Distribution Agreement

In presenting this thesis or dissertation as a partial fulfillment of the requirements for an advanced degree from Emory University, I hereby grant to Emory University and its agents the non-exclusive license to archive, make accessible, and display my thesis or dissertation in whole or in part in all forms of media, now or hereafter known, including display on the world wide web. I understand that I may select some access restrictions as part of the online submission of this thesis or dissertation. I retain all ownership rights to the copyright of the thesis or dissertation. I also retain the right to use in future works (such as articles or books) all or part of this thesis or dissertation.

Signature:

Kaytna Thaker

Date

The concentration of malondialdehyde in exhaled breath is influenced by air pollution exposure during physical activity in adolescents

By

Kaytna Thaker Master of Public Health

Environmental Health

Roby Greenwald, PhD Committee Chair

Parinya Panuwet, PhD Committee Member

Paige Tolbert, PhD Committee Member The concentration of malondialdehyde in exhaled breath is influenced by air pollution exposure during physical activity in adolescents

By

Kaytna Thaker

B.S. Pennsylvania State University 2012

Thesis Committee Chair: Roby Greenwald, PhD

An abstract of A thesis submitted to the Faculty of the Rollins School of Public Health of Emory University in partial fulfillment of the requirements for the degree of Master of Public Health in Environmental Health 2014

Abstract

The concentration of malondialdehyde in exhaled breath is influenced by air pollution exposure during physical activity in adolescents By Kaytna Thaker

Background: The health effects of air pollution have long been a concern, and numerous studies have found associations between air pollution exposure and adverse health effects. One of the key mechanisms through which air pollutants exert toxicity is believed to be oxidative stress. Malondialdehyde (MDA) is a relatively stable common lipid peroxidation product and has been quantified in a variety of biological matrices, including exhaled breath condensate (EBC). **Objectives:** To determine the presence of an association between increased levels of MDA in EBC and increased concentrations of measured air pollutants within this study population of adolescents. *Methods:* Student athletes between the ages of 14 and 18 were recruited from sports practices at two metro-Atlanta high schools. Prior to, and immediately following sports team practices, samples of EBC were collected from each subject. Air quality measurements were recorded for the duration of team practices. MDA concentrations were measured in EBC using High Performance Liquid Chromatographic Mass Spectrometry following derivatization with 2,4-dinitrophenylhydrazine. Single pollutant models using linear regression were used to assess associations between percent change in MDA and pollutants of interest. Results: A non-significant decrease in MDA levels was observed when comparing pre and post-practice MDA concentration among all subject, however, a significant difference was observed in post-practice MDA levels between urban and suburban schools. A significant difference in the mean percent change in MDA levels among subjects participating in indoor and outdoor sports. A statistically significant association was found between particle number count and percent change in MDA levels and a borderline significant association was found between ozone and percent change in MDA levels. Controlling for gender, a statistically significant association was found between ozone and particle number count and percent change in MDA level. *Discussion:* MDA levels increased during sports team practices. Independent of exposure, outdoor sports are associated with increased percent change in MDA levels when compared to indoor sports. Although a significant difference was observed in the mean post-practice MDA concentration among urban and suburban schools, there was no significant difference in the overall percent change in MDA levels between the two schools.

The concentration of malondialdehyde in exhaled breath is influenced by air pollution exposure during physical activity in adolescents

By

Kaytna Thaker

B.S. Pennsylvania State University 2012

Thesis Committee Chair: Roby Greenwald, PhD

A thesis submitted to the Faculty of the Rollins School of Public Health of Emory University in partial fulfillment of the requirements for the degree of Master of Public Health in Environmental Health 2014

Acknowledgements

I would like to thank Dana Barr, Barry Ryan, Elizabeth Marder, Priya Kewada, Anastasia Lopukin, and Gabrielle Fanning-Dowdell for their contributions to this work.

Table of Contents

INTRODUCTION	1
Background	1
Children and Air Pollution	1
Health Effects of Air Pollution	3
Aims	4
METHODS	5
Study Population and Enrollment	5
EBC Sample Collection	6
MDA Standard Solution Preparation	6
EBC Sample Preparation and Derivitization	6
EBC Sample Quantification	7
Analytical Method Performance	8
Data Analysis	8
RESULTS	
Summary statistics	8
Differences in pre vs. post-practice MDA levels between subjects from urban and suburban schools	
Differences in mean MDA levels between subjects from urban and suburban schools	
Differences in pre vs. post-practice MDA levels between subjects participating in indoor and outdoor s	
Differences in mean MDA levels between subjects participating in indoor and outdoor sports	
Pollutants	
Associations between air pollutants and percent change in MDA levels	
DISCUSSION	
Limitations	
CONCLUSIONS	
FIGURES AND TABLES	
Table 1: Study subject characteristics	
Table 2: Mean pre and post-practice MDA concentrations (standard deviation) and M	
% change	
Table 3: Mean pre and post-practice MDA concentrations (standard deviation) and M	
% change	
Table 4: Mean pollutant concentrations (standard deviation) at urban and suburban	
schools and indoor and outdoor sports	23
Table 5: Single pollutant linear regression models examining the relationship between	
ozone, black carbon, and particle number count and percent change in MDA levels	24
Table 6: Single pollutant linear regression models including sex as additional predictor	
percent change in MDA level controlling for sex and age	
Figure 1: Differences in mean MDA levels between subjects from urban and suburbar	n
schools	
Figure 2: Differences in mean MDA levels between subjects participating in indoor an	ıd
outdoor sports	25
Appendix A: Full LC-MS Analytical Method	
Appendix B: MDA Calibration Curve	40

INTRODUCTION

Background

Ambient air pollution is a complex mixture of gases and particles that arise from a variety of sources. Anthropogenic sources of air pollution include stationary sources such as fossil fuel burning power plants and industrial facilities as well as mobile sources such as cars, trucks, and construction and agricultural equipment. The health effects of air pollution have been a concern for several decades, as evidenced by air quality standards mandated by the U.S. EPA and the European Union's Air Quality Limit Values. Exposure to ambient air pollution has been linked to numerous adverse health effects in populations throughout the world. Both short and long term studies have found associations between exposure to air pollutants and increased cardiovascular and respiratory morbidity and mortality, even at low levels of exposure.^{1,2} Recently air pollution has come under increased scrutiny, with the International Agency for Research on Cancer classifying outdoor air pollution as a carcinogen. Additionally, traffic-related pollutants including ozone and particulate matter have been associated with asthma exacerbation and airway hyperresponsiveness among children and cardiovascular disease mortality among adults. ³⁻⁶

Children and Air Pollution

Children in particular may be more susceptible to harmful effects of air pollutants compared to adults, as their lungs are not yet fully developed. Children may have greater exposures than adults, and children generally spend a greater proportion of time spend a lot of time outdoors and engage in physical activities that increase their breathing rate.⁷⁻⁹ Although lung development begins prenatally, with alveoli forming between fetal weeks

