166  |
| <i>S. m</i> high   | 0.87 | 0.73, 1.02 |       |               |
| <i>S. m</i> moderate   | 1.05 | 0.92, 1.19 |       |               |
| <i>S. h</i> high (ref= <i>S. m</i> high)   | 1.36 | 1.07, 1.72 |       |               |
| Secondary infecundity  |      |            |       |               |
| <i>S. h</i> high (ref=absent)  | 1.29 | 1.07, 1.54 | 0.048 | 0.014, 0.076  |
| <i>S. h</i> moderate   | 1.38 | 1.20, 1.59 | 0.124 | 0.073, 0.168  |
| <i>S. m</i> high   | 0.88 | 0.73, 1.06 |       |               |
| <i>S. m</i> moderate   | 1.00 | 0.87, 1.14 |       |               |
| <i>S. h</i> high (ref= <i>S. m</i> high)   | 1.47 | 1.14, 1.89 |       |               |
| Primary infecundity  |      |            |       |               |
| <i>S. h</i> high (ref=absent)  | 1.33 | 0.86, 2.06 |       |               |
| <i>S. h</i> moderate   | 1.28 | 0.91, 1.80 |       |               |
| <i>S. m</i> high   | 0.66 | 0.40, 1.08 |       |               |
| <i>S. m</i> moderate   | 1.46 | 1.05, 2.04 |       |               |
| <i>S. h</i> high (ref= <i>S. m</i> high)   | 2.01 | 1.06, 3.84 |       |               |
| Pregnancy loss   |      |            |       |               |
| <i>S. h</i> high (ref=absent)  | 1.14 | 0.97, 1.33 |       |               |
| <i>S. h</i> moderate   | 0.81 | 0.72, 0.92 |       |               |
| <i>S. m</i> high   | 1.00 | 0.85, 1.18 |       |               |
| <i>S. m</i> moderate   | 0.88 | 0.78, 0.99 |       |               |
| <i>S. h</i> high (ref= <i>S. m</i> high)   | 1.13 | 0.91, 1.41 |       |               |
| <p><i>S. h</i> = <i>Schistosoma haematobium</i><br/> <i>S. m</i> = <i>Schistosoma mansoni</i><br/> PAF = Population Attributable Fraction<br/> N = 17547 clusters N = 1827 model AIC=15606<br/> Method: weighted multi-level mixed logistic regression</p> |      |            |       |               |

**Table 8. Association of impaired fertility with schistosomiasis endemicity of residence, Demographic and Health Surveys Ethiopia 2000, Tanzania 1999, Uganda 2001. Sensitivity analysis, various age restrictions and exposure categories.**

| 1999-2001                        | <i>S.h</i> high (ref=0) | <i>S.h</i> mod    | <i>S.h</i> high (ref= <i>S.m</i> high) |
|----------------------------------|-------------------------|-------------------|--|
| age restricted                   | OR (95%ci)              | OR (95%ci)        | OR (95%ci)                             |
| <b>total infertility</b>         |                         |                   |  |
| <40                              | 1.62 (1.31, 2.01)       | 1.17 (0.99, 1.39) | 1.47 (1.07, 2.01)                      |
| >20                              | 1.62 (1.34, 1.97)       | 1.15 (0.99, 1.34) | 1.46 (1.11, 1.94)                      |
| 20-45                            | 1.69 (1.38, 2.06)       | 1.20 (1.03, 1.40) | 1.54 (1.15, 2.06)                      |
| <b>secondary infertility</b>     |                         |                   |  |
| <40                              | 2.10 (1.60, 2.75)       | 1.11 (0.89, 1.38) | 1.82 (1.22, 2.72)                      |
| >20                              | 1.79 (1.45, 2.21)       | 1.10 (0.93, 1.30) | 1.61 (1.18, 2.20)                      |
| 20-45                            | 1.97 (1.56, 2.48)       | 1.17 (0.97, 1.41) | 1.77 (1.26, 2.48)                      |
| <b>primary infertility</b>       |                         |                   |  |
| <40                              | 1.97 (1.30, 2.98)       | 1.48 (1.05, 2.07) | 1.66 (0.90, 3.05)                      |
| >20                              | 1.88 (1.26, 2.81)       | 1.54 (1.11, 2.13) | 1.56 (0.87, 1.80)                      |
| 20-45                            | 2.10 (1.39, 3.19)       | 1.51 (1.07, 2.12) | 1.60 (0.87, 2.95)                      |
| <b>pregnancy loss</b>            |                         |                   |  |
| <40                              | 1.40 (1.15, 1.70)       | 0.88 (0.76, 1.03) | 1.57 (1.18, 2.10)                      |
| >20                              | 1.24 (1.05, 1.47)       | 0.88 (0.77, 1.00) | 1.32 (1.03, 1.68)                      |
| 20-45                            | 1.36 (1.13, 1.63)       | 0.88 (0.76, 1.02) | 1.47 (1.13, 1.92)                      |
| <b>exposure cut-points</b>       |                         |                   |  |
| <b>total infertility</b>         |                         |                   |  |
| 30%, 5%                          | 1.32 (1.07, 1.64)       | 1.21 (1.05, 1.40) | 1.16 (0.83, 1.60)                      |
| 5% (dichot)                      | 1.25 (1.09, 1.43)       | NA                | 1.24 (1.02, 1.52)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.53 (1.27, 1.85)       | 1.15 (1.00, 1.33) | NA                                     |
| <b>secondary infertility</b>     |                         |                   |  |
| 30%, 5%                          | 1.34 (1.05, 1.71)       | 1.17 (0.99, 1.37) | 1.18 (0.82, 1.0)                       |
| 5% (dichot)                      | 1.22 (1.05, 1.42)       | NA                | 1.30 (1.04, 1.63)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.60 (1.29, 1.97)       | 1.09 (0.93, 1.28) | NA                                     |
| <b>primary infertility</b>       |                         |                   |  |
| 30%, 5%                          | 1.26 (0.84, 1.89)       | 1.37 (1.05, 1.80) | 1.14 (0.62, 2.10)                      |
| 5% (dichot)                      | 1.35 (1.05, 1.74)       | NA                | 1.22 (0.83, 1.79)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.39 (0.98, 1.98)       | 1.35 (1.03, 1.77) | NA                                     |
| <b>pregnancy loss</b>            |                         |                   |  |
| 30%, 5%                          | 1.23 (1.01, 1.49)       | 0.90 (0.79, 1.03) | 1.28 (0.95, 1.73)                      |
| 5% (dichot)                      | 0.97 (0.86, 1.10)       | NA                | 1.08 (0.90, 1.30)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.23 (1.04, 1.46)       | 0.87 (0.76, 0.99) | NA                                     |
| <b>residence duration</b>        |                         |                   |  |
| <b>total infertility</b>         |                         |                   |  |
| >10y                             | 1.47 (1.19, 1.81)       | 1.13 (0.96, 1.33) | 1.35 (1.00, 1.83)                      |
| >5y                              | 1.47 (1.20, 1.80)       | 1.11 (0.95, 1.30) | 1.36 (1.01, 1.82)                      |
| age 1st expos                    | 1.69 (1.31, 2.17)       | 1.18 (0.96, 1.44) | 1.66 (1.15, 2.40)                      |
| <b>secondary infertility</b>     |                         |                   |  |
| >10y                             | 1.74 (1.38, 2.19)       | 1.04 (0.86, 1.25) | 1.55 (1.11, 2.18)                      |
| >5y                              | 1.72 (1.37, 2.16)       | 1.04 (0.87, 1.24) | 1.51 (1.09, 2.10)                      |
| age 1st expos                    | 2.11 (1.59, 2.80)       | 1.09 (0.86, 1.37) | 2.11 (1.39, 3.22)                      |



|                       |                   |                   |                   |
|-----------------------|-------------------|-------------------|-------------------|
| primary infertility   |                   |                   |                   |
| >10y                  | 1.49 (0.95, 2.34) | 1.58 (1.10, 2.27) | 1.29 (0.67, 2.46) |
| >5y                   | 1.74 (1.15, 2.65) | 1.49 (1.06, 2.10) | 1.76 (0.95, 3.28) |
| age 1st expos         | 1.38 (0.82, 2.33) | 1.57 (1.03, 2.40) | 1.18 (0.55, 2.56) |
| pregnancy loss        |                   |                   |                   |
| >10y                  | 1.32 (1.10, 1.59) | 0.89 (0.76, 1.03) | 1.35 (1.03, 1.78) |
| >5y                   | 1.28 (1.07, 1.53) | 0.86 (0.75, 0.99) | 1.29 (1.00, 1.68) |
| age 1st expos         | 1.38 (1.11, 1.73) | 0.92 (0.77, 1.10) | 1.37 (0.98, 1.91) |
|                       |                   |                   |                   |
| countries             |                   |                   |                   |
| total infertility     |                   |                   |                   |
| Eth + Tanz            | 1.58 (1.28, 1.95) | 1.21 (1.02, 1.45) | 1.49 (1.09, 2.03) |
| Tanz + Uga            | 1.68 (1.28, 2.19) | 1.05 (0.81, 1.36) | 1.41 (0.94, 2.13) |
| Tanz alone            | 2.10 (1.02, 4.35) | 1.39 (0.66, 2.93) | 1.77 (0.77, 4.03) |
| secondary infertility |                   |                   |                   |
| Eth + Tanz            | 1.75 (1.38, 2.22) | 1.18 (0.97, 1.44) | 1.59 (1.13, 2.25) |
| Tanz + Uga            | 1.59 (1.19, 2.11) | 1.03 (0.78, 1.36) | 1.41 (0.91, 2.18) |
| Tanz alone            | 2.15 (0.96, 4.81) | 1.48 (0.65, 3.38) | 1.86 (0.74, 4.65) |
| primary infertility   |                   |                   |                   |
| Eth + Tanz            | 1.36 (0.91, 2.02) | 1.35 (0.97, 1.88) | 1.59 (0.88, 2.90) |
| Tanz + Uga            | 1.57 (0.94, 2.62) | 0.90 (0.53, 1.53) | 1.07 (0.50, 2.30) |
| Tanz alone            | 1.42 (0.36, 5.52) | 0.50 (0.12, 2.14) | 1.16 (0.24, 5.59) |
| pregnancy loss        |                   |                   |                   |
| Eth + Tanz            | 1.58 (1.31, 1.90) | 1.04 (0.89, 1.21) | 1.49 (1.13, 1.96) |
| Tanz + Uga            | 0.84 (0.68, 1.04) | 0.78 (0.64, 0.96) | 1.02 (0.74, 1.41) |
| Tanz alone            | 1.25 (0.72, 2.18) | 1.21 (0.68, 2.13) | 1.50 (0.80, 2.83) |