32-36, this process continues well into childhood.¹⁰ Structural and morphological features of the child lung begin to resemble those of an adult lung between ages 3 and 4. Using cell culture and animal models including non-human primates, toxicological studies have demonstrated exposure to air pollution alters the morphological construction of the airways, ultimately altering their functional capacity.¹¹⁻¹⁴ Researchers have observed in animal models that differentiation of the lung epithelium can be altered by insult at a very young age and because repair mechanisms are not fully developed, these alterations persist.¹⁵

Epidemiological studies indicate that exposure to air pollutants are associated with decreased lung growth. The Children's Health Study conducted in California from 1993 to 2001 observed deficits in lung function and decreased lung development correlated with PM_{2.5} and elemental carbon levels.¹⁶ However this study only evaluated lung development between the ages of 10 to 18 and did not consider deficits in development that may have resulted from exposure to air pollutants in early childhood. A sub-study within this cohort was conducted to evaluate the association between playing team sports and development of asthma during a 5 year follow up period.¹⁷ Researchers found that development of physician-diagnosed asthma was associated with high ambient levels of ozone. These results were consistent among both children with previous history of asthma symptoms and those who had not previously exhibited symptoms of asthma such as wheezing. A prospective cohort study conducted among Austria schoolchildren from 1994-1997 observed decreases in pulmonary function associated with PM_{10} , and Jedrychowski et al. also found consistent associations between air pollutants and decreased lung growth among preadolescent children in Krakow, Poland.^{18,19}

Health Effects of Air Pollution

Oxidative stress is believed to be one of the key underlying features of air pollutant toxicity.²⁰ Oxidative stress is generally described as an imbalance between free radicals (often reactive oxygen species or reactive nitrogen species) and antioxidants; excess oxidative products can damage DNA, form protein adducts, and damage lipid membranes leading to the formation of lipid peroxidation products.^{21,22} Although oxidative stress can target a variety of cellular components, lipids are the most involved class of biomolecule due to the abundance of lipids in the cell membrane and various other cellular constituents.²³

Using cell and animal models, toxicological studies report air pollution induced oxidative stress and additional human exposure studies have found indications of airway inflammation following exposure to traffic-related air pollution in both healthy and asthmatic subjects.²⁴⁻²⁸ Epidemiological studies have demonstrated that short term increases in pollutants such as fine particulate matter (PM_{2.5}), ozone, and elemental or black carbon are associated with increased airway oxidative stress in children and adolescents.^{29,30} Vossoughi et al. found evidence of increased airway and systemic inflammation in a cohort following long-term exposure to traffic and industrial air pollution, and numerous respiratory diseases are associated with airway inflammation, including asthma, chronic bronchitis, and chronic obstructive pulmonary disease.^{31,32}

Although air pollutants appear to exacerbate airway inflammation, the extent to which oxidative stress contributes to air pollution induced toxicity and airway inflammation remains unclear.^{20,33,34} As such, a variety of techniques have been developed to quantify biomarkers of airway inflammation and oxidative stress.

Noninvasive techniques such as the collection of exhaled breath condensate have proven useful. Exhaled breath condensate (EBC) is a fluid formed by cooling exhaled air. Although this fluid consists mostly of condensed water vapor, it also contains respiratory fluid droplets released from the lung epithelial lining fluid, which contain traces of volatile and non-volatile solutes that can be recovered in EBC samples.³⁵ Biomolecules including aldehydes, leukotrienes, 8-isoprostane, prostaglandins, hydrogen peroxide, nitric oxide-derived products, and hydrogen ions can be measured in EBC.³⁶

Malondialdehyde (MDA) is a relatively stable common lipid peroxidation product and has been quantified in a variety of biological matrices, including EBC. ³⁷⁻⁴¹ Several studies have reported increased levels of MDA along with increased levels of air pollutants. Barregard et al. reported higher levels of MDA following exposure to wood smoke, supporting the hypothesis that particulate air pollution causes oxidative stress and distal pulmonary inflammation.⁴² Among asthmatic schoolchildren in Mexico City, Romieu et al. reported associations between levels of MDA in EBC and levels of ozone and particulate matter.³⁹ Additionally, MDA levels were inversely associated with lung function. A study conducted during the 2008 Beijing Olympics found associations between MDA levels in healthy individuals and both single pollutants and multi-pollutant combinations, indicating the oxidative stress effects of air pollution.³⁸

Aims

Through the collection and analysis of health measurements and air quality data, this analysis aims to demonstrate an association between increased levels of MDA in EBC and increased levels of air pollutants. EBC samples will be analyzed to determine the levels of MDA present in each sample. MDA levels will be compared with air pollutant data to determine the presence of an association between increased levels of MDA in EBC and increased concentrations of measured air pollutants within this study population of adolescents.

METHODS

Study Population and Enrollment

The Study of Air Pollution and Physical Activity (SAPPA) seeks to determine the effect of physical activity upon air pollution exposure. The study primary investigator is Roby Greenwald, PhD (Dept. of Environmental Health), and it was funded by NIH grant # 1K25ES020355-01A1. The study protocol was reviewed and approved by Emory University's Institutional Review Board.

Demographic and health information was gathered through a questionnaire. The questionnaire included questions about date of birth, sex, race/ethnicity, height and weight, smoking status, and history of asthma.

Student athletes between the ages of 14 and 18 were recruited from sports practices at two metro-Atlanta high schools. Sports included cross country, football, soccer, basketball, track & field, and marching band. Study consent was obtained from the subjects, or in the case of subjects under the age of 18, from the subject's parent/guardian. Prior to, and immediately following sports team practices, biological samples and health measurements were obtained from each subject. Air quality measurements were recorded for the duration of team practices and constitute exposure. Measured levels of pollutants are representative of ambient concentrations to which subjects were exposed. Inhaled dose represents the amount of pollutants taken into the lungs and varies among subjects depending on breathing rate and lung capacity. For the purposes of this study, ambient concentrations were used as model inputs.

Subjects who self-identified as a current smoker were excluded from the study. A total of 179 subjects have participated in the study to date. For the purposes of this analysis, a subset of 58 subjects was randomly selected for analysis.

EBC Sample Collection

Samples of exhaled breath condensate (EBC) were collected from each subject prior to and following exposure using an R-tube® collection device with an aluminum sleeve chilled to -80° Celsius. Subjects breathed through the R-tube® for approximately 10-15 minutes, or until at least 1 mL of condensate was produced. An aliquot of 245 µL of EBC was added to 5 µL of butylated hydroxytoluene (BHT), to prevent further oxidation of MDA in the EBC fluid. Subsequently, EBC samples were stored at -80° C until analysis.