**Table 9. Association of impaired fertility with schistosomiasis endemicity of residence, Demographic and Health Surveys Ethiopia2011, Kenya2009, Tanzania2001, Uganda2011. Sensitivity analysis, various age restrictions and exposure categories.**

| 2009-2011                        | <i>S.h</i> high (ref=0) | <i>S.h</i> mod    | <i>S.h</i> high (ref= <i>S.m</i> high) |
|----------------------------------|-------------------------|-------------------|--|
| age restricted                   | OR (95%ci)              | OR (95%ci)        | OR (95%ci)                             |
| total infertility                |                         |                   |  |
| <40                              | 1.34 (1.11, 1.63)       | 1.35 (1.16, 1.56) | 1.62 (1.24, 2.12)                      |
| >20                              | 1.26 (1.07, 1.49)       | 1.31 (1.15, 1.49) | 1.47 (1.17, 1.86)                      |
| 20-45                            | 1.25 (1.05, 1.49)       | 1.24 (1.09, 1.42) | 1.52 (1.19, 1.95)                      |
| secondary infertility            |                         |                   |  |
| <40                              | 1.40 (1.12, 1.75)       | 1.43 (1.21, 1.70) | 1.70 (1.24, 2.32)                      |
| >20                              | 1.28 (1.07, 1.52)       | 1.35 (1.18, 1.54) | 1.53 (1.19, 1.96)                      |
| 20-45                            | 1.27 (1.04, 1.54)       | 1.28 (1.10, 1.49) | 1.54 (1.17, 2.03)                      |
| primary infertility              |                         |                   |  |
| <40                              | 1.50 (0.98, 2.30)       | 1.29 (0.92, 1.80) | 2.81 (1.48, 5.33)                      |
| >20                              | 1.62 (1.10, 2.38)       | 1.32 (0.97, 1.80) | 3.14 (1.76, 5.62)                      |
| 20-45                            | 1.62 (1.07, 2.45)       | 1.33 (0.95, 1.84) | 3.27 (1.74, 6.14)                      |
| pregnancy loss                   |                         |                   |  |
| <40                              | 1.11 (0.92, 1.33)       | 0.81 (0.71, 0.94) | 1.17 (0.90, 1.50)                      |
| >20                              | 1.14 (0.97, 1.33)       | 0.82 (0.72, 0.92) | 1.14 (0.91, 1.41)                      |
| 20-45                            | 1.10 (0.93, 1.30)       | 0.80 (0.70, 0.91) | 1.16 (0.91, 1.48)                      |
| total infecundity                |                         |                   |  |
| <40                              | 1.32 (1.08, 1.62)       | 1.35 (1.16, 1.57) | 1.55 (1.16, 2.05)                      |
| >20                              | 1.26 (1.07, 1.50)       | 1.35 (1.18, 1.54) | 1.39 (1.10, 1.77)                      |
| 20-45                            | 1.24 (1.03, 1.48)       | 1.27 (1.10, 1.46) | 1.43 (1.11, 1.86)                      |
| secondary infecundity            |                         |                   |  |
| <40                              | 1.38 (1.09, 1.74)       | 1.41 (1.18, 1.69) | 1.61 (1.16, 2.24)                      |
| >20                              | 1.29 (1.08, 1.54)       | 1.38 (1.20, 1.58) | 1.47 (1.14, 1.90)                      |
| 20-45                            | 1.26 (1.03, 1.54)       | 1.29 (1.11, 1.51) | 1.48 (1.11, 1.06)                      |
| primary infecundity              |                         |                   |  |
| <40                              | 1.32 (0.81, 2.16)       | 1.20 (0.82, 1.76) | 2.07 (1.00, 4.29)                      |
| >20                              | 1.42 (0.91, 2.21)       | 1.24 (0.88, 1.76) | 2.27 (1.18, 4.37)                      |
| 20-45                            | 1.42 (0.88, 2.29)       | 1.24 (0.85, 1.80) | 2.36 (1.16, 4.81)                      |
| exposure cut-points              |                         |                   |  |
| total infertility                |                         |                   |  |
| 30%, 5%                          | 1.20 (1.00, 1.45)       | 1.33 (1.18, 1.51) | 1.54 (1.18, 2.00)                      |
| 5% (dichot)                      | 1.29 (1.15, 1.46)       | NA                | 1.32 (1.12, 1.56)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.25 (1.06, 1.47)       | 1.31 (1.16, 1.49) | NA                                     |
| secondary infertility            |                         |                   |  |
| 30%, 5%                          | 1.17 (0.96, 1.42)       | 1.35 (1.19, 1.54) | 1.48 (1.12, 1.95)                      |
| 5% (dichot)                      | 1.30 (1.15, 1.48)       | NA                | 1.32 (1.12, 1.56)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.28 (1.07, 1.53)       | 1.35 (1.18, 1.55) | NA                                     |
| primary infertility              |                         |                   |  |
| 30%, 5%                          | 1.70 (1.17, 2.46)       | 1.31 (1.00, 1.72) | 2.91 (1.62, 5.24)                      |
| 5% (dichot)                      | 1.35 (1.04, 1.74)       | NA                | 1.25 (0.88, 1.79)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.47 (1.00, 2.16)       | 1.34 (0.99, 1.81) | NA                                     |
| pregnancy loss                   |                         |                   |  |
| 30%, 5%                          | 1.08 (0.91, 1.29)       | 0.84 (0.75, 0.95) | 1.05 (0.82, 1.35)                      |
| 5% (dichot)                      | 0.89 (0.80, 1.00)       | NA                | 1.00 (0.85, 1.17)                      |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.14 (0.98, 1.34)       | 0.81 (0.72, 0.92) | NA                                     |

|                                  |                   |                   |                   |
|----------------------------------|-------------------|-------------------|-------------------|
| total infecundity                |                   |                   |                   |
| 30%, 5%                          | 1.21 (1.00, 1.46) | 1.37 (1.20, 1.55) | 1.43 (1.09, 1.87) |
| 5% (dichot)                      | 1.32 (1.17, 1.49) | NA                | 1.30 (1.10, 1.54) |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.25 (1.05, 1.47) | 1.35 (1.19, 1.54) |                   |
| secondary infecundity            |                   |                   |                   |
| 30%, 5%                          | 1.18 (0.97, 1.44) | 1.39 (1.22, 1.59) | 1.39 (1.05, 1.84) |
| 5% (dichot)                      | 1.34 (1.18, 1.52) | NA                | 1.36 (1.14, 1.63) |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.29 (1.08, 1.54) | 1.38 (1.20, 1.59) | NA                |
| primary infecundity              |                   |                   |                   |
| 30%, 5%                          | 1.58 (1.04, 2.39) | 1.25 (0.93, 1.69) | 2.31 (1.21, 4.43) |
| 5% (dichot)                      | 1.28 (0.96, 1.70) | NA                | 1.07 (0.72, 1.59) |
| <i>S.h</i> alone (no <i>Sm</i> ) | 1.26 (0.81, 1.96) | 1.29 (0.91, 1.81) | NA                |
|                                  |                   |                   |                   |
| countries                        |                   |                   |                   |
| total infertility                |                   |                   |                   |
| Ken + Tanz + Uga                 | 1.49 (1.24, 1.79) | 1.50 (1.28, 1.76) | 1.58 (1.22, 2.05) |
| Eth + Tanz + Uga                 | 1.26 (1.06, 1.50) | 1.23 (1.07, 1.42) | 1.50 (1.17, 1.92) |
| Tanz alone                       | 1.27 (0.81, 1.99) | 1.18 (0.75, 1.86) | 1.64 (1.01, 2.65) |
| secondary infertility            |                   |                   |                   |
| Ken + Tanz + Uga                 | 1.41 (1.17, 1.71) | 1.50 (1.28, 1.77) | 1.44 (1.10, 1.89) |
| Eth + Tanz + Uga                 | 1.26 (1.05, 1.51) | 1.23 (1.06, 1.42) | 1.46 (1.12, 1.89) |
| Tanz alone                       | 1.24 (0.78, 1.96) | 1.19 (0.75, 1.89) |                   |
| primary infertility              |                   |                   |                   |
| Ken + Tanz + Uga                 | 2.41 (1.54, 3.76) | 1.69 (1.10, 2.57) | 3.36 (1.76, 6.42) |
| Eth + Tanz + Uga                 | 1.38 (0.98, 1.94) | 1.33 (1.01, 1.76) | 2.21 (1.32, 3.71) |
| Tanz alone                       | 1.94 (0.66, 5.75) | 1.62 (0.54, 4.83) | 3.23 (1.04, 9.99) |
| pregnancy loss                   |                   |                   |                   |
| Ken + Tanz + Uga                 | 0.91 (0.77, 1.06) | 0.73 (0.64, 0.84) | 0.87 (0.70, 1.09) |
| Eth + Tanz + Uga                 | 1.12 (0.95, 1.33) | 0.85 (0.74, 0.97) | 1.19 (0.94, 1.51) |
| Tanz alone                       | 0.89 (0.63, 1.27) | 0.86 (0.61, 1.22) | 0.90 (0.62, 1.31) |
| total infecundity                |                   |                   |                   |
| Ken + Tanz + Uga                 | 1.51 (1.26, 1.82) | 1.53 (1.30, 1.80) | 1.52 (1.16, 1.97) |
| Eth + Tanz + Uga                 | 1.28 (1.07, 1.52) | 1.25 (1.08, 1.45) | 1.41 (1.10, 1.82) |
| Tanz alone                       | 1.39 (0.89, 2.17) | 1.20 (0.77, 1.88) | 1.59 (0.99, 2.58) |
| secondary infecundity            |                   |                   |                   |
| Ken + Tanz + Uga                 | 1.45 (1.20, 1.75) | 1.53 (1.30, 1.81) | 1.42 (1.09, 1.87) |
| Eth + Tanz + Uga                 | 1.29 (1.07, 1.56) | 1.25 (1.07, 1.46) | 1.40 (1.07, 1.83) |
| Tanz alone                       | 1.37 (0.87, 2.17) | 1.23 (0.77, 1.95) | 1.54 (0.94, 2.53) |
| primary infecundity              |                   |                   |                   |
| Ken + Tanz + Uga                 | 2.26 (1.37, 3.74) | 1.51 (0.93, 2.45) | 2.51 (1.23, 5.19) |
| Eth + Tanz + Uga                 | 1.26 (0.84, 1.88) | 1.35 (0.98, 1.86) | 1.91 (1.05, 3.49) |
| Tanz alone                       | 1.83 (0.57, 5.93) | 1.21 (0.37, 3.98) | 2.18 (0.64, 7.43) |

## FIGURES

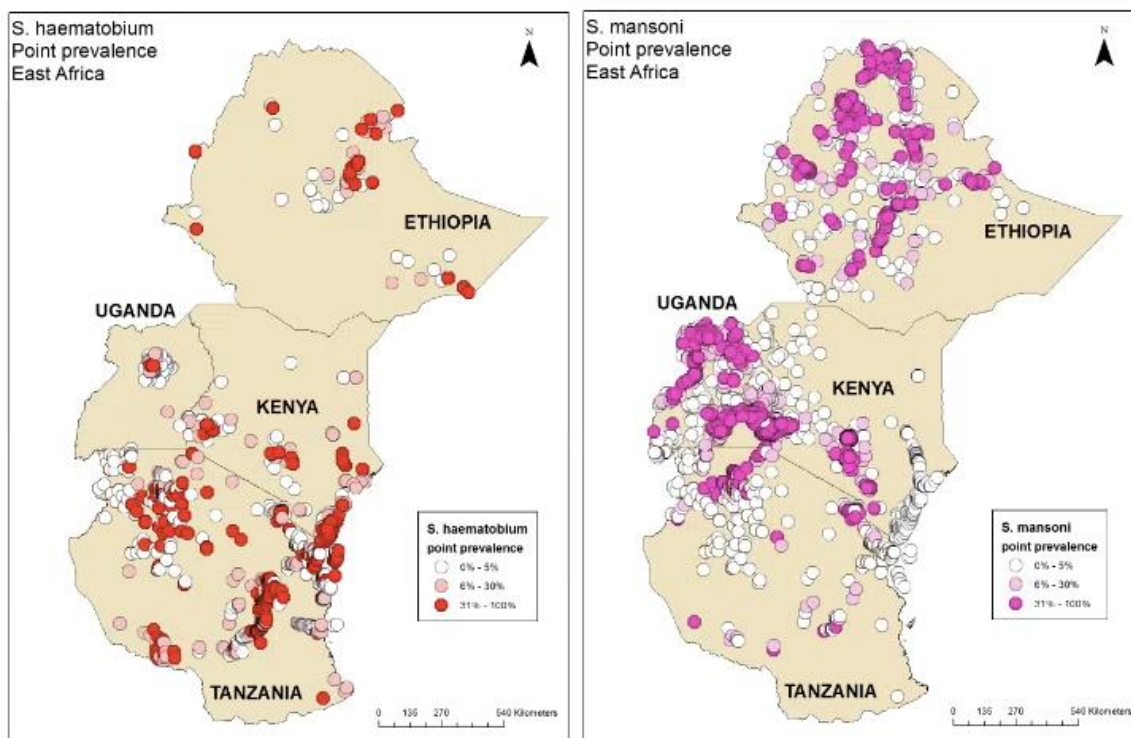


Figure 1: Map of schistosomiasis distribution in East Africa

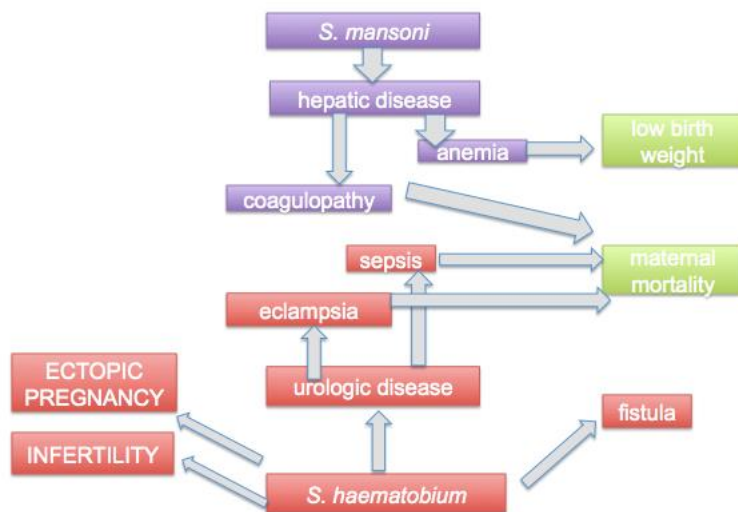


Figure 2: Potential effects of schistosomiasis in pregnancy.

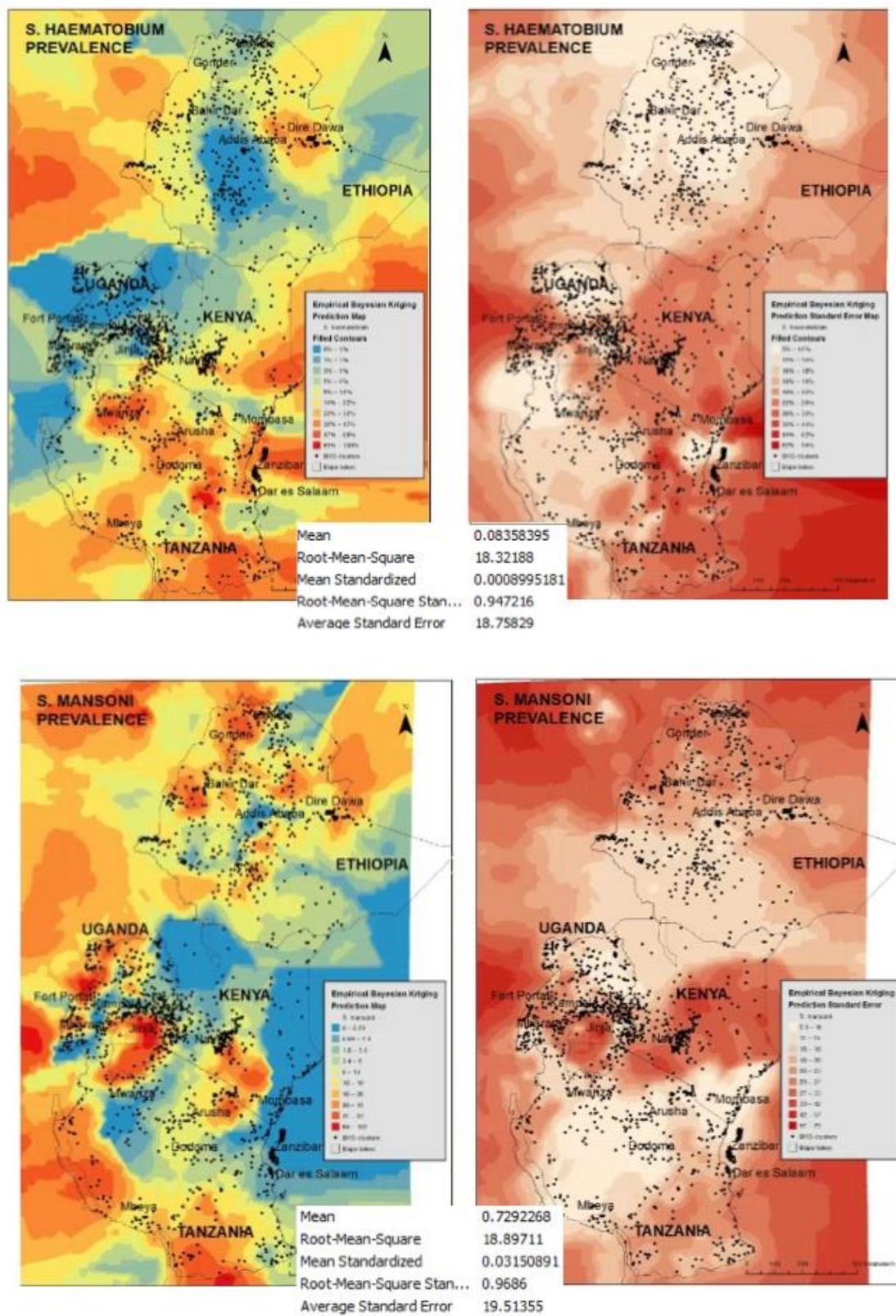
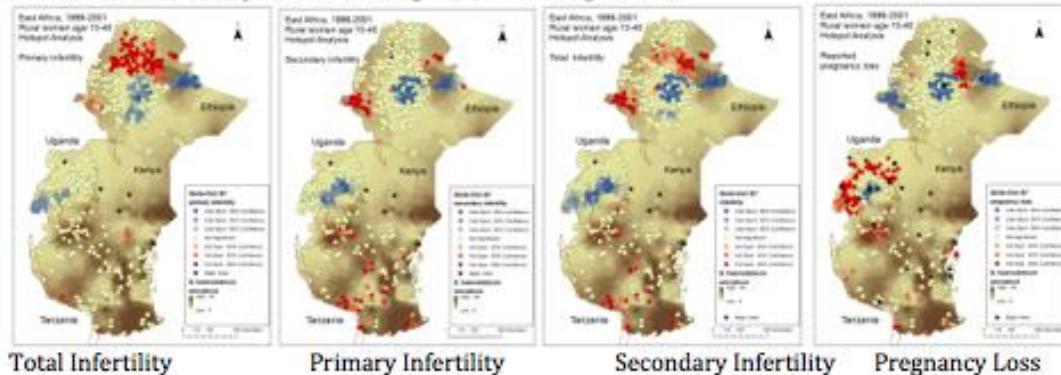


Figure 3. Predicted schistosomiasis distribution in East Africa, surface interpolated by Empirical Bayesian Kriging. Prediction maps (left), prediction standard error (right).

Getis-Ord  $G_i^*$  cluster analysis

1999-2001 DHS Ethiopia, Tanzania, Uganda, women age 15-50



2009-2011 DHS Ethiopia, Kenya, Tanzania, Uganda, women age 15-50

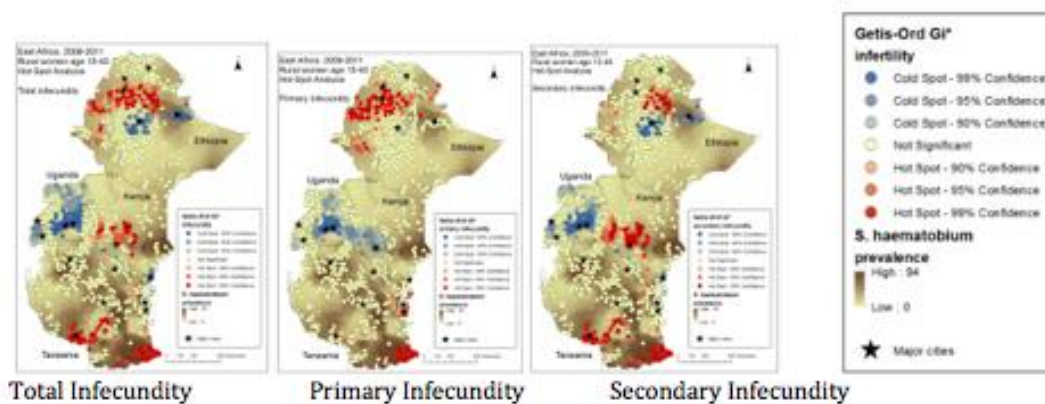
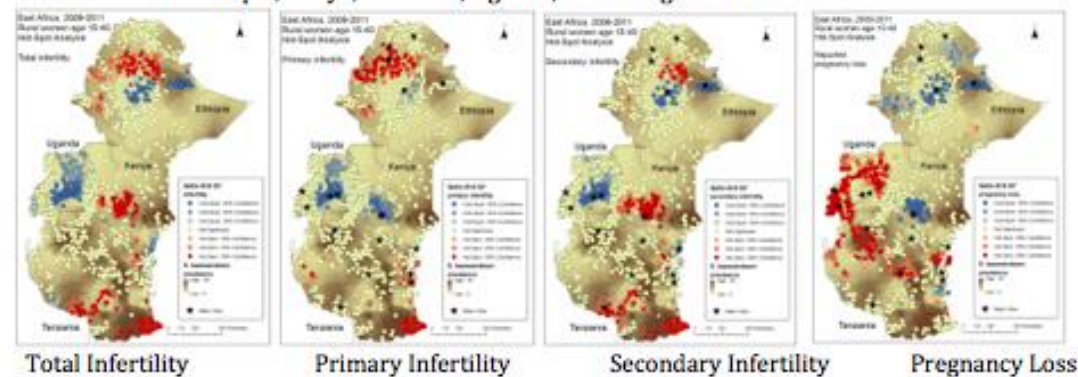
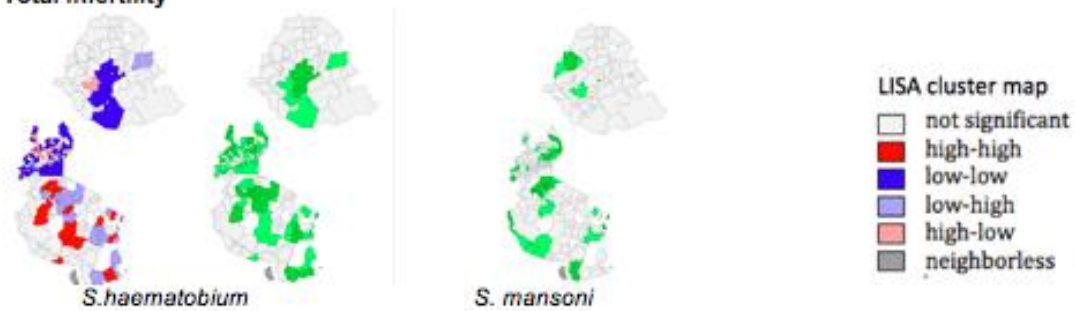


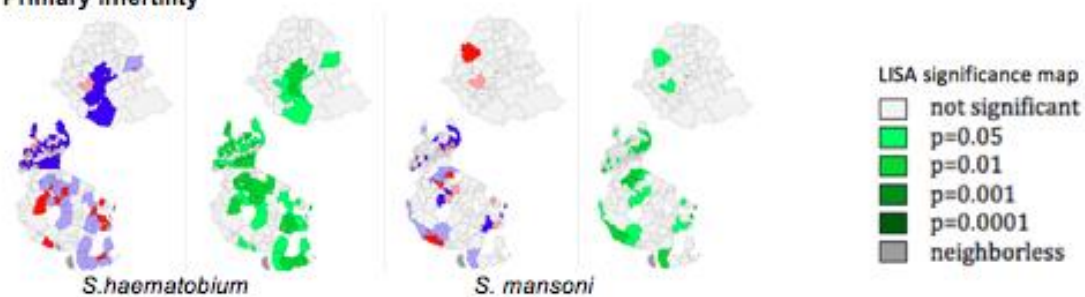
Figure 4. Local clustering of impaired fertility outcomes by Getis-Ord  $G_i^*$  hotspot analysis. Rural women under age 40, Demographic and Health Surveys Ethiopia, Kenya, Tanzania, Uganda