MDA Standard Solution Preparation

MDA standard solutions were prepared by dissolving malondialdehyde tetrabutylammonium salt in acetonitrile to yield a 10 mM stock solution. This stock solution was diluted with deionized H₂O to create yield a linear calibration solutions with concentrations ranging from 0.001 μ M to 1.0 μ M (Appendix B, Figure 2).

EBC Sample Preparation and Derivitization

Malondialdehyde (MDA) concentrations were measured in EBC using an adaptation of the method described by Chen et al.⁴³ First, EBC samples were thawed and vortexed. Then, a 50 μ L aliquot of each EBC sample was mixed with 25 μ L of an internal standard solution (1 μ M of d₂-MDA) and with 20 μ L of 1M HCl. The EBC samples were

derivitized using 25 μ L of a saturated 2,4-dinitrophenylhydrazine (DNPH) solution to form a DNPH-MDA adduct under an incubation temperature of 37 ° Celsius for 70 mins. Blank samples, consisted of deionized H₂O, and 7 standard calibrants were prepared concurrently with the EBC samples, using the same procedure, for each batch of sample analysis.

EBC Sample Quantification

EBC samples containing a DNPH-MDA adduct were separated and analyzed using a 6190 High Performance Liquid Chromatographic Mass Spectrometric (LC-MS) system (Agilent, USA). The analytical column was a C18 column, 150×4.6 mm I.D., particle size 5 µm (Phenomenex, USA). Column compartment temperature was set at 45° C. Mobile phases consisted of acetonitrile (containing 0.1% formic acid, v/v) and deionized water (containing 0.1% formic acid, v/v). The instrument flow rate was set at 0.8 mL/min. with an injection volume was 100µL.

Mass spectrometric analysis of the DNPH-MDA adduct was performed under a positive Electrospray Ionization (EIS), using a Single Ion Monitoring (SIM) technique. An *m/z* of 235 was monitored for native DNPH-MDA adduct, while an *m/z* of 237 was monitored for its label isotope analogue. Mass spectrometric parameters were as follows: 120 fragmentor, 594 dwell time, 360°C sheath gas temperature, 325°C drying gas temperature, 10 L/min sheath gas flow, 5 L/min drying gas flow, 35 psi nebulizer pressure, 4000V capillary voltage, and 800V nozzle voltage. See Appendix A for detailed method information.

Analytical Method Performance

The method limit of quantification is defined as the concentration of the lowest standard solution, which contained 0.001 μ M MDA. The accuracy of this method was assessed by taking subtracting the expected concentration of each standard from the observed concentration and averaging these values. Using these parameters, the accuracy of this method is 102%. The method precision, reported as an average relative standard deviation (RSD), was below 15%. Note that all analytical parameters reported here were in compliance with the requirements set by the FDA. *Data Analysis*

Analyses were performed using SAS version 9.3. Exploratory analyses were performed and values for each variable. Univariate summaries were used to analyze continuous variables and relative frequency tables were used to analyze categorical variables and check for implausible or missing values. Exposures were recorded as mean concentration of each pollutant of interest. Single pollutant models using linear regression were used to assess associations between percent change in MDA and pollutants of

RESULTS

interest.

Summary statistics

A subset of 58 subjects was randomly selected for this analysis. Subjects were recruited at various high school sports team practices. Table 1 presents the characteristics of the study subjects. Prior to and immediately following sports team practices, each subject completed a series of health measurements and provided samples of exhaled breath condensate. Subjects with an MDA % change greater than 3 standard deviations from the mean were removed from analysis. Results are based on the remaining 52 subjects. Mean MDA concentrations before and after sports team practices and MDA % change are presented in Table 2.

			Gender		Spe	ort
School	Total Subjects	Mean Age (yr)	Male	Female	Indoor	Outdoor
Urban	34	16.88 <u>+</u> 1.14	33	1	19	15
Suburban	18	17.03 ± 1.16	6	12	6	12

Table 1	:: S	Study	sub	ject o	characteristics
---------	------	-------	-----	--------	-----------------

 Table 2: Mean pre and post-practice MDA concentrations (standard deviation) and

 MDA % change

	Urban School	Suburban School	All Subjects
Pre-practice (µM)	0.0144 (0.0228)	0.0158 (0.0127)	0.0149 (0.0198)
Post-practice (µM)	0.0120 (0.0066)	0.0268 (0.0299)	0.0171 (0.0194)
Percent Change (%)	42.85 (84.00)	81.06 (109.75)	56.07 (94.43)

Differences in pre vs. post-practice MDA levels between subjects from urban and suburban schools

Using the mean MDA concentrations as indicated in Table 2, a non-significant decrease in MDA levels was observed when comparing post and pre-practice MDA concentration among all subjects (-0.0022 μ M, p=0.4569). A similar non-significant increase was observed for subjects from the urban school (0.0024 μ M, p=0.5113), however a significant decrease in post vs. pre-practice MDA levels was observed among subjects from the suburban school (-0.0110 μ M, p=0.0358). These differences do not take into account air pollutant levels.

Differences in mean MDA levels between subjects from urban and suburban schools

The mean pre and post-practice MDA levels and percent change in MDA among subjects at urban and suburban schools were compared using one-way ANOVA (Figure 1). No significant difference was observed in pre-practice MDA levels (p=0.8064) or percent change in MDA (p=0.1673), however a significant difference was observed in post-practice MDA levels between urban and suburban schools (p=0.0076).

Figure 1: Differences in mean MDA levels between subjects from urban and suburban schools



Differences in pre vs. post-practice MDA levels between subjects participating in

indoor and outdoor sports

Using the mean MDA concentrations as indicated in Table 3, a non-significant decrease in MDA levels was observed when comparing post and pre-practice MDA concentration among all subjects (-0.0022 μ M, p=0.4569). A non-significant increase was observed for subjects participating in indoor sports (0.0019 μ M, p=0.7614), however a borderline significant decrease in post vs. pre-practice MDA levels was observed among

subjects participating in outdoor sports (-0.0026 μ M, p=0.0594). These differences do not take into account air pollutant levels.

Table 3: Mean pre and post-practice MDA concentrations (standard deviation) and MDA % change

	Indoor Sports	Outdoor Sports	All Subjects
Pre-practice (µM)	0.0220 (0.0254)	0.0084 (0.0090)	0.0149 (0.0198)
Post-practice (µM)	0.0238 (0.0253)	0.0110 (0.0081)	0.0171 (0.0194)
Percent Change (%)	24.8 (70.15)	85.03 (05.54)	56.07 (94.43)

Differences in mean MDA levels between subjects participating in indoor and outdoor

sports

The mean pre and post-practice MDA levels and percent change in MDA among subjects participating in indoor and outdoor sports were compared using one-way ANOVA (Figure 2). A significant difference was observed in pre-practice MDA levels (p=0.012) and post-practice MDA levels (p=0.0153). Additionally, a significant difference in the percent change in MDA levels (p=0.02).