## 1999-2009 Demographic and Health Survey Ethiopia, Tanzania, Uganda, women age 15-50

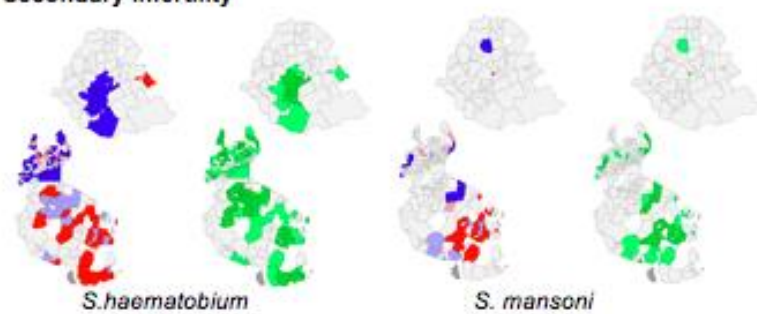
## Total infertility



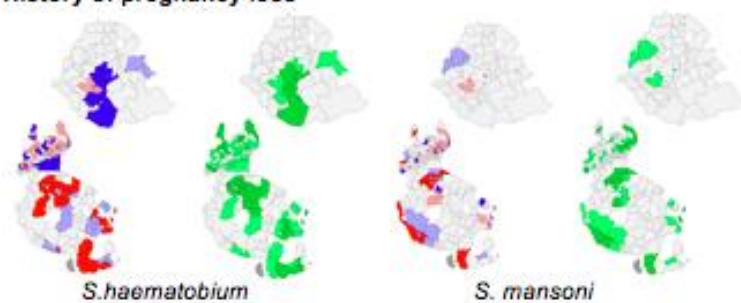
## Primary infertility



## Secondary infertility

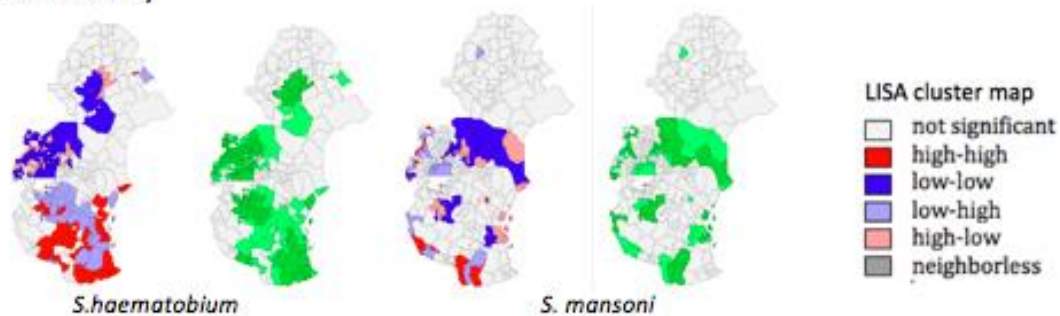


## History of pregnancy loss

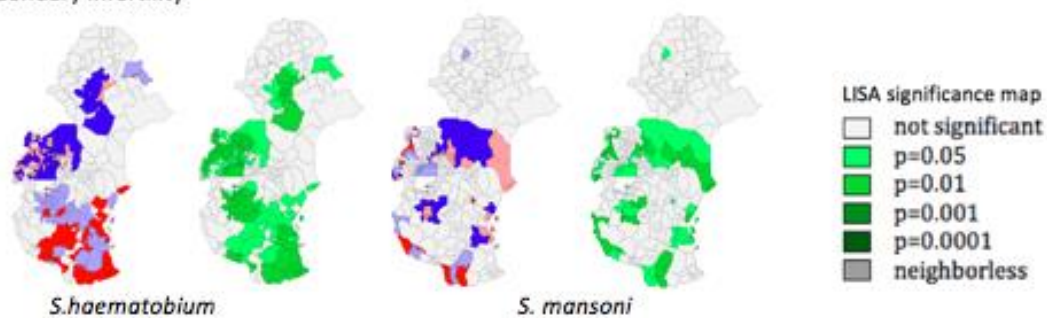


## 2009-2011 Demographic and Health Survey Ethiopia, Kenya, Tanzania, Uganda, women age 15-50

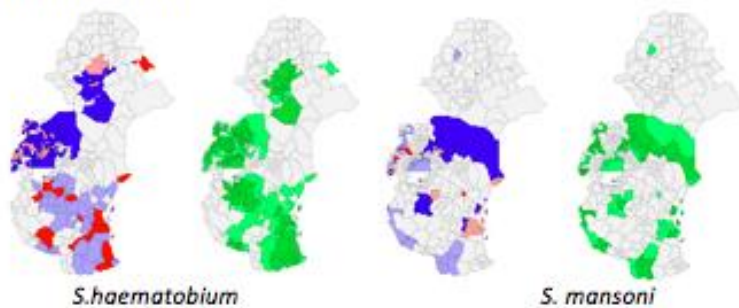
## Total infertility



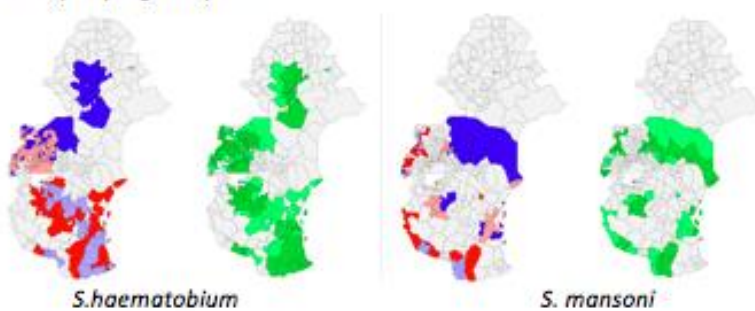
## Secondary infertility



## Primary infertility



## History of pregnancy loss





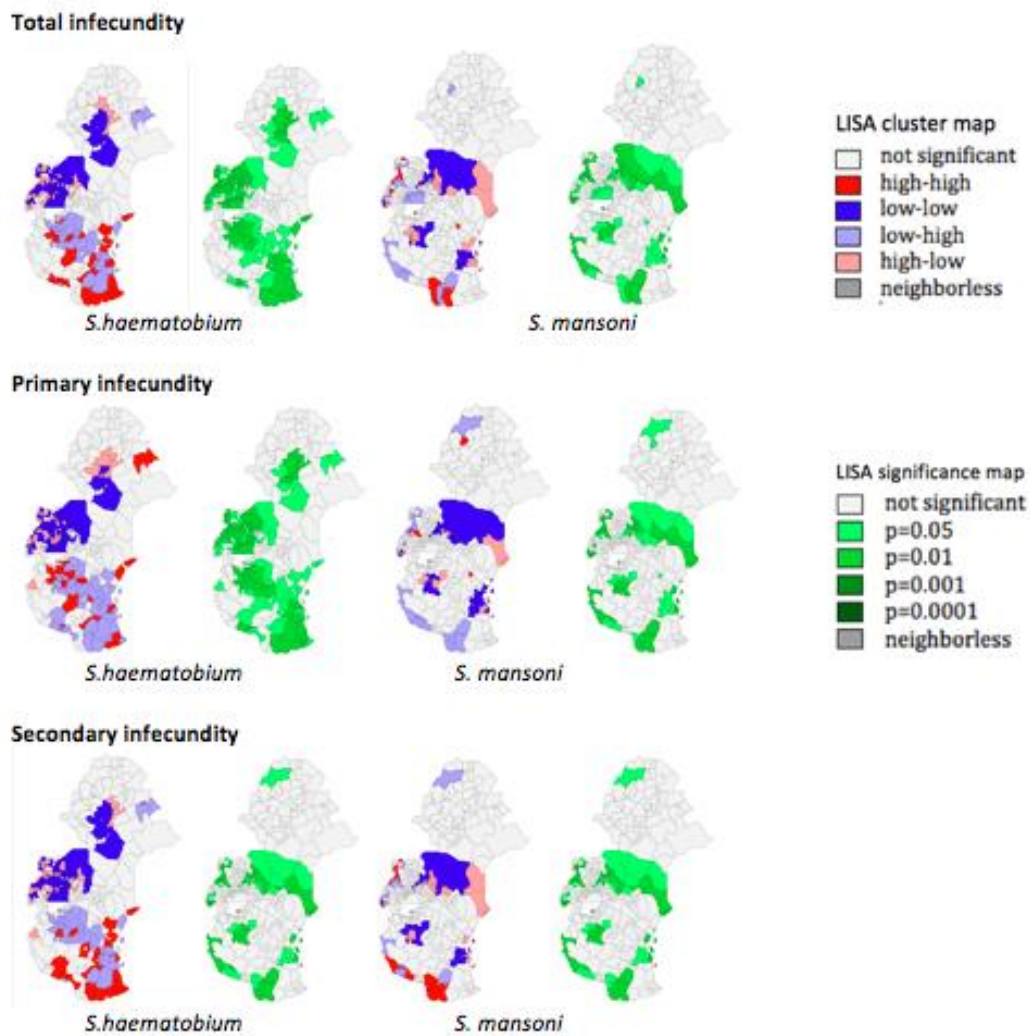


Figure 5. | Measures of fertility impairment: Bivariate Local Indices of Spatial Autocorrelation (LISA). Visual exploration of associations of high and low clustering of outcomes with areas of high and low prevalence of *Schistosoma haematobium* and *S. mansoni*

## REFERENCES

1. Colley DG, Bustinduy AL, Secor WE, King CH. Human schistosomiasis. *Lancet*. 2014 Jun 28;383(9936):2253-64.
2. Hotez PJ, Fenwick A. Schistosomiasis in Africa: an emerging tragedy in our new global health decade. *PLoS Neglected Tropical Diseases* [electronic resource]. 2009;3(9): e485.
3. King CH, Dangerfield-Cha M. The unacknowledged impact of chronic schistosomiasis. *Chronic Illness*. 2008 Mar;4(1):65-79.
4. van der Werf MJ, de Vlas SJ, Brooker S, Looman CWN, Nagelkerke NJD, Habbema JDF, et al. Quantification of clinical morbidity associated with schistosome infection in sub-Saharan Africa. *Acta Trop*. 2003 May;86(2-3):125-39.
5. Kittur N, Castleman JD, Campbell CHJ, King CH, Colley DG. Comparison of *Schistosoma mansoni* Prevalence and Intensity of Infection, as Determined by the Circulating Cathodic Antigen Urine Assay or by the Kato-Katz Fecal Assay: A Systematic Review. *American Journal of Tropical Medicine & Hygiene*. 2016 Mar 2;94(3):605-10.
6. Brooker S. Spatial epidemiology of human schistosomiasis in Africa: risk models, transmission dynamics and control. *Transactions of the Royal Society of Tropical Medicine & Hygiene*. 2007 Jan;101(1):1-8.
7. Abdel-Malek E. Factors conditioning the habitat of bilharziasis intermediate hosts of the family Planorbidae. *Bull World Health Organ*. 1958;18(5-6):785-818.
8. Moodley I, Kleinschmidt I, Sharp B, Craig M, Appleton C. Temperature-suitability maps for schistosomiasis in South Africa. *Annals of Tropical Medicine & Parasitology*. 2003 Sep;97(6):617-27.
9. Olds GR. Administration of praziquantel to pregnant and lactating women. *Acta Trop*. 2003 May;86(2-3):185-95.
10. Fenwick A, Webster JP. Schistosomiasis: challenges for control, treatment and drug resistance. *Curr Opin Infect Dis*. 2006 Dec;19(6):577-82.
11. Ross AGP, Olveda RM, Chy D, Olveda DU, Li Y, Harn DA, et al. Can mass drug administration lead to the sustainable control of schistosomiasis? *J Infect Dis*. 2015 Jan 15;211(2):283-9.
12. Hotez PJ. Mass drug administration and integrated control for the world's high-prevalence neglected tropical diseases. *Clinical Pharmacology & Therapeutics*. 2009 Jun;85(6):659-64.
13. Lelo AE, Mburu DN, Magoma GN, Mungai BN, Kihara JH, Mwangi IN, et al. No apparent reduction in schistosome burden or genetic diversity following four years of school-based mass drug administration in mwea, central kenya, a heavy transmission area. *PLoS Neglected Tropical Diseases* [electronic resource]. 2014 Oct;8(10): e3221.