Figure 2: Differences in mean MDA levels between subjects participating in indoor and outdoor sports

Pollutants

Ozone, black carbon, and particle number count were recorded during sports team practices. Mean concentrations of each pollutant are presented in Table 4. Both ozone and black carbon mean concentrations are greater at the suburban school compared to the urban school. Ozone, black carbon, and particle number count are all substantially greater among outdoor sports when compared to indoor sports.

 Table 4: Mean pollutant concentrations (standard deviation) at urban and suburban schools and indoor and outdoor sports

School			Sport		
Pollutant	Urban School	Suburban School	Indoor Sports	Outdoor Sports	
Ozone (ppb)	20 (18)	33 (22)	5 (0)	42 (11)	
Black carbon (µg/m ³)	0.20 (0.22)	0.29 (0.41)	0.013 (0.009)	0.33 (0.32)	
Particle number count (#/cm ³)	9434 (5848)	2906 (598)	5364 (1544)	9854 (7366)	

Associations between air pollutants and percent change in MDA levels

Single pollutant linear regression models were used to assess associations between air pollutants and percent change in MDA levels. As indicated in Table 5, no statistically significant association was found black carbon and percent change in MDA levels. A statistically significant association was found between particle number count and percent change in MDA levels and a borderline significant association was found between ozone and percent change in MDA levels. Using the interquartile range (IQR), an increase of 37 ppb in ozone levels is associated with a 45.9% change in MDA levels and an increase of 3365 particles/cm³ is associated with a 17.16% increase in MDA levels. When multi-pollutant linear regression models were considered, no statistically significant associations at the α =0.05 level were found between pollutants and percent change in MDA level.

Table 5: Single pollutant linear regression models examining the relationship between ozone, black carbon, and particle number count and percent change in MDA levels

Pollutant	Parameter estimate	Standard error	P-value
Ozone (ppb)	1.24	0.63	0.0563
Black carbon (µg/m ³)	-31.04	54.84	0.5749
Particle number count (#/cm ³)	0.0051	0.0023	0.0350

When sex and age are included as additional predictors a statistically significant association was found between ozone and particle number count and percent change in MDA level. Because age was not a significant predictor of percent change in MDA levels, it was removed from the models. Gender was coded as 0/1, with females (coded as 1) exhibiting a larger percent change. Controlling for gender, a 37 ppb increase in ozone is associated with a 47.4% increase in percent change of MDA levels and an increase of

3365 particles/cm³ is associated with a 26.25% increase in MDA levels.

Equation 1: Single pollutant linear regression model controlling for sex

Percent change MDA = $\beta_0 + \beta_1$ *Pollutant concentration + β_2 *Gender

Pollutant	Parameter Estimate	Standard Error	P-value
Ozone	1.28	0.59	0.035
Sex	81.21	27.39	0.0047
Black carbon	-17.53	54.67	0.750
Sex	57.69	38.53	0.143
Particle number count	0.0078	0.002	0.002
Sex	87.72	32.18	0.0064

 Table 6: Single pollutant linear regression models including sex as additional predictors of percent change in MDA level controlling for sex and age

When considering multi-pollutant linear regression models including age and sex as additional predictors, no statistically significant association at the α =0.05 level was found between pollutants and percent change in MDA level.

DISCUSSION

The mean percent change in MDA levels among all subjects was 56% and when stratifying by urban and suburban school, the mean percent change in MDA levels was 43% and 81% respectively. These values indicate that MDA levels increased during sports team practices which constituted the exposure period. Independent of exposure, outdoor sports are associated with increased percent change in MDA levels when compared to indoor sports. Several studies have reported increased MDA levels following exposure to traffic related air pollution, and these results are in agreement.^{38,39} Although a significant difference was observed in the mean post-practice MDA concentration among urban and suburban schools, there was no significant difference in the overall percent change in MDA levels between the two schools.

Previous studies have found increased levels of MDA in EBC following exposure to traffic-related pollutants including ozone, black carbon, and PM_{2.5} among asthmatic and healthy subjects. ^{38,39} Additionally, exposure to traffic-related pollutants has been associated with decreased lung function, asthma exacerbation, and airway inflammation.^{4,16,44} In this study, particle number count was found to be significantly associated with percent change in MDA levels. Although not statistically significant, the results of this study suggest that ozone may be associated with percent change in MDA levels. Black carbon was not found to be significantly associated with percent change in MDA levels.

For the purposes of this study, ambient concentrations were used as model inputs. It is for this reason that exposure may not necessarily reflect inhaled dose. Additionally, exposure levels for ozone, particle number count, and black carbon were relatively low compared to other studies. This is due to the fact that concentrations of air pollutants in indoor environments are typically lower than outdoor concentrations. Also, levels of measured pollutants tend to increase during the summer months in Atlanta. However, when compared to previous summers, the summer of 2013 was unusually cool and rainy. The relatively small number of subjects included in this analysis is also a limitation. Ongoing analysis of additional samples will improve our ability to determine the magnitude of the associations between pollutants and percent change in MDA levels.

When controlling for age and gender, both particle number count and ozone were found to be significantly associated with percent change in MDA levels. Age does not appear to have a significant impact upon percent change in MDA levels, however based on the results of this analysis, gender appears to be a significant predictor of the percent change in MDA levels. This association between gender and percent change in MDA levels has not been observed in previous studies. Further analyses are needed to determine if this association is merely a function of the small sample size of this analysis. The significant associations between particle number count and percent change in MDA levels are in agreement with previous studies. However the results of this analysis failed to find a significant association between ozone and black carbon and percent change in MDA levels.

The influence of physical activity upon MDA levels remains unclear.^{45,46} The results of this study indicate that MDA levels increased over the course of sports team practices, presumably while subjects were engaging in physical activity. A greater percent change in MDA levels was observed among subjects participating in outdoor sports (85%) compared to subjects participating in indoor sports (25%). These results appear to indicate that independent of exposure to air pollutants, engaging in physical activity corresponds to an increase in MDA levels.

Limitations

Due to the noninvasive nature of collection, EBC is a promising matrix in which a variety of biomolecules can be quantified. However, variability exists in the collection of EBC. Although aluminum sleeves are originally chilled to -80° Celsius, over time the temperature of sleeves increases. Ambient temperature has been found to impact the pH of EBC samples which in turn can affect the electrochemical properties and reaction

kinetics of the EBC.⁴⁷ The R-tube® collection devices are designed in a manner so as to limit salivary contamination of EBC fluid; however the method used in this analysis cannot determine if samples of EBC have been contaminated with saliva. It is necessary to test for high concentrations of phosphate or amylase to determine if samples are contaminated.

The high degree of variability in measurement is a limitation of the EBC matrix. Because water vapor is the major constituent of EBC, the concentration of analytes, including malondialdehyde, exhibit significant interindividual variations as well as intraindividual variations.⁴⁸ Numerous studies have compared levels of MDA across individual subjects, however in order to do so more accurately, further research is necessary to determine an accurate method of quantifying the dilution factor of EBC samples.⁴⁹ EBC was collected at specific time points (immediately before and after sports team practices) in order to best capture variations in MDA concentrations due to exposure however further research is necessary to determine the optimal collection time point.

CONCLUSIONS

The data from this study indicate that exposure to particulate matter and ozone is associated with increased MDA levels although further research is needed to determine the true magnitude of this association. This study focuses on children, as they may be more susceptible to adverse health effects of air pollution exposure compared to adults. Additionally, the associations observed in this study provide further insight into the pollutants most likely to induce oxidative stress and airway inflammation. Increased

References

- **1.** Brunekreef B, Holgate ST. Air pollution and health. *The Lancet*. 2002;360(9341):1233-1242.
- 2. Samet JM, Dominici F, Curriero FC, Coursac I, Zeger SL. Fine particulate air pollution and mortality in 20 U.S. cities, 1987-1994. *The New England journal of medicine*. Dec 14 2000;343(24):1742-1749.
- **3.** Jerrett M, Shankardass K, Berhane K, et al. Traffic-related air pollution and asthma onset in children: a prospective cohort study with individual exposure measurement. *Environ Health Perspect*. Oct 2008;116(10):1433-1438.
- **4.** Evans KA, Halterman JS, Hopke PK, Fagnano M, Rich DQ. Increased ultrafine particles and carbon monoxide concentrations are associated with asthma exacerbation among urban children. *Environ Res.* 2014;129(0):11-19.
- 5. Beelen R, Stafoggia M, Raaschou-Nielsen O, et al. Long-term Exposure to Air Pollution and Cardiovascular Mortality: An Analysis of 22 European Cohorts. *Epidemiology*. Feb 28 2014.
- 6. Kim B-J, Lee S-Y, Kwon J-W, et al. Traffic-related air pollution is associated with airway hyperresponsiveness. *J Allergy Clin Immun*. (0).
- 7. La Grutta S, Indinnimeo L, di Coste A, et al. Environmental risk factors and lung diseases in children: From guidelines to health effects. *Early Human Development*. 2013;89, Supplement 3(0):S59-S62.
- 8. Bateson TF, Schwartz J. Children's response to air pollutants. *Journal of toxicology and environmental health. Part A.* 2008;71(3):238-243.
- **9.** Esposito S, Tenconi R, Lelii M, et al. Possible molecular mechanisms linking air pollution and asthma in children. *BMC pulmonary medicine*. 2014;14(1):31.
- **10.** Burri PH. Structural Aspects of Postnatal Lung Development Alveolar Formation and Growth. *Neonatology*. 2006;89(4):313-322.
- **11.** Fanucchi MV, Buckpitt AR, Murphy ME, Plopper CG. Naphthalene Cytotoxicity of Differentiating Clara Cells in Neonatal Mice. *Toxicology and Applied Pharmacology*. 1997;144(1):96-104.
- 12. Lakari E, PÄÄKkÖ P, Pietarinen-Runtti P, Kinnula VL. Manganese Superoxide Dismutase and Catalase Are Coordinately Expressed in the Alveolar Region in Chronic Interstitial Pneumonias and Granulomatous Diseases of the Lung. *American journal of respiratory and critical care medicine*. 2000/02/01 2000;161(2):615-621.
- **13.** Plopper CG, Weir AJ, Nishio SJ, et al. Elevated susceptibility to 4-ipomeanol cytotoxicity in immature Clara cells of neonatal rabbits. *Journal of Pharmacology and Experimental Therapeutics*. May 1, 1994 1994;269(2):867-880.
- **14.** Fanucchi MV, Day KC, Clay CC, Plopper CG. Increased vulnerability of neonatal rats and mice to 1-nitronaphthalene-induced pulmonary injury. *Toxicology and Applied Pharmacology*. 2004;201(1):53-65.
- **15.** Smiley-Jewell SM, Nishio SJ, Weir AJ, Plopper CG. Neonatal Clara cell toxicity by 4-ipomeanol alters bronchiolar organization in adult rabbits. *American Journal of Physiology Lung Cellular and Molecular Physiology*. 1998;274(4):L485-L498.

- **16.** Gauderman WJ, Avol E, Gilliland F, et al. The effect of air pollution on lung development from 10 to 18 years of age. *The New England journal of medicine*. Sep 9 2004;351(11):1057-1067.
- **17.** McConnell R, Berhane K, Gilliland F, et al. Asthma in exercising children exposed to ozone: a cohort study. *Lancet.* Feb 2 2002;359(9304):386-391.
- **18.** Jedrychowski W, Flak E, Mroz E. The adverse effect of low levels of ambient air pollutants on lung function growth in preadolescent children. *Environ Health Perspect.* Aug 1999;107(8):669-674.
- **19.** Horak F, Studnicka M, Gartner C, et al. Particulate matter and lung function growth in children: a 3-yr follow-up study in Austrian schoolchildren. *European Respiratory Journal*. May 1, 2002 2002;19(5):838-845.
- **20.** Romieu I, Castro-Giner F, Kunzli N, Sunyer J. Air pollution, oxidative stress and dietary supplementation: a review. *European Respiratory Journal*. January 1, 2008 2008;31(1):179-197.
- **21.** Bowler RP, Crapo JD. Oxidative stress in allergic respiratory diseases. *J Allergy Clin Immun.* 2002;110(3):349-356.
- **22.** Valavanidis A, Vlachogianni T, Fiotakis K, Loridas S. Pulmonary oxidative stress, inflammation and cancer: respirable particulate matter, fibrous dusts and ozone as major causes of lung carcinogenesis through reactive oxygen species mechanisms. *International journal of environmental research and public health*. Sep 2013;10(9):3886-3907.
- **23.** Del Rio D, Stewart AJ, Pellegrini N. A review of recent studies on malondialdehyde as toxic molecule and biological marker of oxidative stress. *Nutrition, Metabolism and Cardiovascular Diseases.* 2005;15(4):316-328.
- **24.** Li R, Ning Z, Cui J, et al. Ultrafine particles from diesel engines induce vascular oxidative stress via JNK activation. *Free Radical Biology and Medicine*. 2009;46(6):775-782.
- **25.** Araujo JA, Barajas B, Kleinman M, et al. Ambient Particulate Pollutants in the Ultrafine Range Promote Early Atherosclerosis and Systemic Oxidative Stress. *Circulation Research.* March 14, 2008 2008;102(5):589-596.
- **26.** González-Flecha B. Oxidant mechanisms in response to ambient air particles. *Molecular Aspects of Medicine*. 2004;25(1–2):169-182.
- 27. Salvi S, Blomberg A, Rudell B, et al. Acute inflammatory responses in the airways and peripheral blood after short-term exposure to diesel exhaust in healthy human volunteers. *American journal of respiratory and critical care medicine*. Mar 1999;159(3):702-709.
- **28.** Stenfors N, Nordenhäll C, Salvi SS, et al. Different airway inflammatory responses in asthmatic and healthy humans exposed to diesel. *European Respiratory Journal*. January 1, 2004 2004;23(1):82-86.
- **29.** Delfino RJ, Staimer N, Gillen D, et al. Personal and ambient air pollution is associated with increased exhaled nitric oxide in children with asthma. *Environ Health Persp.* Nov 2006;114(11):1736-1743.
- **30.** Barraza-Villarreal A, Sunyer J, Hernandez-Cadena L, et al. Air pollution, airway inflammation, and lung function in a cohort study of Mexico City schoolchildren. *Environ Health Persp.* Jun 2008;116(6):832-838.