14. Deribe K, Meribo K, Gebre T, Hailu A, Ali A, Aseffa A, et al. The burden of neglected tropical diseases in Ethiopia, and opportunities for integrated control and elimination. *Parasites & Vectors* [Electronic Resource]. 2012; 5: 240.
15. Mwandawiro CS, Nikolay B, Kihara JH, Ozier O, Mukoko DA, Mwanje MT, et al. Monitoring and evaluating the impact of national school-based deworming in Kenya: study design and baseline results. *Parasites & Vectors* [Electronic Resource]. 2013; 6: 198.
16. Mazigo HD, Nuwaha F, Kinung'hi SM, Morona D, Pinot de Moira A, Wilson S, et al. Epidemiology and control of human schistosomiasis in Tanzania. *Parasites & Vectors* [Electronic Resource]. 2012; 5: 274.
17. Parker M, Allen T. Does mass drug administration for the integrated treatment of neglected tropical diseases really work? Assessing evidence for the control of schistosomiasis and soil-transmitted helminths in Uganda. *Health Research Policy & Systems*. 2011; 9: 3.
18. Knopp S, Stothard JR, Rollinson D, Mohammed KA, Khamis IS, Marti H, et al. From morbidity control to transmission control: time to change tactics against helminths on Unguja Island, Zanzibar. *Acta Trop*. 2013 Nov;128(2):412-22.
19. Doumenge J, Mott K, Cheung C, Villenave D, Chapuis O. Atlas of the global distribution of schistosomiasis. Geneva: World Health Organization; 1987.
20. Brooker S, Hay SI, Issae W, Hall A, Kihamia CM, Lwambo NJ, et al. Predicting the distribution of urinary schistosomiasis in Tanzania using satellite sensor data. *Tropical Medicine & International Health*. 2001 Dec;6(12):998-1007.
21. Clements ACA, Lwambo NJS, Blair L, Nyandindi U, Kaatano G, Kinung'hi S, et al. Bayesian spatial analysis and disease mapping: tools to enhance planning and implementation of a schistosomiasis control programme in Tanzania. *Tropical Medicine & International Health*. 2006 Apr;11(4):490-503.
22. Simoonga C, Utzinger J, Brooker S, Vounatsou P, Appleton CC, Stensgaard AS, et al. Remote sensing, geographical information system and spatial analysis for schistosomiasis epidemiology and ecology in Africa. *Parasitology*. 2009 Nov;136(13):1683-93.
23. Schur N, Hurlimann E, Stensgaard AS, Chimfwembe K, Mushinge G, Simoonga C, et al. Spatially explicit *Schistosoma* infection risk in eastern Africa using Bayesian geostatistical modelling. *Acta Trop*. 2013 Nov;128(2):365-77.
24. Helling-Giese G, Sjaastad A, Poggensee G, Kjetland EF, Richter J, Chitsulo L, et al. Female genital schistosomiasis (FGS): relationship between gynecological and histopathological findings. *Acta Trop*. 1996 Dec 30;62(4):257-67.
25. Feldmeier H, Poggensee G, Krantz I, Helling-Giese G. Female genital schistosomiasis. New challenges from a gender perspective. *Tropical & Geographical Medicine*. 1995;47(2 Suppl): S2-15.

26. Poggensee G, Feldmeier H. Female genital schistosomiasis: facts and hypotheses. *Acta Trop*. 2001 Jun 22;79(3):193-210.
27. Feldmeier H, Daccal RC, Martins MJ, Soares V, Martins R. Genital manifestations of schistosomiasis mansoni in women: important but neglected. *Mem Inst Oswaldo Cruz*. 1998;93(Suppl 1):127-33.
28. Zaher MF, Badr MM, Fawzy RM. Bilharzial urinary fistula with report on 50 cases. *J Egypt Med Assoc*. 1959; 42: 412-9.
29. Kjetland EF, Ndhlovu PD, Gomo E, Mduluzi T, Midzi N, Gwanzura L, et al. Association between genital schistosomiasis and HIV in rural Zimbabwean women. *AIDS*. 2006 Feb 28;20(4):593-600.
30. Leutscher P, Ravaoalimalala VE, Raharisolo C, Ramarokoto CE, Rasendramino M, Raobelison A, et al. Clinical findings in female genital schistosomiasis in Madagascar. *Tropical Medicine & International Health*. 1998 Apr;3(4):327-32.
31. Downs JA, Mguta C, Kaatano GM, Mitchell KB, Bang H, Simplicie H, et al. Urogenital schistosomiasis in women of reproductive age in Tanzania's Lake Victoria region. *American Journal of Tropical Medicine & Hygiene*. 2011 Mar;84(3):364-9.
32. Poggensee G, Kiwelu I, Weger V, Goppner D, Diedrich T, Krantz I, et al. Female genital schistosomiasis of the lower genital tract: prevalence and disease-associated morbidity in northern Tanzania. *J Infect Dis*. 2000 Mar;181(3):1210-3.
33. Kjetland EF, Poggensee G, Helling-Giese G, Richter J, Sjaastad A, Chitsulo L, et al. Female genital schistosomiasis due to *Schistosoma haematobium*. Clinical and parasitological findings in women in rural Malawi. *Acta Trop*. 1996 Dec 30;62(4):239-55.
34. Talaat M, Watts S, Mekheimer S, Farook Ali H, Hamed H. The social context of reproductive health in an Egyptian hamlet: a pilot study to identify female genital schistosomiasis. *Soc Sci Med*. 2004 Feb;58(3):515-24.
35. Poggensee G, Feldmeier H, Krantz I. Schistosomiasis of the female genital tract: public health aspects. *Parasitology Today*. 1999 Sep;15(9):378-81.
36. Hegertun IE, Sulheim Gundersen KM, Kleppa E, Zulu SG, Gundersen SG, Taylor M, et al. *S. haematobium* as a common cause of genital morbidity in girls: a cross-sectional study of children in South Africa. *PLoS Neglected Tropical Diseases* [electronic resource]. 2013;7(3): e2104.
37. Leutscher P, Raharisolo C, Pecarrere JL, Ravaoalimalala VE, Serieye J, Rasendramino M, et al. *Schistosoma haematobium* induced lesions in the female genital tract in a village in Madagascar. *Acta Trop*. 1997 Jun 24;66(1):27-33.
38. Stecher CW, Kallestrup P, Kjetland EF, Vennervald B, Petersen E. Considering treatment of male genital schistosomiasis as a tool for future HIV prevention: a systematic review. *International Journal of Public Health*. 2015 Nov;60(7):839-48.

39. Salawu OT, Odaibo AB. Maternal schistosomiasis: a growing concern in sub-Saharan Africa. *Pathogens and Global Health*. 2014 Sep;108(6):263-70.
40. Siegrist D, Siegrist-Obimpeh P. *Schistosoma haematobium* infection in pregnancy. *Acta Trop*. 1992 Apr;50(4):317-21.
41. Ben-Chetrit E, Lachish T, Morch K, Atias D, Maguire C, Schwartz E. Schistosomiasis in pregnant travelers: a case series. *Journal of Travel Medicine*. 2015 Mar-Apr;22(2):94-8.
42. Qunhua L, Jiawen Z, Bozhao L, Zhilan P, Huijie Z, Shaoying W, et al. Investigation of association between female genital tract diseases and *Schistosomiasis japonica* infection. *Acta Trop*. 2000 Nov 2;77(2):179-83.
43. Siza JE. Risk factors associated with low birth weight of neonates among pregnant women attending a referral hospital in northern Tanzania. *Tanzan J Health Res*. 2008 Jan;10(1):1-8.
44. Friedman JF, Mital P, Kanzaria HK, Olds GR, Kurtis JD. Schistosomiasis and pregnancy. *Trends Parasitol*. 2007 Apr;23(4):159-64.
45. King CH, Dickman K, Tisch DJ. Reassessment of the cost of chronic helminthic infection: a meta-analysis of disability-related outcomes in endemic schistosomiasis. *Lancet*. 2005 Apr 30;365(9470):1561-9.
46. Ajanga A, Lwambo NJ, Blair L, Nyandindi U, Fenwick A, Brooker S. *Schistosoma mansoni* in pregnancy and associations with anaemia in northwest Tanzania. *Trans R Soc Trop Med Hyg*. 2006 Jan;100(1):59-63.
47. Muhangi L, Woodburn P, Omara M, Omoding N, Kizito D, Mpairwe H, et al. Associations between mild-to-moderate anaemia in pregnancy and helminth, malaria and HIV infection in Entebbe, Uganda. *Trans R Soc Trop Med Hyg*. 2007 Sep;101(9):899-907.
48. Geelhoed D, Agadzi F, Visser L, Ablordeppey E, Asare K, O'Rourke P, et al. Severe anemia in pregnancy in rural Ghana: a case-control study of causes and management. *Acta Obstet Gynecol Scand*. 2006;85(10):1165-71.
49. Kagu MB, Kawuwa MB, Gadzama GB. Anaemia in pregnancy: a cross-sectional study of pregnant women in a Sahelian tertiary hospital in Northeastern Nigeria. *J Obstet Gynaecol*. 2007 Oct;27(7):676-9.
50. Ndibazza J, Muhangi L, Akishule D, Kiggundu M, Ameke C, Oweka J, et al. Effects of deworming during pregnancy on maternal and perinatal outcomes in Entebbe, Uganda: a randomized controlled trial. *Clinical Infectious Diseases*. 2010 Feb 15;50(4):531-40.
51. Tanabe M. Haemostatic abnormalities in hepatosplenic schistosomiasis mansoni. *Parasitol Int*. 2003 Dec;52(4):351-9.
52. Kayange NM, Smart LR, Tallman JE, Chu EY, Fitzgerald DW, Pain KJ, et al. Kidney disease among children in sub-Saharan Africa: systematic review. *Pediatr Res*. 2015 Feb;77(2):272-81.

53. Nmorsi OPG, Kwandu UNCD, Ebiaguanye LM. Schistosoma haematobium and urinary tract pathogens co-infections in a rural community of Edo State, Nigeria. *J Commun Dis*. 2007 Jun;39(2):85-90.
54. Uneke CJ, Ugwuoru CD, Ngwu BA, Ogbu O, Agala CU. Public health implication of bacteriuria and antibiotic susceptibility of bacteria isolates in schistosoma haematobium-infected school pupils in Southeast Nigeria. *World Health & Population [Electronic Resource]*. 2006;8(3):66-76.
55. Ossai OP, Dankoli R, Nwodo C, Tukur D, Nsubuga P, Ogbuabor D, et al. Bacteriuria and urinary schistosomiasis in primary school children in rural communities in Enugu State, Nigeria, 2012. *The Pan African medical journal*. 2014;18(Suppl 1):15.
56. Kopelman JN, Miyazawa K. Hepatosplenic schistosomiasis in pregnancy: report of a case and review of the literature. *Am J Perinatol*. 1990 Oct;7(4):380-3.
57. Soto-Albors CE, Rayburn WF, Taylor L, Musselman M. Portal hypertension and hypersplenism in pregnancy secondary to chronic schistosomiasis. A case report. *J Reprod Med*. 1984 May;29(5):345-8.
58. Wirth HP, Casanova C, Meyenberger C, Hammer B, Ammann R, Blum HE. [Hepatosplenic schistosomiasis: case report and clinical review]. *Schweiz Med Wochenschr*. 1993 Oct 23;123(42):1991-5.
59. Obata NH, Kurauchi O, Kikkawa F, Yamada M, Fukuda Y, Itakura A. Preeclampsia with fetal death in a patient with schistosomiasis japonica. *Arch Gynecol Obstet*. 1998;261(2):101-4.
60. Goyaux N, Leke R, Keita N, Thonneau P. Ectopic pregnancy in African developing countries. *Acta Obstet Gynecol Scand*. 2003 Apr;82(4):305-12.
61. Olack B, Cosmas L, Mogeni D, Montgomery J. Causes of mortality in women of reproductive age living in an urban slum (Kibera) in Nairobi ; 2014.
62. Amoko DH, Buga GA. Clinical presentation of ectopic pregnancy in Transkei, South Africa. *East Afr Med J*. 1995 Dec;72(12):770-3.
63. Lindow SW, Moore PJ. Ectopic pregnancy: analysis of 100 cases. *International Journal of Gynaecology & Obstetrics*. 1988 Dec;27(3):371-5.
64. Ville Y, Leruez M, Glowaczower E, Fernandez H. [Fertility after extra-uterine pregnancy in Africa. Follow-up of a cohort of 145 patients over 5 years]. *J Gynecol Obstet Biol Reprod*. 1991;20(1):27-32.
65. Bugalho A, Strolego F, Pregazzi R, Osman N, Ching C. Extrauterine pregnancy in Mozambique. *Int J Gynaecol Obstet*. 1991 Mar;34(3):239-42.
66. Bland KG, Gelfand M. The influence of urinary bilharziasis on vesico-vaginal fistula in relation to causation and healing. *Transactions of the Royal Society of Tropical Medicine & Hygiene*. 1970;64(4):588-92.

67. Cui W. Mother or nothing: the agony of infertility. *Bull World Health Organ.* 2010 Dec 1;88(12):881-2.
68. Gurunath S, Pandian Z, Anderson RA, Bhattacharya S. Defining infertility - a systematic review of prevalence studies. *Hum Reprod Update.* 2011 Sep-Oct;17(5):575-88.
69. Rutstein, Shea O. and Iqbal H. Shah. Infecundity, Infertility, and Childlessness in Developing Countries. DHS Comparative Reports. Calverton, Maryland, USA: ORC Macro and the World Health Organization; 2004. Report No.: 9.
70. Menken J, Larsen U. Estimating the incidence and prevalence and analyzing the correlates of infertility and sterility. *Ann N Y Acad Sci.* 1994 Feb 18; 709:249-65.
71. World Health Organization. Reproductive Health Indicators for Global Monitoring: Report of the Second Interagency Meeting. Geneva: WHO; 2000. Report No.: WHO/RHR/01.19.
72. Bongaarts J, Frank O, Lesthaeghe R. The Proximate Determinants of Fertility in Sub-Saharan Africa John Bongaarts, Odile Frank and Ron Lesthaeghe, Vol. 10, No. 3 (Sep., 1984), pp. 511-537. *Population and Development Review.* 1984;10(3):511.
73. Ericksen K, Brunette T. Patterns and predictors of infertility among African women: a cross-national survey of twenty-seven nations. *Soc Sci Med.* 1996 Jan;42(2):209-20.
74. Larsen U. Research on infertility: which definition should we use? *Fertility & Sterility.* 2005 Apr;83(4):846-52.
75. Mascarenhas MN, Cheung H, Mathers CD, Stevens GA. Measuring infertility in populations: constructing a standard definition for use with demographic and reproductive health surveys. *Population Health Metrics.* 2012;10(1):17.
76. Larsen U, Menken J. Individual-level sterility: a new method of estimation with application to sub-Saharan Africa. *Demography.* 1991 May;28(2):229-47.
77. Larsen U, Menken J. Measuring sterility from incomplete birth histories. *Demography.* 1989 May;26(2):185-201.
78. Mascarenhas MN, Flaxman SR, Boerma T, Vanderpoel S, Stevens GA. National, regional, and global trends in infertility prevalence since 1990: a systematic analysis of 277 health surveys. *PLoS Medicine / Public Library of Science.* 2012;9(12): e1001356.
79. Cates W, Farley TM, Rowe PJ. Worldwide patterns of infertility: is Africa different? *Lancet.* 1985 Sep 14;2(8455):596-8.
80. Kitilla T. Hysterosalpingography in the evaluation of infertility: a five years review. (FGAE, 2001 -5). *Ethiop Med J.* 2010 Oct;48(4):267-75.
81. Stewart-Smythe GW, van Iddekinge B. Lessons learned from infertility investigations in the public sector. *South African Medical Journal.Suid-Afrikaanse Tydskrif Vir Geneeskunde.* 2003;93(2):141-3.

82. Umeora OU, Mbazor JO, Okpere EE. Tubal factor infertility in Benin City, Nigeria - sociodemographics of patients and aetiopathogenic factors. *Trop Doct.* 2007 Apr;37(2):92-4.
83. Marais NF, Wessels PH, Smith MS, Gericke A. [The prevalence of Chlamydia trachomatis infections in new patients at the Infertility Clinic, UOFS, Bloemfontein]. *South African Medical Journal.Suid-Afrikaanse Tydskrif Vir Geneeskunde.* 1990;77(5):232-3.
84. Siemer J, Theile O, Larbi Y, Fasching PA, Danso KA, Kreienberg R, et al. Chlamydia trachomatis infection as a risk factor for infertility among women in Ghana, West Africa. *American Journal of Tropical Medicine & Hygiene.* 2008 Feb;78(2):323-7.
85. Margolis K, Wranz PA, Kruger TF, Joubert JJ, Odendaal HJ. Genital tuberculosis at Tygerberg Hospital--prevalence, clinical presentation and diagnosis. *South African Medical Journal.Suid-Afrikaanse Tydskrif Vir Geneeskunde.* 1992;81(1):12-5.
86. Oosthuizen AP, Wessels PH, Hefer JN. Tuberculosis of the female genital tract in patients attending an infertility clinic. *South African Medical Journal.Suid-Afrikaanse Tydskrif Vir Geneeskunde.* 1990;77(11):562-4.
87. Kjetland EF, Kurewa EN, Mduluzi T, Midzi N, Gomo E, Friis H, et al. The first community-based report on the effect of genital Schistosoma haematobium infection on female fertility. *Fertility & Sterility.* 2010 Sep;94(4):1551-3.
88. Miller-Fellows S, Hildebrand V, Furin J, King C. Schistosomiasis haematobia and infertility in Coast Province, Kenya American Society of Tropical Medicine and Hygiene Annual Meeting. 2014; Abstract #55.
89. Frank O. Infertility in sub-Saharan Africa: Estimates and Implications. *Population and Development Review.* 1983;9(1):137.
90. Romaniuk A. Infertility in Tropical Africa. In: Caldwell JC PC, editor. *The Population of Tropical Africa.* 1968. p. 214.
91. Allard R. [Gynecological contribution to the study of sterility among the Mongo people of Befale]. *Ann Soc Belg Med Trop.* 1955 Dec 31;35(6):631-48.
92. Velghe A. [Study of the demographic situation of populations in Kibombo and Kasongo territories: medical causes of the decline in birth rate]. Belgium: FBEI; 1952.
93. Sunil TS, Pillai VK. Sterility in zambia. *Ann Hum Biol.* 2002 Jul-Aug;29(4):414-21.
94. Larsen U. Primary and secondary infertility in sub-Saharan Africa. *Int J Epidemiol.* 2000 Apr;29(2):285-91.
95. Tesfaghiorghis H. Infecundity and subfertility among the rural population of Ethiopia. *J Biosoc Sci.* 1991 Oct;23(4):461-75.
96. Larsen U, Raggars H. Levels and trends in infertility in sub-Saharan Africa. In: Boerma J, Mgalla Z, editors. ; 2001. p. 25-69.



97. Blacker J, Opiyo C, Jasseh M, Sloggett A, Ssekamatte-Ssebuliba J. Fertility in Kenya and Uganda: a comparative study of trends and determinants. *Population Studies*. 2005 Nov;59(3):355-73.
98. Hurlimann E, Schur N, Boutsika K, Stensgaard A, Laserna de Himpsl M, Ziegelbauer K, et al. Toward an open-access global database for mapping, control, and surveillance of neglected tropical diseases. *PLoS Neglected Tropical Diseases* [electronic resource]. 2011 Dec;5(12): e1404.
99. Brooker S, Hotez PJ, Bundy DAP. The global atlas of helminth infection: mapping the way forward in neglected tropical disease control. *PLoS Neglected Tropical Diseases* [electronic resource]. 2010;4(7): e779.
100. Flueckiger RM, Nikolay B, Gelderblom HC, Smith JL, Haddad D, Tack W, et al. Integrating data and resources on neglected tropical diseases for better planning: the NTD mapping tool (NTDmap.org). *PLoS Neglected Tropical Diseases* [electronic resource]. 2015 Feb;9(2): e0003400.
101. Perez-Heydrich C, Warren J, Burgert C, Emch M. Guidelines On the Use Of DHS GPS Data. Spatial Analysis Reports No. 8. Calverton, Maryland: ICF International; 2013.
102. Measure DHS. Survey Organization Manual for Demographic and Health Surveys. Calverton, MD: ICF International; 2012.
103. Measure DHS. Demographic and Health Survey Sampling and Household Listing Manual. Calverton, MD: ICF International; 2012.
104. Haws RA, Mashasi I, Mrisho M, Schellenberg JA, Darmstadt GL, Winch PJ. "These are not good things for other people to know": how rural Tanzanian women's experiences of pregnancy loss and early neonatal death may impact survey data quality. *Soc Sci Med*. 2010 Nov;71(10):1764-72.
105. Burgert CR, Bradley SEK, Arnold F, Eckert E. Improving estimates of insecticide-treated mosquito net coverage from household surveys: using geographic coordinates to account for endemicity. *Malaria Journal*. 2014; 13:254.
106. Curtis S, Hossain M. West Africa spatial analysis prototype exploratory analysis: The effect of aridity on child nutritional status. DHS Spatial Analysis Reports No. 2. Calverton, MD: Macro International; 1998.
107. Anselin L, Syabri I, Kho Y. GeoDa: An Introduction to Spatial Data Analysis. *Geographical Analysis*. 2006;38(1):5.
108. Burgert C. Spatial Interpolation with Demographic and Health Survey Data: Key Considerations. DHS Spatial Analysis Reports No. 9. Rockville, Maryland: ICF International; 2014.

109. Gething P, Tatem A, Bird T, Burgert-Brucker C. Creating Spatial Interpolation Surfaces with DHS Data DHS Spatial Analysis Reports No. 11. Rockville, Maryland: ICF International; 2015.
110. Ren R. Note on DHS Standard Weight De-normalization ICF International.
111. Bell B, Ene M, Smiley W, Schoeneberger J. A Multilevel Model Primer Using SAS Proc Mixed. SAS Global Forum 2013. Columbia, SC: University of South Carolina; 2013.
112. Ene M, Leighton E, Blue G, Bell B. Multilevel Models for Categorical Data Using SAS Proc GLIMMIX: the Basics. Paper 3430-2015. Columbia, SC: University of South Carolina; 2015.
113. Adeniji KA. Morphological features and organ distribution of schistosomal infection. African Journal of Medicine & Medical Sciences. 2001 Mar-Jun;30(1-2):53-6.
114. Bailey SL, Price J, Llewelyn M. Fluke infertility: the late cost of a quick swim. Journal of Travel Medicine. 2011 Jan-Feb;18(1):61-2.
115. Balasch J, Martinez-Roman S, Creus M, Campo E, Fortuny A, Vanrell JA. Schistosomiasis: an unusual cause of tubal infertility. Human Reproduction. 1995 Jul;10(7):1725-7.
116. Billy-Brissac R, Foucan L, Gallais A, Wan-Ajouhu G, Roudier M. [Genital Schistosoma mansoni bilharziasis in women: apropos of 2 cases in Guadeloupe]. Med Trop. 1994;54(4):345-8.
117. Bland KG, Gelfand M. The effects of schistosomiasis on the fallopian tubes in the African female. Journal of Obstetrics & Gynaecology of the British Commonwealth. 1970 Nov;77(11):1024-7.
118. Boule M, Notelovitz M. Bilharzia of the female genital tract. South African Journal of Obstetrics and Gynaecology. 1964(6 June):48.
119. Bullough CH. Infertility and bilharziasis of the female genital tract. British Journal of Obstetrics & Gynaecology. 1976 Oct;83(10):819-22.
120. Cornier E, Jouhanet H, Feintuch H, Delafontaine D, Dao C, Bouccara L. [Intra-uterine pregnancy after microsurgical salpingostomy for bilharziasis of the tubal ampulla]. Nouvelle Presse Medicale. 1981 Jul 11-25;10(30):2514-5.
121. Crump JA, Murdoch DR, Chambers ST, Aickin DR, Hunter LA. Female genital schistosomiasis. Journal of Travel Medicine. 2000 Jan;7(1):30-2.
122. DeMille PA, Bourquin P, Sun CC, Kauffman L. Cytologic diagnosis of Schistosoma haematobium in routine cervicovaginal smear from an infertile woman. Diagn Cytopathol. 1995 Aug;13(2):181-2.
123. Diouf B, Spay G, Toure P. [Genital bilharziasis in women]. Bulletin de la Societe Medicale d'Afrique Noire de Langue Francaise. 1973;18(4):517-9.