- **31.** Shelhamer JH, Levine SJ, Wu T, Jacoby DB, Kaliner MA, Rennard SI. NIH conference. Airway inflammation. *Annals of internal medicine*. Aug 15 1995;123(4):288-304.
- **32.** Vossoughi M, Schikowski T, Vierkotter A, et al. Air pollution and subclinical airway inflammation in the SALIA cohort study. *Immunity & ageing : I & A*. Mar 19 2014;11(1):5.
- **33.** Liu L, Poon R, Chen L, et al. Acute Effects of Air Pollution on Pulmonary Function, Airway Inflammation, and Oxidative Stress in Asthmatic Children. *Environ Health Persp.* Apr 2009;117(4):668-674.
- **34.** Chen B-Y, Chan C-C, Lee C-T, et al. The Association of Ambient Air Pollution With Airway Inflammation in Schoolchildren. *American journal of epidemiology*. April 15, 2012 2012;175(8):764-774.
- **35.** Rosias PPR, Dompeling E, Hendriks HJE, Heijnens JWCM, Donckerwolcke RAMG, Jöbsis Q. Exhaled breath condensate in children: Pearls and pitfalls. *Pediatric Allergy and Immunology*. 2004;15(1):4-19.
- **36.** Montuschi P. Review: Analysis of exhaled breath condensate in respiratory medicine: methodological aspects and potential clinical applications. *Therapeutic Advances in Respiratory Disease*. October 1, 2007 2007;1(1):5-23.
- **37.** Bartoli ML, Novelli F, Costa F, et al. Malondialdehyde in Exhaled Breath Condensate as a Marker of Oxidative Stress in Different Pulmonary Diseases. *Mediators of Inflammation*. 2011;2011.
- **38.** Gong JC, Zhu T, Kipen H, et al. Malondialdehyde in exhaled breath condensate and urine as a biomarker of air pollution induced oxidative stress. *J Expo Sci Env Epid.* May-Jun 2013;23(3):322-327.
- **39.** Romieu I, Barraza-Villarreal A, Escamilla-Nunez C, et al. Exhaled breath malondialdehyde as a marker of effect of exposure to air pollution in children with asthma. *J Allergy Clin Immun.* Apr 2008;121(4):903-909.
- **40.** Larstad M, Ljungkvist G, Olin AC, Toren K. Determination of malondialdehyde in breath condensate by high-performance liquid chromatography with fluorescence detection. *J Chromatogr B*. Jan 5 2002;766(1):107-114.
- **41.** Andreoli R, Manini P, Corradi M, Mutti A, Niessen WMA. Determination of patterns of biologically relevant aldehydes in exhaled breath condensate of healthy subjects by liquid chromatography/atmospheric chemical ionization tandem mass spectrometry. *Rapid Communications in Mass Spectrometry*. 2003;17(7):637-645.
- **42.** Barregard L, Sällsten G, Andersson L, et al. Experimental exposure to wood smoke: effects on airway inflammation and oxidative stress. *Occupational and environmental medicine*. May 1, 2008 2008;65(5):319-324.
- **43.** Chen JL, Huang YJ, Pan CH, Hu CW, Chao MR. Determination of urinary malondialdehyde by isotope dilution LC-MS/MS with automated solid-phase extraction: a cautionary note on derivatization optimization. *Free radical biology* & *medicine*. Nov 1 2011;51(9):1823-1829.
- **44.** McCreanor J, Cullinan P, Nieuwenhuijsen MJ, et al. Respiratory Effects of Exposure to Diesel Traffic in Persons with Asthma. *New England Journal of Medicine*. 2007;357(23):2348-2358.

- **45.** Child RB, Wilkinson DM, Fallowfield JL, Donnelly AE. Elevated serum antioxidant capacity and plasma malondialdehyde concentration in response to a simulated half-marathon run. *Medicine and science in sports and exercise*. Nov 1998;30(11):1603-1607.
- **46.** Clarkson PM, Thompson HS. Antioxidants: what role do they play in physical activity and health? *The American Journal of Clinical Nutrition*. August 1, 2000 2000;72(2):637s-646s.
- **47.** Koczulla AR, Noeske S, Herr C, et al. Ambient temperature impacts on pH of exhaled breath condensate. *Respirology*. 2010;15(1):155-159.
- **48.** Kubáň P, Foret F. Exhaled breath condensate: Determination of non-volatile compounds and their potential for clinical diagnosis and monitoring. A review. *Analytica Chimica Acta*. 2013;805(0):1-18.
- **49.** Horváth I, Hunt J, Barnes PJ, Condensate ObotAETFoEB. Exhaled breath condensate: methodological recommendations and unresolved questions. *European Respiratory Journal*. September 1, 2005 2005;26(3):523-548.