124. Ekoukou D, Luzolo-Lukanu A, Mulard C, Bazin C, Ng Wing Tin L. [Peritoneal and tubal *Schistosoma haematobium* bilharziasis. Two case reports]. *J Gynecol Obstet Biol Reprod*. 1995;24(8):819-24.
125. El-Mahgoub S. Pelvic schistosomiasis and infertility. *International Journal of Gynaecology & Obstetrics*. 1982 Jun;20(3):201-6.
126. El-maraghy MA, Elyan A, El-Leithy AG, El-Tehewey FA, Senna IA, El-Tawil A, et al. Bilharziasis of the female genital tract; new concepts. *J Egypt Soc Parasitol*. 1982 Jun;12(1):179-86.
127. Gilbert B. Schistosomiasis of the female genital tract and neighboring tissues. *Journal of Obstetrics & Gynaecology of the British Empire*. 1943;50(5):317.
128. Harouny A, Pedersen H. Pelveo-peritoneal schistosomiasis as a cause of primary infertility. *International Journal of Gynaecology & Obstetrics*. 1988 Dec;27(3):467-9.
129. Hoffmann H, Bauerfeind I. High tissue egg burden mechanically impairing the tubal motility in genital schistosomiasis of the female. *Acta Obstet Gynecol Scand*. 2003 Oct;82(10):970-1.
130. Krolkowski A, Janowski K, Larsen JV. Asherman syndrome caused by schistosomiasis. *Obstetrics & Gynecology*. 1995 May;85(5 Pt 2):898-9.
131. Le Guyader A, Kekeh K, Richier ME, Ferrand B, Chevrel ML. [Genital bilharziasis in women. A case of endometritis due to *Schistosoma haematobium*]. *Archives d Anatomie Pathologique*. 1965 Dec;13(4):256-8.
132. Morice P, Chapron C, Vacher Lavenu MC, Terrasse G, Dubuisson JB. [Genital bilharziasis and female infertility. Review of the literature and three case reports]. *Fertilite Contraception Sexualite*. 1996 Jan;24(1):56-61.
133. Mouktar M. Functional disorders due to bilharzial infection of the female genital tract. *Journal of Obstetrics & Gynaecology of the British Commonwealth*. 1966; 73:307.
134. Nouhou H, Seve B, Idi N, Moussa F. [Schistosomiasis of the female genital tract: anatomoclinical and histopathological aspects. Apropos of 26 cases]. *Bulletin de la Societe de Pathologie Exotique*. 1998;91(3):221-3.
135. O'Leary M. Case records of the Massachusetts General Hospital. Weekly clinicopathological exercises. Case 1-1994. A 27-year-old woman with secondary infertility and a bladder mass. *N Engl J Med*. 1994 Jan 6;330(1):51-7.
136. Ogunniyi SO, Nganwuchu AM, Adenle MA, Dare FO. Pregnancy following infertility due to pelvic schistosomiasis-a case report. *West Afr J Med*. 1994 Apr-Jun;13(2):132-3.
137. Picaud A, Bennani S, Mba Allo L, Mouely G, Nlome-Nze AR, Ogowet-Igumu N. [Unusual causes of hemoperitoneum of genital origin]. *J Gynecol Obstet Biol Reprod*. 1990;19(4):441-5.

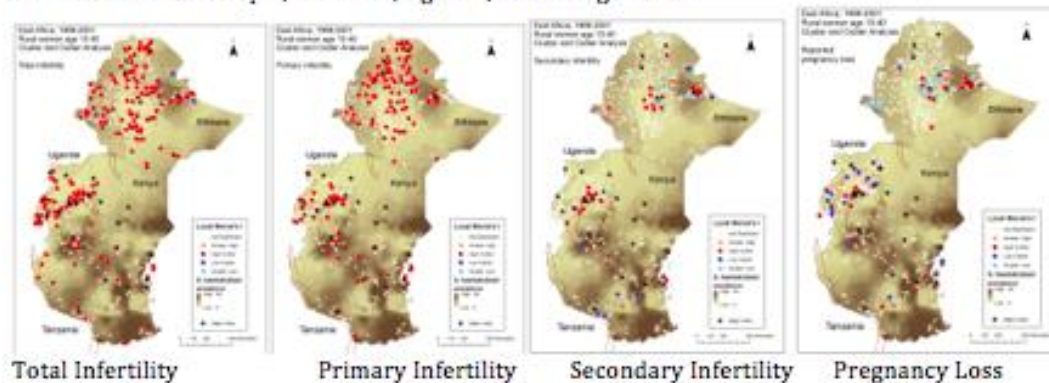
138. Schanz A, Richter J, Beyer I, Baldus SE, Hess AP, Kruessel JS. Genital schistosomiasis as a cause of female sterility and acute abdomen. *Fertility & Sterility*. 2010 Apr;93(6): 2075.e7, 2075.e9.
139. Schroers B, Peschen-van Issum M, Reinert RR, Fuzesi L, Biesterfeld, Winkler M. [Sterility as a sequela of tubal schistosomiasis]. *Geburtshilfe Frauenheilkd*. 1995 Mar;55(3):140-2.
140. Sheorey H, Charles PG, Pyman J. Ectopic schistosomiasis in a returned traveler. *Journal of Travel Medicine*. 2004 Jul-Aug;11(4):251-2.
141. Swai B, Poggensee G, Mtweve S, Krantz I. Female genital schistosomiasis as an evidence of a neglected cause for reproductive ill-health: a retrospective histopathological study from Tanzania. *BMC Infectious Diseases*. 2006; 6:134.
142. Vass AC, Lucey JJ. Bilharzial granuloma of the fallopian tube. Case report. *British Journal of Obstetrics & Gynaecology*. 1982 Oct;89(10):867-9.
143. Aminu MB, Abdullahi K, Dattijo LM. Tubal ectopic gestation associated with genital schistosomiasis: a case report. *Afr J Reprod Health*. 2014 Jun;18(2):144-6.
144. Bahrami S, Alatassi H, Slone SP, O'Connor DM. Tubal gestation and schistosomiasis: a case report. *J Reprod Med*. 2006 Jul;51(7):595-8.
145. Bugalho A, Strolego F, Benussi G, Pregazzi R, Osman N. [Schistosomiasis: possible cause of ectopic pregnancy. Four clinical cases]. *Minerva Ginecol*. 1991 Dec;43(12):577-9.
146. El-Bedri L. Ectopic pregnancy caused by *Schistosoma haematobium* infection of the fallopian tube; a case report. *American Journal of Obstetrics & Gynecology*. 1958 Sep;76(3):515-7.
147. Eogan M, O'Malley A, Flavin R, Gillan J, McKenna P, Coulter-Smith S. Ectopic pregnancy associated with tubal schistosomiasis. *Ir Med J*. 2002 Sep;95(8):250.
148. Garba M, Almoustapha T, Garba A, Nouhou H. [Extra uterine pregnancy associated with a tubal schistosomiasis due to *Schistosoma haematobium*. A case report from Niger]. *Bulletin de la Societe de Pathologie Exotique*. 2004 Feb;97(1):41-2.
149. Hassim AM. Tubal gestation associated with schistosomiasis. *Journal of Obstetrics & Gynaecology of the British Commonwealth*. 1966 Oct;73(5):855-6.
150. Laxman VV, Adamson B, Mahmood T. Recurrent ectopic pregnancy due to *Schistosoma haematobium*. *Journal of Obstetrics & Gynaecology*. 2008 May;28(4):461-2.
151. Mayat MG. Ectopic pregnancy in a tube infested with *Bilharzia*. *South African Medical Journal/Suid-Afrikaanse Tydskrif Vir Geneeskunde*. 1959;33(11):219-20.

152. Mohammed AZ, Uzoho CC, Galadanci HS, Ashimi A. Ruptured tubal gestation: an unusual presentation of *Schistosoma haematobium* infection of the fallopian tube. *Trop Doct.* 2004 Jan;34(1):48-9.
153. Okonofua FE, Ojo OS, Odunsi OA, Odesanmi WO. Ectopic pregnancy associated with tubal schistosomiasis in a Nigerian woman. *International Journal of Gynaecology & Obstetrics.* 1990 Jul;32(3):281-4.
154. Owusu-Bempah A, Odoi AT, Dassah ET. Genital schistosomiasis leading to ectopic pregnancy and subfertility: a case for parasitic evaluation of gynaecologic patients in schistosomiasis endemic areas. *Case Reports in Obstetrics and Gynecology.* 2013: 634264.
155. Sahu L, Tempe A, Singh S, Khurana N. Ruptured ectopic pregnancy associated with tubal schistosomiasis. *J Postgrad Med.* 2013 Oct-Dec;59(4):315-7.
156. Scheller R. [Gynecologic bilharziasis and ectopic pregnancy]. *Zentralbl Gynakol.* 1974 Jan 18;96(3):88-92.
157. Schneider D, Steyn DW. Genital schistosomiasis presenting as suspected ectopic pregnancy in the Western Cape. *South African Medical Journal.Suid-Afrikaanse Tydskrif Vir Geneeskunde.* 2000;90(6):609.
158. Ville Y, Leruez M, Picaud A, Walter P, Fernandez H. Tubal schistosomiasis as a cause of ectopic pregnancy in endemic areas; a report of three cases. *European Journal of Obstetrics, Gynecology, & Reproductive Biology.* 1991 Nov 3;42(1):77-9.

## APPENDIX: ADDITIONAL MAPS

## Local Moran's I

1999-2001 DHS Ethiopia, Tanzania, Uganda, women age 15-50



2009-2011 DHS Ethiopia, Kenya, Tanzania, Uganda, women age 15-50

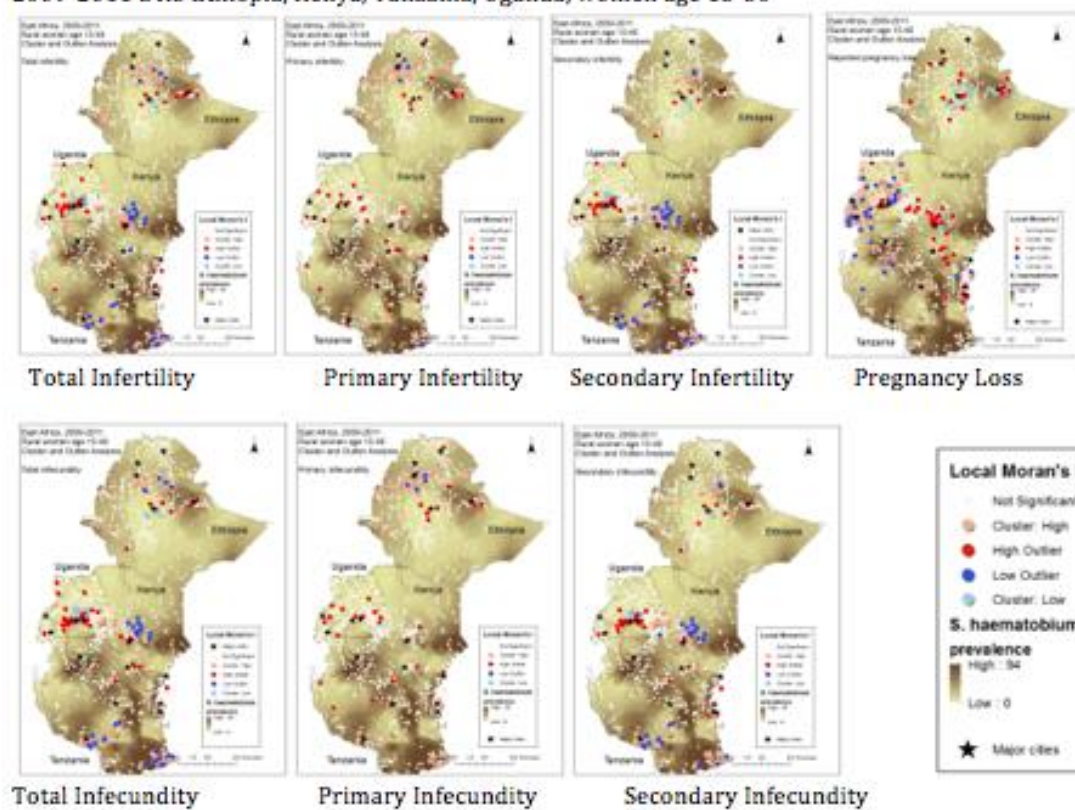
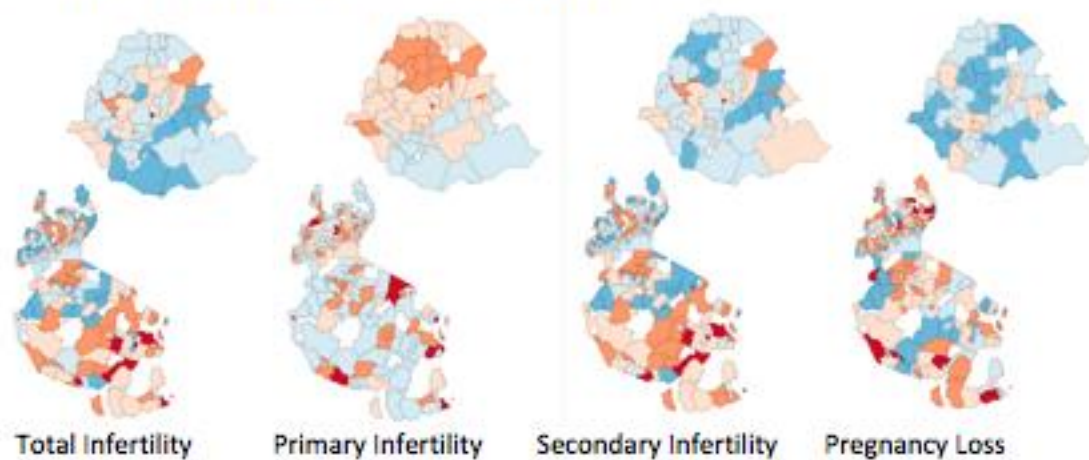


Figure 6. Local clustering of impaired fertility outcomes by Local Moran's I Cluster and Outlier Analysis. Rural women under age 40, Demographic and Health Surveys Ethiopia, Kenya, Tanzania, Uganda

1999-2001 DHS Ethiopia, Tanzania, Uganda, women age 15-50



2009-2011 DHS Ethiopia, Kenya, Tanzania, Uganda, women age 15-50

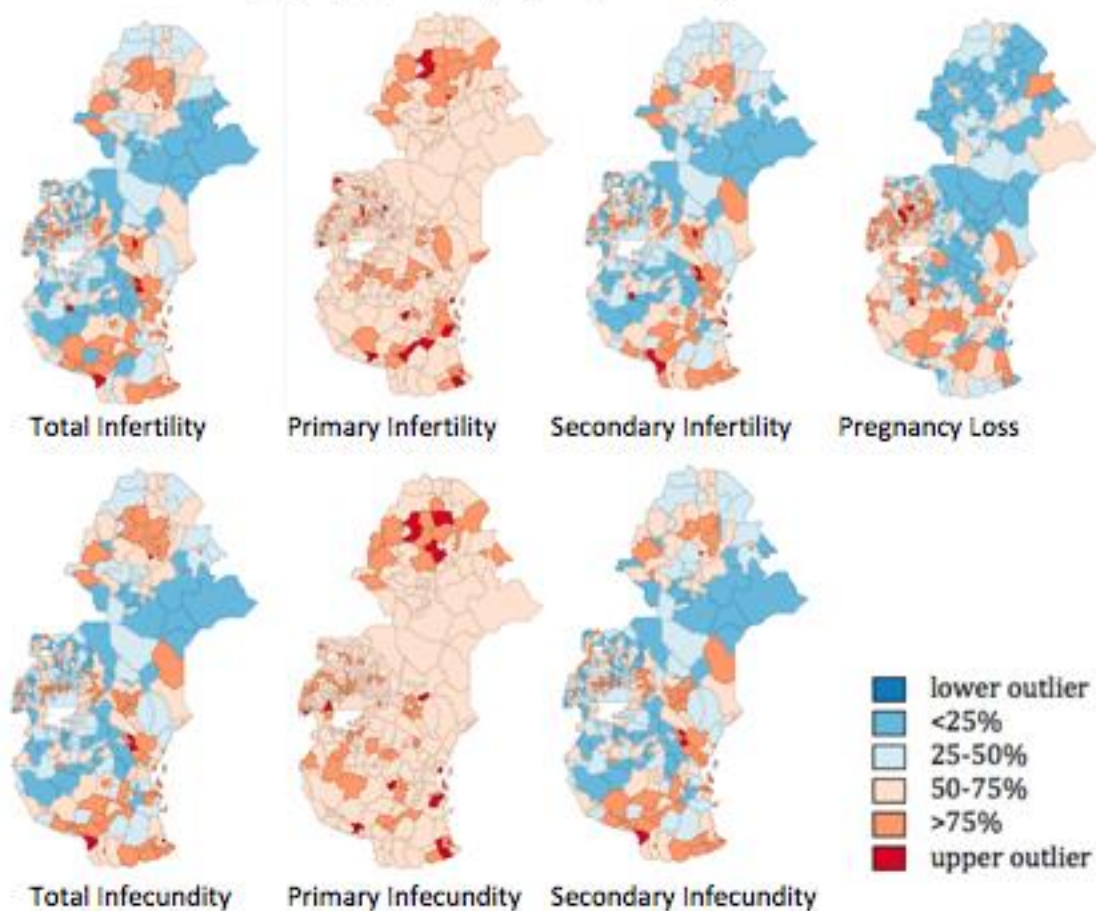
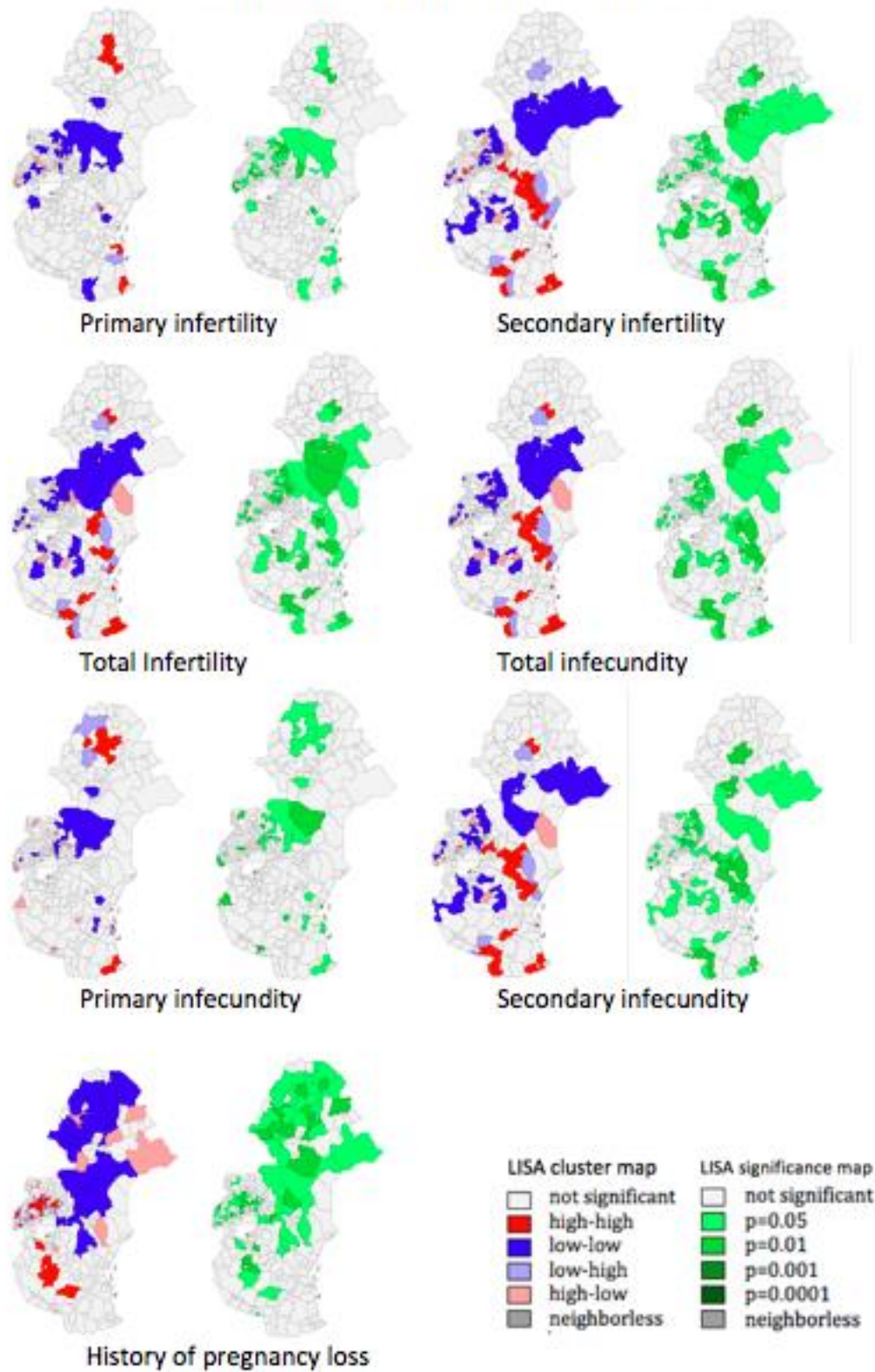


Figure 7. Measures of fertility impairment as box maps (quartiles, non-parametric data). Women age 15-50, Demographic and Health Surveys.

## 2009-2011 DHS Ethiopia, Kenya, Tanzania, Uganda, women age 15-50





1999-2001 DHS Ethiopia, Tanzania, Uganda, women age 15-50

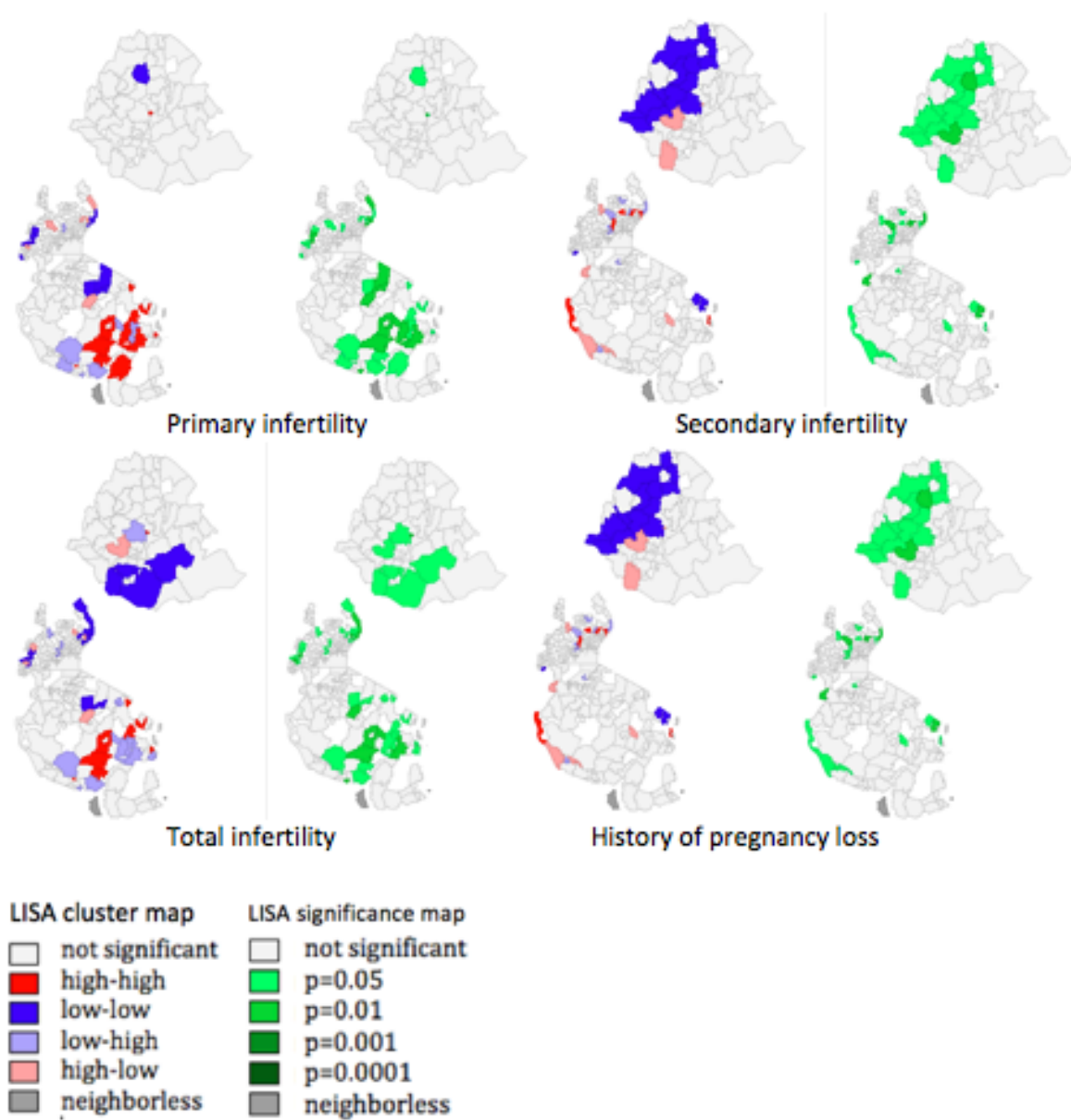


Figure 8. Measures of fertility impairment: Local Indices of Spatial Autocorrelation (LISA)