FIGURES AND TABLES

School	Mean Age (yr)	Gender (M/F)	Sport (Indoor/Outdoor)	Total Subjects
Urban	16.88 ± 1.14	33/1	19/15	34
Suburban	17.03 <u>+</u> 1.16	6/12	6/12	18

Table 1: Study subject characteristics

Table 2: Mean pre and post-practice MDA concentrations (standard deviation) and MDA % change

	Urban School	Suburban School	All Subjects
Pre-practice (µM)	0.0144 (0.0228)	0.0158 (0.0127)	0.0149 (0.0198)
Post-practice (µM)	0.0120 (0.0066)	0.0268 (0.0299)	0.0171 (0.0194)
Percent Change (%)	42.85 (84.00)	81.06 (109.75)	56.07 (94.43)

Table 3: Mean pre and post-practice MDA concentrations (standard deviation) and MDA % change

T	Indoor Sports	Outdoor Sports	All Subjects
Pre-practice (µM)	0.0220 (0.0254)	0.0084 (0.0090)	0.0149 (0.0198)
Post-practice (µM)	0.0238 (0.0253)	0.0110 (0.0081)	0.0171 (0.0194)
Percent Change (%)	24.8 (70.15)	85.03 (05.54)	56.07 (94.43)

Table 4: Mean pollutant concentrations (standard deviation) at urban and suburban schools and indoor and outdoor sports

	School		Sport	
Pollutant	Urban School	Suburban	Indoor Sports	Outdoor
		School		Sports
Ozone (ppb)	20 (18)	33 (22)	5 (0)	42 (11)
Black carbon (µg/m ³)	0.20 (0.22)	0.29 (0.41)	0.013 (0.009)	0.33 (0.32)
Particle number count (#/cm ³)	9434 (5848)	2906 (598)	5364 (1544)	9854 (7366)

Pollutant	Parameter estimate	Standard error	P-value
Ozone (ppb)	1.24	0.63	0.0563
Black carbon (µg/m ³)	-31.04	54.84	0.5749
Particle number count (#/cm ³)	0.0051	0.0023	0.0350

Table 5: Single pollutant linear regression models examining the relationship between ozone, black carbon, and particle number count and percent change in MDA levels

Table 6: Single pollutant linear regression models including sex as additional predictors of percent change in MDA level controlling for sex and age

Pollutant	Parameter Estimate	Standard Error	P-value
Ozone	1.28	0.59	0.035
Sex	81.21	27.39	0.0047
Black carbon	-17.53	54.67	0.750
Sex	57.69	38.53	0.143
Particle number count	0.0078	0.002	0.002
Sex	87.72	32.18	0.0064





Figure 1: Differences in mean MDA levels between subjects from urban and suburban





Appendix A: Full LC-MS Analytical Method

Method Information

Method: C:\Chem32\1\METHODS\MDA-DNPH_F2.M Modified: 3/5/2014 at 2:21:11 PM

MS final method for MDA-DNPH: Positive SIM mode

Method Audit Trail

Operator : Date : 3/5/2014 12:50:53 PM Change Info: This method was created at 3/5/2014 12:50:53 PM and based on method C:\Chem32\1\METHODS\MDA-DNPH_F.M

Operator : Date : 3/5/2014 12:51:07 PM Change Info: Method saved. User comment: "Test Meth 030414"

Operator : Date : 3/5/2014 12:58:55 PM Change Info: Method saved. User comment: ""

Operator : Date : 3/5/2014 2:21:11 PM Change Info: Method saved. User comment: ""

Run Time Checklist

Pre-Run Cmd/Macro: off

Data Acquisition: on

Standard Data Analysis: off

Customized Data Analysis: off

Save GLP Data: off

Post-Run Cmd/Macro: off

Save Method with Data: off

======= Binary Pump =======

Binary Pump (G4220A)

Flow: Use Solvent Types: Stroke Mode: Low Pressure Limit: High Pressure Limit: Max. Flow Ramp Up: Max. Flow Ramp Down: Expected Mixer:	0.100 ml/min Yes Synchronized 0.00 bar 550.00 bar 100.000 ml/min ² 100.000 ml/min ² No check	
Stroke A Automatic Stroke Calculation A	A: Yes	
Compress A Compressibility Mode A: Compressibility A:	Compressibility Value Set 45 10e-6/bar	
Compress B Compressibility Mode B: Compressibility B:	Compressibility Value Set 75 10e-6/bar	
Stop Time Stoptime Mode: Stoptime:	Time set 10.00 min	
Post Time Posttime Mode: Posttime:	Time set 1.00 min	
Timetable Time Function Paramete	r	
0.00 Change FlowFlow: 0.8 ml/min1.00 Change Solvent Com. Solvent composition A: 35.0 % B:65.0 %5.00 Change Solvent Com. Solvent composition A: 40.0 % B:60.0 %5.50 Change Solvent Com. Solvent composition A: 0.0 % B:100.0 %		

8.00 Change Solvent Com. Solvent composition A: 0.0 % B:100.0 %
8.50 Change Solvent Com. Solvent composition A: 50.0 % B:50.0 %
10.00 Change Solvent Com. Solvent composition A: 50.0 % B:50.0 %

Solvent Composition
ChannelCh. 1 Solv.Name 1

A 100.0 % Water V.02 0.1% NH4OH B 100.0 % Acetonitrile V.02 0.1% Formic Acid

Ch2 Solv. Name 2 Selected Used

100.0 % Water V.02 0.1% Formic AcidCh. 2Yes100.0 % Methanol V.020.1% Acetic AcidCh. 1Yes

Percent %

.....

50.0 50.0

Instrument Curves	
Store Pressure:	Yes
Store Flow:	Yes
Store Solvent Ratio A:	Yes
Store Solvent Ratio B:	Yes

HiP Sampler

HiP Sampler (G4226A)

Post Time Posttime Mode:	Off
Auxiliary Draw Speed: Eject Speed: Draw Position Offset: Wait Time After Drawing: Sample Flush Out Factor: Vial/Well bottom sensing:	100.0 μl/min 200.0 μl/min 0.0 mm 2.0 s 5.0 No
Injection Injection Mode: Injection Volume:	Standard injection 20.00 µL
High throughput Valve To Bypass After Injection	n Enabled: No
Overlapped Injection Enable Overlapped Injection:	No
Valve Switching Valve Movements:	0
Valve Switch Time 1 Switch Time 1 Enabled:	No
Valve Switch Time 2 Switch Time 2 Enabled:	No
Valve Switch Time 3 Switch Time 3 Enabled:	No
Valve Switch Time 4 Switch Time 4 Enabled:	No
Stop Time Stoptime Mode:	As pump/No limit

Timetable

HiP Sampler (G4226A)

Function	Parameter
Repeat	Repeat 4 time(s)
Draw	Draw 20 µL from sample with default spee
	d using default offset
Wait	Wait 0.02 min
Eject	Eject default volume to seat with defau
	lt speed using default offset
End Repeat	End Repeat
Draw	Draw 20 µL from sample with default spee
	d using default offset
Wait	Wait 0.02 min
Inject	Inject

====== Column Comp.		
Column Comp. (G1316C)		
Ready when front door open:	Yes	
Left Temperature Control Temperature Control Mode: Temperature:	Temperature Set 45.00 °C	
Enable Analysis Left Temperature		

Enable Analysis Left Temperature Enable Analysis Left Temperature		Yes 0.80 °C
Right Temperature Control Right temperature Control Mode:		Combined
Enable Analysis Right Temperatu Enable Analysis Right Temperatu Enable Analysis Right Temperatu	re On:	Yes 0.80 °C
Stop Time Stoptime Mode:	As pump/i	njector
Post Time Posttime Mode:	Of	f
Timetable		
Instrument Curves Store Left Temperature: Store Right Temperature:		No No

Mass Spectrometer Detector

_____ _____

General Information

Use MSD	: Enabled
Tune File	: atunes.tun
StopTime	: No Limit
Time Filter	: Enabled
Data Storage	: Condensed
Peakwidth	: 0.20 min
Fast Scan	: Disabled

Fast Scan Data Reconstruction: Disabled Polarity Switch Delay : 20 ms Ionization Switch Delay : 50 ms

Signals

[Signal 1]

Not Active

[Signal 2]

Ionization Mode: AJS-ESPolarity: PositiveFragmentor Ramp: Not ApplicablePercent Cycle Time: 100.00 %Sim On Target Mass: Disabled

Sim Parameters

[Signal 3]

Not Active

[Signal 4]

Not Active

Spray Chamber

[MSZones]

SGas Temp	: 360 C	maximum 360 C
Gas Temp	: 325 C	maximum 350 C
DryingGas	: 5.0 l/min	maximum 13.0 l/min
SGas Flow	: 10.0 l/min	maximum 12.0 l/min
Neb Pres	: 35 psig	maximum 60 psig
$\mathbf{V}\mathbf{C}$. 4000 17	

VCap (Positive)	:4000 V
VCap (Negative)	: 4000 V
NozzleVolt (Positive)	: 800 V
NozzleVolt (Negative) : 800 V

[Time Table]

Time Table is empty.

END OF MS ACQUISITION PARAMETERS

==== **FIA Series** ____ FIA Series in this Method : Disabled Time Setting Time between Injections : 0.15 min Injection Loop Flush Time : 0.17 min

Estimated Sample Purity Calculation

General

Estimated Sample Purity Calculation is disabled.

The Data Analysis Parameters of the used Method are :

=======

Integration Events

Non signal specific	c Integration Events

Event	Value
Tangent Skim Mode	Standard
Baseline Correction	Advanced
Tail Peak Skim Height Ratio	0.000
Front Peak Skim Height Ratio	0.000
Skim Valley Ratio	20.000
Peak to Valley Ratio	500.000

Default Integration Event Table "Event"

Event	Value Time
Initial Slope Sensitivity Initial Peak Width Initial Area Reject Initial Height Reject Initial Shoulders	 1.000 Initial 0.020 Initial 1.000 Initial 1.700 Initial OFF Initial

Detector Default Integration Event Table "Event_DAD"

Event	Value Time
Initial Slope Sensitivity Initial Peak Width Initial Area Reject Initial Height Reject Initial Shoulders	 5.000 Initial 0.020 Initial 5.000 Initial 1.000 Initial OFF Initial

Event	Value Time
Initial Slope Sensitivity	1.000 Initial
Initial Peak Width	0.020 Initial
Initial Area Reject	1.000 Initial
Initial Height Reject	1.700 Initial
Initial Shoulders	OFF Initial
Detector Default Integration	
Event	Value Time
Initial Slope Sensitivity	1.000 Initial
Initial Peak Width	0.020 Initial
Initial Area Reject	1.000 Initial
Initial Height Reject	1.700 Initial
Initial Shoulders	OFF Initial
Detector Default Integration Event	Value Time
Initial Slope Sensitivity Initial Peak Width	10000.000 Initial 0.020 Initial
Initial Area Reject	1000.000 Initial
Initial Height Reject	100.000 Initial
Initial Shoulders	OFF Initial
Signal Specific Integration Ev	
Event	Value Time
Initial Slope Sensitivity	100000.000 Initial
Initial Peak Width	0.020 Initial
Initial Area Reject	10000.000 Initial
Initial Height Reject	1000.000 Initial
Initial Shoulders	OFF Initial

Detector Default Integration Event Table "Event_ADC"

Signal Specific Integration Event Table "Event_MSD1SPC"

Event	Value Time
Initial Slope Sensitivity(Full Scan)	1.000 Initial
Initial Peak Width(Full Scan)	0.250 Initial
Initial Slope Sensitivity(Cond. Scar	n/SIM) 0.100 Initial
Initial Peak Width(Cond. Scan/SIM	I) 0.050 Initial
Initial Area Reject	0.000 Initial
Initial Height Reject	5.000 Initial
Initial Shoulders	OFF Initial

Detector Default Integration Event Table "Event_VWD"

Event	alue 7	Гime
Initial Slope Sensitivity Initial Peak Width Initial Area Reject Initial Height Reject Initial Shoulders	1.00 0.020 1.000 1.70	 DO Initial O Initial O Initial O Initial Initial

Detector Default Integration Event Table "Event_ECD"

Event	Value Time
Initial Slope Sensitivity	1.000 Initial
Initial Peak Width	0.020 Initial
Initial Area Reject	1.000 Initial
Initial Height Reject	1.700 Initial
Initial Shoulders	OFF Initial

Detector Default Integratio	on Event	t Table	"Event_	_MWD"

Event	Value	Tin	ne
Initial Slope Sensitivity	1	.000	Initial

Initial Peak Width	0.020	Initial
Initial Area Reject	1.000	Initial
Initial Height Reject	1.700	Initial
Initial Shoulders	OFF	Initial

Apply Method's Manual Integration Events: No

Specify Report ======= Calculate: Area Percent Use Multiplier & Dilution Factor with ISTDs

Use Sample Data from Data File Destination: Screen Quantitative Results sorted by: Signal **Report Style:** Short Sample info on each page: No Add Chromatogram Output: Yes Chromatogram Output: Portrait Size in Time direction: 100 % of Page Size in Response direction: 40 % of Page Uncalibrated Peaks: Report with Calibrated Peaks

Signal Options

Include: Axes, Retention Times, Baselines, Tick Marks Font: Arial, Size: 8

Ranges: Full Multi Chromatograms: Separated, Each in full Scale 37

Calibration Table

Calib. Data Modified :

Rel. Reference Window :5.000 %Abs. Reference Window :0.000 minRel. Non-ref. Window :5.000 %Abs. Non-ref. Window :0.000 minUncalibrated Peaks :not reportedPartial Calibration :Yes, identified peaks are recalibratedCorrect All Ret. Times:No, only for identified peaks

Curve Type : Linear Origin : Included Weight : Equal

Recalibration Settings:

Average Response:Average all calibrationsAverage Retention Time:Floating Average New 75%

Calibration Report Options :

Printout of recalibrations within a sequence:
Calibration Table after Recalibration
Normal Report after Recalibration
If the sequence is done with bracketing:
Results of first cycle (ending previous bracket)

Peak Sum Table

No Entries in table

Sample related custom fields			
======= Custom Field	Туре	Mand. Default Value	
None defined			
======== ====== Comj	pound related	custom fields	
Custom Field	• 1	Mand. Default Value	
None defined			

Appendix B: MDA Calibration Curve

